# Immune checkpoint inhibitors in metastatic melanoma therapy (Review)

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Received October 31, 2023; Accepted January 26, 2024

DOI: 10.3892/mi.2024.137

Abstract. An increase in the incidence of melanoma has been observed in recent decades, which poses a significant challenge due to its poor prognosis in the advanced and metastatic stages. Previously, chemotherapy and high doses of interleukin-2 were available treatments for melanoma; however, they offered limited survival benefits and were associated with severe toxicities. The treatment of metastatic melanoma has been transformed by new developments in immunotherapy. Immune checkpoint inhibitors (ICIs), monoclonal antibodies that target cytotoxic T-lymphocyte-associated antigen-4 (CTLA-4), programmed cell death protein 1 (PD-1) and its ligand, PDL-1, have emerged as promising therapeutic options. Commonly used ICIs, such as ipilimumab, nivolumab and pembrolizumab, have been found to be associated with an improved median overall survival, recurrence-free survival and response rates compared to traditional chemotherapies. Combination therapies involving different types of ICIs, such as anti-PD1 with anti-CTLA-4, have further enhanced the overall survival and response rates by targeting various phases of T-cell activation. Additionally, the development of novel biomarkers has facilitated the assessment of responses to ICI therapy, with tissue and serum-based prognostic and predictive biomarkers now available. The increased response observed with ICIs also provides potential for immune-related adverse effects on various organ systems. Further research is required to evaluate the efficacy and safety of various combinations of ICIs, while ongoing clinical trials explore the potential of newer ICIs. Concerns regarding the development of resistance to ICIs also warrant attention. The present review summarizes and discusses the advent of ICIs with a marked significant breakthrough in the treatment of metastatic melanoma, providing improved outcomes compared to traditional therapies.

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## 1. Introduction

The incidence of melanoma has increased over the past two decades (1). Each year, melanoma affects more than 325,000 people. Males experience more frequent occurrences, with 174,000 yearly cases compared to females, with 151,000 cases (2). Of note, 20% of patients with melanoma eventually develop unresectable or distant metastatic disease labeled as stage III/IV (3). Unfortunately, for such an advanced-stage disease, the prognosis remains bleak. However, understanding the advanced stages of melanoma growth and progression has led to the development of promising new therapeutic alternatives.

Until 2011, chemotherapy was the initial treatment for metastatic melanoma; however, it only provided a 6-month median survival time and a 25% 1-year overall survival rate. High-dose interleukin-2 (IL-2) was the only immunotherapy available, but was associated with severe toxicities and only benefited a limited number of patients (4). Currently, advances in immunotherapy

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*Key words:* ipilimumab, nivolumab, pembrolizumab, melanoma, immune checkpoint inhibitors

and studies on cell cycle regulatory molecules have facilitated the creation of immune checkpoint inhibitors (ICIs), a group of monoclonal antibodies that block co-inhibitory molecules, such as cytotoxic T-lymphocyte-associated antigen-4 (CTLA-4), programmed cell death protein 1 (PD-1) and its ligand, PDL1 (5-7). Ipilimumab, nivolumab and pembrolizumab were the first class of medications shown to improve the overall survival of patients with metastatic melanoma (4).

The present review provides comprehensive evidence regarding the role of ICIs and their utilization in advanced melanoma cases. The outcomes of ICIs, such as ipilimumab, nivolumab and pembrolizumab are highlighted, including the improved survival rates and response rates associated with their use compared to traditional chemotherapies, while also focusing on the mechanisms and demonstrating the potentially adverse effects of these therapies.

Furthermore, the present review focuses on combination therapies, including anti-PD1 with anti-CTLA-4, showcasing their importance compared to monotherapy. The improved outcomes of combination therapies over traditional therapies are highlighted, with an emphasis on the need for ongoing research, optimized treatment approaches and strategies which can be used to overcome resistance. In addition to discussing the development of novel biomarkers for assessing ICI therapeutic responses in both tissue and serum-based prognostic and predictive markers, tumor metabolic dependencies and targeting the metabolic pathways by combining ICIs are also discussed. This could provide an improved efficacy, which, to the best of our knowledge, has not been described commonly in the available literature focusing on ICIs used in melanoma.

# 2. Immune checkpoint inhibitor drugs and their role in metastatic melanoma

The surface of immune cells, such as T-cells, assists in the regulation of the immune response through various receptors. When activated by certain ligands, these receptors inhibit immune cells from attacking the body's own cells. However, in cancer, tumor cells can take advantage by binding to these checkpoint-inhibitory receptors through their own ligands and suppressing the immune response, as illustrated in Fig. 1.

Commonly implicated inhibitory receptors include (CTLA-4, PD-1, T-cell immunoglobulin domain and mucin domain-3 (TIM-3), killer cell immunoglobulin-like receptor (KIR), lymphocyte-activation gene 3 (LAG3), glucocorticoid-induced tumor necrosis factor receptor (GITR), B- and T-lymphocyte attenuator (BTLA) and V-domain immunoglobulin (Ig)-containing suppressor of T-cell activation (VISTA) (8-10), as presented in Fig. 2.

Over time, several drugs have been introduced targeting these receptors. Ipilimumab was one of the first ICI drugs to be approved by the Food and Drug Administration (FDA) for the treatment of metastatic melanoma, which functions by blocking CTLA-4 (11). There are numerous additional comparable drugs in early phase III, phase II, or preclinical research. These include pidilizumab, atezolizumab, durvalumab and tremelimumab (formerly known as ticilimumab) (12,13). To increase the immune system's defense against cancer cells, these medications also work against various immunological checkpoints (12,13).

# **3.** Mechanisms underlying the effectiveness of ICIs in metastatic melanoma

*CTLA-4 and anti-CTLA-4 drugs*. CTLA-4, a B7/CD28 family member, is a coinhibitory receptor expressed on the surface of T-cells that eventually inhibits T-cells, and it is expressed by regulatory T-cells (Tregs) (14). Discovered in 1987, it was considered to function as a negative regulator of T-cell activation until the mid-1990s (15-17).

When CTLA-4 is expressed on the surface of CD4<sup>+</sup> and CD8<sup>+</sup> T-cells, the binding affinity increases and is higher to CD80 and CD86, which are the costimulatory receptors present on antigen-presenting cells (APCs) compared to CD28 which is another costimulatory receptor (18). The expression of CTLA-4 increases when there is an activation of T-cell receptors and the release of cytokines such as IL-12 and interferon (IFN)- $\gamma$ . This upregulation creates a feedback inhibition loop on T-effector cells which are activated, leading to CTLA-4 acting as a natural 'brake' on CD4<sup>+</sup> and CD8<sup>+</sup> T-cell activation induced by APCs, as shown in Fig. 3.

Tregs also play a key role in maintaining immune homeostasis by inhibiting excessive immune responses. One of the mechanisms through which Tregs suppress effector T-cell activity is via CTLA-4 signaling (19). Two anti-CTLA-4 drugs have been studied in patients with melanoma: i) Ipilimumab, the first ICI evaluated and approved for the treatment of melanoma is a fully human immunoglobulin anti-CTLA-4 monoclonal antibody (20,21); ii) tremelimumab, a fully human immunoglobulin anti-CTLA-4 monoclonal antibody which is still under investigation (12).

There are two major mechanisms through which these drugs act. First, the inhibition of CTLA-4 signaling in cytotoxic T-cells that specifically target tumors can directly affect these cells by enabling them to evade a state of anergy and enter an active proliferative effector phase. Once activated, these effector T-cells are more likely to penetrate the tumor and exhibit direct cytotoxic effects on tumor cells, while also releasing cytokines such as IL-2 and IFN- $\gamma$  to stimulate an immunogenic tumor microenvironment. Thus, by blocking the CTLA-4 pathway, T-cells that were previously inactive can become activated and effectively target the tumor cells, causing a more powerful immune response against the cancer. This new approach holds promise as a potential immunotherapy for the treatment of cancer (22).

The second major mechanism driving these drugs is the blocking of CTLA-4 signaling in Tregs, which may impair their ability to halt the activity of effector T-cells. This inhibition of CTLA-4 signaling can either cause a decrease in the number of Tregs or reduce their function without affecting their population size. Therefore, blocking CTLA-4 on Tregs may disrupt this suppression and lead to increased immune activation against tumor cells (19,23,24).

Patients with melanoma are treated with the primary aim of suppressing the molecular interplay between the melanoma cells and immune effector cells. Ipilimumab, which has mainly been approved for the treatment of more advanced stages, such as unresectable or metastatic melanoma, has been shown to be associated with a marked overall survival rate confirmed from a phase 3 clinical trial (25). The interference of ipilimumab on CTLA-4 expressed on the subset of tumor-specific T-cell

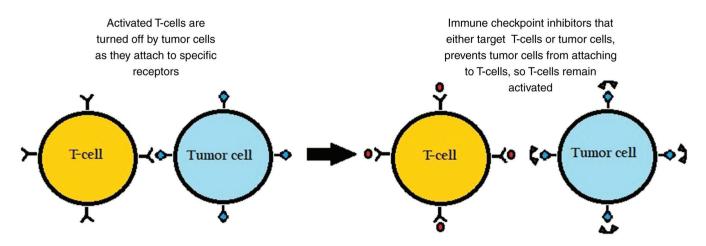


Figure 1. Illustration demonstrating the role of immune checkpoint inhibitors in suppressing the activation of T-cells or preventing the release of cytokines from tumor cells.

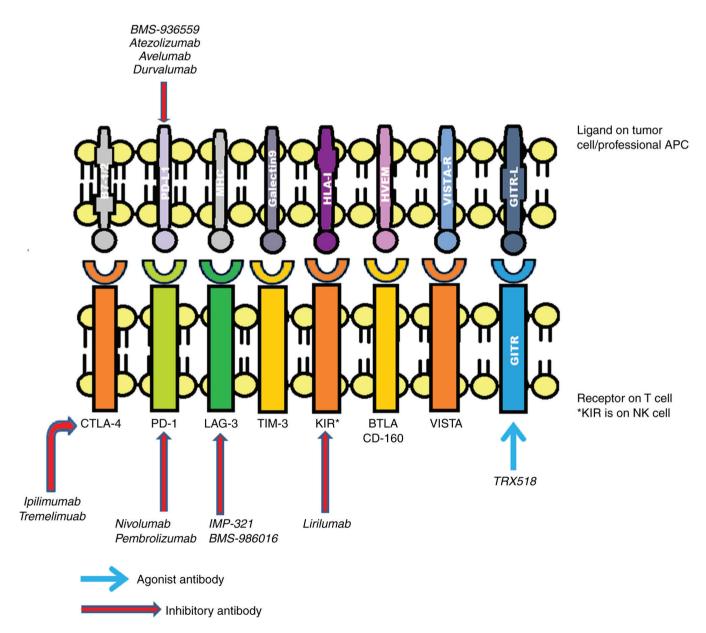


Figure 2. Various types of immune receptors on T-cells and their interaction with ligands present on tumor cells or immune checkpoint inhibitors in the treatment of melanoma. APC, antigen-presenting cell; CTLA-4, cytotoxic T-lymphocyte-associated antigen-4; PD-1, programmed cell death protein 1; LAG-3, lymphocyte-activation gene-3; TIM-3, T-cell immunoglobulin domain and mucin domain-3; BTLA, B- and T-lymphocyte attenuator; KIR, killer cell immunoglobulin-like receptor; VISTA, V-domain immunoglobulin (Ig)-containing suppressor of T-cell activation; NK cell, natural killer cell.

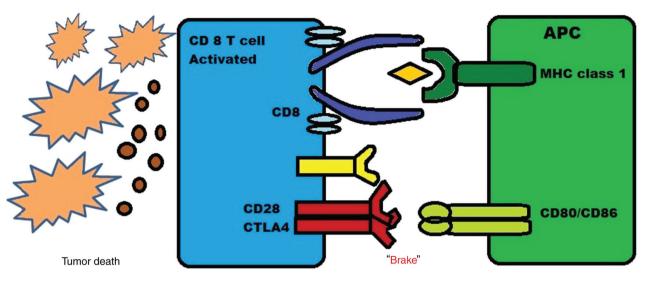


Figure 3. Illustration demonstrating the interaction of CTLA-4 expression on T-cells and CD86 on APCs. CTLA-4, cytotoxic T-lymphocyte-associated antigen-4; APC, antigen-presenting cell; MHC, major histocompatibility complex.

proliferation and B7 molecules on antigen-presenting cells is expected to prevent tumor development (26).

*PD-1 pathway and anti-PD-1/PD-L1 drugs*. PD-1 is a protein present on the surface of T-cells, B-cells and natural killer (NK) cells. It functions as an inhibitory molecule by binding to PD-L1 (or B7-H1) and PD-L2 (B7-H2). PD-L1 is expressed in numerous types of tissue, including hematopoietic cells and certain tumors such as melanoma, where they are expressed in 40-50% of cases. PD-L2 is mainly expressed in hematopoietic cells. The binding of PD-1 to PD-L1/2 inhibits the death of tumor cells and promotes the conversion of T-effector cells into Tregs, while also inducing exhaustion in peripheral T-effector cells, as illustrated in Fig. 4 (27,28).

PD-1 and/or PD-L1 are also expressed on cells, such as NK cells, monocytes and dendritic cells (27,28). The PD-1 pathway operates through various mechanisms, such as reducing the activity of T-cells during an inflammatory response, increasing the proliferation and suppressive activity of Tregs, and reducing the lytic activity of B-cells and NK cells (29).

The affinity between PD-1 and PD-L1 is 3-fold stronger than the affinity between PD-1 and PD-L2. When PD-L1 binds with PD-1 on T-cells, it results in T-cell exhaustion, dysfunction, neutralization and the production of IL-10 within the tumor mass. This process allows tumors that overexpress PD-L1 to protect themselves from being attacked and killed by CD8<sup>+</sup> cytotoxic T-cells (30).

Pro-effector cytokines, namely IL-12 and IFN-γ, can upregulate the expression of PD-1 and PD-L1/L2, which helps to prevent excessive T-effector cell activity. It is worth noting that PD-L1 has also been shown to inhibit CD80, indicating the existence of complex interactions between CTLA-4, PD-1 and other pathways (31,32).

The PD-1 and PD-L1 antibody inhibitors were created with the aim of preventing the PD-1 or PD-L1 side from functioning, reactivating T-cells and promoting an immune response against cancer cells (13).

Based on promising results from clinical trials, antibodies that inhibit PD-1 (such as pembrolizumab, nivolumab-IgG4

fully humanized and dostarlimab), as well as those that inhibit PD-L1 (such as avelumab, atezolizumab, and durvalumab) are being evaluated for use in melanoma cases and various other malignancies (NCT04020809, NCT04274816, NCT03313206, NCT03842943 and NCT05928962). However, it is not yet known which inhibitor, PD-1 or PD-L1, is more efficient (13). Of these, nivolumab and pembrolizumab are the two major FDA-approved anti-PD-1 monoclonal antibodies available for the treatment of advanced and metastatic melanomas.

Melanoma cells exhibit increased levels of PD-L1, which promotes the apoptosis of the likewise increased levels of T-cells (33). It has also been found that the circulating melanoma antigen-specific T-cells and tumor-infiltrating lymphocytes express PD-1 abnormally. It is considered that melanoma cells are capable of initiating, as well as sustaining PD-1 signals, T-cell fatigue and dysfunction (33). Hence, by blocking PD-1 in patients with melanoma, one could possibly restore abnormal activation and signaling and eventually recover the immune effect. Pembrolizumab or lambrolizumab were used in unresectable or metastatic melanoma in the study by Hamid *et al* (34), in an aim to elucidate the effects of PD-1 medications in melanoma.

However, due to the heterogeneous nature of tumors, the expression of PD-L1 is not uniform throughout. The extent of PD-L1 expression can differ in various locations within the tumor, resulting in varying levels of PD-L1 in immunohistochemical staining. Moreover, the effectiveness of PD-L1/PD-1 inhibitors can also be influenced by several other factors, such as the type of cancer, the patient's immune system and the genetic profile of the tumor. Thus, a more in-depth understanding of these factors is essential for developing effective treatment strategies that consider the heterogeneity of tumors and the variability in PD-L1 expression (35).

The combination ICI therapy used in the treatment of patients with metastatic melanoma primarily involves CTLA-1 and PD-1 inhibitors. This amplified the inhibitions that can be simultaneously intervened during different phases of the interaction among melanoma cells and the immune system. This, for example, includes anti-CTLA-4 inhibiting

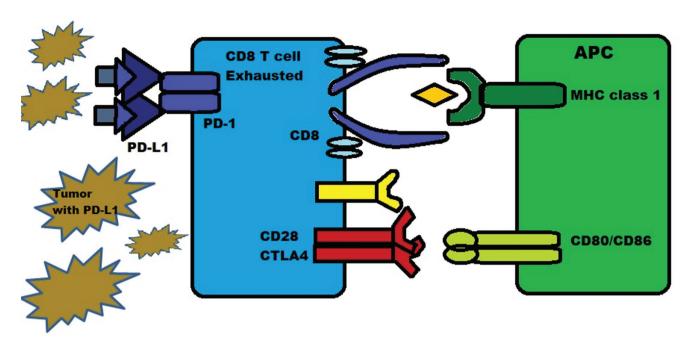


Figure 4. Illustration demonstrating the interaction between PD-1 and PD-L1 among T-cells and tumor cells. PD-1, programmed cell death protein 1; PD-L1, programmed cell death protein ligand 1; CTLA-4, cytotoxic T-lymphocyte-associated antigen-4; MHC, major histocompatibility complex.

the priming stage at the same time anti-PD-1 inhibits the effector stage (36,37). It has also been noted that the use of anti-CTLA-4 inhibitors results in an increased expression of PD-1; hence, using combination therapy results in a more robust treatment response in patients with melanoma (38).

Newer drugs. An increased understanding of immunological mechanisms has led to the identification of additional potential targets for checkpoint inhibition in the treatment of cancer. Some of these potential targets include BTLA, VISTA, TIM-3, CD47 and LAG-3. i) The blockade of BTLA has been shown to enhance New York esophageal squamous cell carcinoma 1) specific CD8<sup>+</sup> T-cell function and enhance the efficacy of anti-PD-1 (39-41). ii) VISTA blockade has been shown to increase T-cell infiltration and function in tumors, thereby reducing tumor growth (10,42). iii) TIM-3 blockade causes T-helper-1 cell hyperproliferation and cytokine release, leading to tumor shrinkage in a mouse model when combined with anti-CTLA-4 or anti-PD-1 (43-47). iv) Targeting CD47 with a humanized anti-CD47 monoclonal antibody in combination with rituximab has shown to lead to objective responses in half of the heavily pretreated patients with relapsed or refractory non-Hodgkin's lymphoma, including a complete response in more than one-third of patients (48). v) An immune pathway known as LAG-3 has been identified as a potential complement to the PD-1/PDL1 pathway in enhancing the immune response against cancer. LAG-3 is an immune checkpoint receptor that regulates the function of T-cells. BMS-986016 is a therapy that targets LAG-3 and is currently under investigation in combination with nivolumab, which targets PD-1, to enhance the immune response against cancer cells. The combination of these two therapies has the potential to create a synergistic effect, leading to improved treatment outcomes for patients with cancer (49).

# 4. Combination therapy with immune checkpoint inhibitors in melanoma

The ability of anti-CTLA-4 and anti-PD-1/PD-L1 monoclonal antibodies to target various T-cell activation locations and phases is the rationale for their combined use. PD-1 is primarily expressed on antigen-experienced T-cells in peripheral tissues, while CTLA-4 is expressed by naive T-cells in the lymph nodes. According to pre-clinical research, combining ICIs is more effective than treatment with with monotherapy for managing melanoma (50-53).

In pre-clinical investigations, anti-CTLA-4 and anti-PD-1/PD-L1 monoclonal antibodies have been shown to induce the infiltration of CD8<sup>+</sup> T-cells and the expansion of an inducible T-cell co-stimulator (ICOS)<sup>+</sup> T helper 1-like CD4 fraction, which in turn induces the response of CD4<sup>+</sup> effector T-cells. Based on this, the sequencing or combination of nivolumab with ipilimumab in metastatic cutaneous melanoma has been researched (50-53). Other combination studies are on nivolumab, relatlimab and combination therapy with pembrolizumab with low-dose ipilimumab (54,55). The data from the CheckMate and RELATIVITIY047 trials on the combination of ICIs are presented in Table I, which demonstrate a favorable response for such therapies (54,56,57). Table II presents data from a meta-analysis, comparing monotherapy and combination therapy (47-64).

### 5. Comparison between the immune checkpoint inhibitors

Along with the major breakthrough in melanoma treatment with the use of selective BRAF inhibitors, after ~6 months of the median duration, resistance to therapy began to develop. The BRAF mutation drives the tumor proliferation exponentially by activating mitogen-activated kinase pathway (MAP),

Study name	Treatment group	Objective response rate (%)	Median PFS	5-year PFS rate	Median overall survival	5-Year overall survival rate	(Refs.)
CheckMate 064	Nivolumab followed by ipilimumab	41	-	-	76%	-	(56)
CheckMate 064	Ipilimumab followed by nivolumab	20	-	-	54%	-	(56)
Checkmate 067	Nivolumab-ipilimumab	58	11.5 months	36%	36.9 months	52%	(57)
Checkmate 067	Nivolumab	45	6.9 months	29%	-	44%	(57)
Checkmate 067	Ipilimumab	19	2.9 months	8%	19.9 months	26%	(57)
<b>RELATIVITY-047</b>	Nivolumab-relatlimab	47.7	10.1 months	-	-	-	(54)
<b>RELATIVITY-047</b>	Nivolumab	36	4.6 months	-	_	-	(54)

Table I. Summary of the results of the CheckMate and RELATIVITY047 trials performed on the combination of immune checkpoint inhibitors for the treatment of melanoma.

Table II. Summary of results obtained from a previous meta-analysis of clinical trials conducted by Pradeep *et al* (58), which compared the efficacy and safety of ICIs between monotherapy and combined ICI therapy in advanced melanomas.

Outcome [(Refs.); as in the meta-analysis by Pradeep <i>et al</i> (58)]	Comparison	Effect size in terms of HR or RR (95% CI)	P-value
Overall survival (52-56)	Nivolumab with ipilimumab vs. monotherapy	HR, 0.65 (0.53-0.79)	<0.0001
	Nivolumab with ipilimumab vs. nivolumab alone	HR, 0.84 (0.71-0.99)	0.04
	Nivolumab with ipilimumab vs. ipilimumab alone	HR, 0.54 (0.48-0.62)	< 0.00001
Progression-free survival (53,55-58)	Nivolumab with ipilimumab vs. monotherapy	HR, 0.48 (0.38-0.60)	< 0.0001
	Nivolumab with ipilimumab vs. nivolumab alone	HR, 0.68 (0.49-0.94)	0.02
	Nivolumab + ipilimumab vs. ipilimumab alone	HR, 0.42 (0.37-0.47)	< 0.00001
Overall response rate (52,54-57,59)	Nivolumab with ipilimumab vs. monotherapy	RR, 2.15 (1.63-2.84)	< 0.00001
L	Nivolumab with ipilimumab vs. nivolumab alone	RR, 1.32 (1.22-1.43)	< 0.00001
	Nivolumab with ipilimumab vs. ipilimumab alone	RR, 3.09 (2.74-3.50)	< 0.00001

In the meta-analysis by Pradeep *et al* (58), overall survival, progression-free survival and objective response rates were the outcomes studied. HR, hazard ratio; RR, relative risk.

and the development of resistance to BRAF inhibitors in both MAP kinase-dependent and MAP kinase-independent pathways (65-67). Resistance in MAP kinase-dependent pathways includes secondary mutations in NRAS, the increased expression of COT kinase, CRAF activation and acquired mutations in MEK1 (65,68-71). MAP kinase-independent pathways include the upregulation of platelet-derived growth factor receptor, additional receptor tyrosine kinases activation including AXL, Erb-B2 receptor tyrosine kinase 4) and insulin like growth factor 1 receptor, the activation of PI3K/AKT signaling, and the loss of phosphatase and tensin homolog (PTEN) (65,69,71-75).

*Ipilimumab*. Ipilimumab is a fully human monoclonal antibody developed to antagonize CTLA-4. A clinical study was conducted on patients who had unresectable stage III or IV melanoma, where they were randomly assigned in a 3:1:1 ratio to receive ipilimumab (3 mg/kg) plus the glycoprotein 100 (gp100) vaccine, ipilimumab alone, or gp100 alone. The patients who received ipilimumab plus gp100 had a longer median overall survival rate of 10 months compared to 6.4 months for those who received gp100 alone, with a hazard ratio (HR) for mortality of 0.68 and a statistically significant P-value of <0.001. The median overall survival rate of patients who received ipilimumab alone was 10.1 months (HR, 0.66; P=0.003) compared to those who received gp100 alone (20).

In the study by Robert *et al* (25), patients with untreated metastatic melanoma were randomly assigned to receive either ipilimumab plus dacarbazine or dacarbazine alone. A significantly longer median overall survival (11.2 months) was found in the ipilimumab plus dacarbazine group compared to the dacarbazine alone group (9.1 months). In a follow-up maintenance study, patients who received ipilimumab plus dacarbazine alone (8.8%). These findings suggest that ipilimumab in combination with dacarbazine may be an effective treatment option for metastatic melanoma patients (76).

In a trial for ipilimumab as an adjuvant treatment performed by Eggermont *et al* (77), patients with stage III cutaneous melanoma who had undergone complete resection were randomly administered either ipilimumab (10 mg/kg) (n=475) or a placebo (n=476). Following a median follow-up of 5.3 years, the ipilimumab group had a 5-year recurrence-free survival rate of 40.8%, while the placebo group had a rate of 30.3% (HR, 0.76; P<0.001). In terms of the 5-year overall survival rate, the ipilimumab group had a rate of 65.4% compared to 54.4% in the placebo group (HR, 0.72; P=0.001) (77).

In the study by Ascierto *et al* (78), patients with unresectable stage III or IV melanoma were randomly assigned to receive either 10 or 3 mg/kg ipilimumab. The median overall survival rate was 15.7 months for the 10 mg/kg group and 11.5 months for the 3 mg/kg group (HR,0.84; P=0.04). Overall, their study suggested that ipilimumab treatment improved the survival outcomes of patients with unresectable stage III or IV melanomas, with higher doses of the drug (10 mg/kg) leading to an improved overall survvial compared to lower doses (3 mg/kg) (78).

Nivolumab. Nivolumab is a monoclonal antibody that inhibits the interaction of PD-1 with PD-L1. The clinical study performed by Robert et al (79) compared the efficacy of nivolumab with the standard therapy of dacarbazine. In their study, patients who had metastatic melanoma without a BRAF mutation were randomly divided into two groups (1:1 with nivolumab at 3 mg/kg once every 2 weeks (n=210) and dacarbazine (n=208). The survival rate at 1 year was 72.9% in patients treated with nivolumab compared to 42.1% in patients who were assigned dacarbazine (HR, 0.42, P<0.001). The objective response rate was 40% with nivolumab compared to 13.9% with dacarbazine (odds ratio, 4.06; P<0.001) (79). Another randomized controlled trial was carried out between 2012-2014 on patients with advanced melanoma who progressed after ipilimumab therapy or a combination of ipilimumab and a BRAF inhibitor if they were found to be positive for a V600E mutation (80). That study assessed the role of nivolumab as a second-line treatment in the management of patients with advanced melanoma. Patients were divided into three groups in a 2:1 pattern where one group (n=272) received nivolumab at 3 mg/kg once every 2 weeks and another group (n=133) received the investigator's choice of chemotherapy (ICC), which was either dacarbazine or paclitaxel plus carboplatin (80). An interim analysis of that study found that, in the first 120 patients of the nivolumab group, 38 patients (31.7%) experienced confirmed objective responses, whereas only 5 out of the 47 patients (10.8%) receiving the ICC treatment exhibited similar responses. Subsequently, upon further analysis of that trial, it was revealed that the median overall survival rate of patients who received nivolumab was 16 months, while for those who received ICC, it was 14 months (81). The HR was 0.95, indicating that nivolumab did not improve the survival rate of patients who had ipilimumab-refractory metastatic melanoma when compared to ICC. However, nivolumab had a higher overall response rate of 27% vs. 10% for ICC, and the median duration of response was also longer for nivolumab at 32 months compared to 13 months for ICC (77). Hence, nivolumab exhibiting a higher overall response rate and a longer duration of response suggests that it may be a more effective treatment option for some patients (81). Another study was also carried out to compare the efficacy of nivolumab compared to ipilimumab as an adjuvant therapy in patients who had resected advanced melanoma. In patients with stage III or stage IV melanoma, adjuvant therapy was administered with either nivolumab (n=453) or ipilimumab (n=453) and follow-up was performed after 18 months (82). The 12-month rate of recurrence-free survival was significantly higher in the nivolumab group at 70.5%, vs. 60.8% in the ipilimumab group with a HR of 0.65 (P<0.001). It was also noted that treatment-related adverse events were 14.4% for patients treated with nivolumab and 45.9% for those treated with ipilimumab. Therefore, patients who received ipilimumab therapy experienced more severe side-effects than those who received nivolumab therapy. This suggests that nivolumab may be a more effective and tolerable treatment option for patients with stage IIIB, IIIC, or IV melanoma following surgical resection (82).

Pembrolizumab. Pembrolizumab is a monoclonal antibody which functions by blocking the PD-1 on T-cells and allowing these T-cells to identify and kill cancer cells. Similar to nivolumab, pembrolizumab was also compared with ICC in ipilimumab-refractory melanoma. A randomized controlled study was conducted on patients with advanced melanoma and have progressed even after receiving ipilimumab and/or standard BRAF therapy (83). Patients were divided into three groups as follows: One group (n=181) received 10 mg/kg pembrolizumab, one group (n=180) received 2 mg/kg pembrolizumab, and another group (n=179) received ICC. The 6-month progression-free survival rate was found to be 38% in patients treated with pembrolizumab at 10 mg/kg (HR, 0.5 vs. ICC; P<0.0001), 34% in the 2 mg/kg group (HR, 0.57 vs. ICC; P<0.0001) and 16% in the ICC group (83). Another study was conducted by Robert et al (84), this time comparing pembrolizumab with ipilimumab. Patients with advanced melanoma were divided at a 1:1:1 ratio to receive pembrolizumab at 10 mg/kg once every 2 weeks or pembrolizumab at 2 mg/kg once every 3 weeks or four doses of ipilimumab at 3 mg/kg for once every 3 weeks. An interim analysis was performed which revealed that the 6-month progression-free survival of the patients treated with pembrolizumab once every 2 weeks was 47.3% (HR, 0.58 vs. ipilimumab; P<0.001), 46.4% for those treated with pembrolizumab once every 3 weeks (HR, 0.58 vs. ipilimumab; P<0.001) and 26.5% for those treated with ipilimumab (84). A final analysis revealed that the median overall survival rate was not reached in both pembrolizumab groups; however, it was noted to be 16 months in the ipilimumab group (HR, 0.68 for pembrolizumab once every 2 weeks vs. ipilimumab, P=0.0009; and HR, 0.68 for pembrolizumab once every 3 weeks vs. ipilimumab, P=0.0008) (85). Similarly, in the study by Robert et al (84) the 24-month overall survival rate was 55% in the group treated once every 2 weeks, 55% in the group treated once every 3 weeks and 43% in the ipilimumab group. Not only do nivolumab and pembrolizumab prolong overall survival, but they also maintain the quality of life of patients with melanoma (86,87). These findings have led to the FDA approval of pembrolizumab for ipilimumab and/or BRAF inhibitory refractory advanced melanoma. A summary of the comparison among ICIs and their outcomes in patients is presented in Table III.

ICI drug	Compared drug	Outcome	(Refs.)
Ipilimumab	Ipilimumab (3 mg/kg) (n=137) vs. Gp100 (n=136) vs. ipilimumab (3 mg/kg) with gp100 (n=403)	Median overall survival (months): 10.1 vs. 6.4 vs. 10.0	(20)
	Ipilimumab (10 mg/kg) with dacarbazine (n=251) vs. dacarbazine (n=251)	Median overall survival (months): 11.2 vs. 9.1	(76)
	Ipilimumab (10 mg/kg) adjuvant (n=475) vs. placebo (n=476)	Recurrence-free survival (%): 40.8 vs. 30.3	(77)
	Ipilimumab (10 mg/kg) (n=365) vs. ipilimumab (3 mg/kg) (n=362)	Median overall survival (months): 15.7 vs. 11.5	(78)
Nivolumab	Nivolumab(3 mg/kg) once every 2 weeks (n=210) vs. dacarbazine (n=208)	1-Year survival rate (%): 72.9 vs. 42.1; objective response rate (%): 40 vs. 13.9	(79)
	Nivolumab (3 mg/kg) (n=272) vs. ICC (dacarbazine or paclitaxel plus carboplatin) (n=133)	Median overall survival rate (months): 16 vs. 14; overall response rate (%): 27 vs. 10	(80)
	Nivolumab (n=453) vs. ipilimumab (n=453)	12-Month recurrence-free survival rate (%): 70.5 vs. 60.8; treatment-related adverse events (%): 14.4 vs. 45.9	(82)
Pembrolizumab	Pembrolizumab (10 mg/kg) (n=181) vs. pembrolizumab (2 mg/kg) (n=180) vs. ICC (paclitaxel plus carboplatin, paclitaxel, carboplatin, dacarbazine, or oral temozolomide) (n=179)	<ul><li>6-Month progression-free survival rate (%):</li><li>38 vs. 34 vs. 16</li></ul>	(83)
	Pembrolizumab (10 mg/kg once every 2 weeks) (n=279) vs. pembrolizumab (2 mg/kg once every 3 weeks) (n=277) vs. four doses of ipilimumab (3 mg/kg once every 3 weeks) (n=278)	6-Month progression-free survival rate (%): 47.3 vs. 46.4 vs. 26.5; median overall survival rate (months): NA vs. NA vs. 16; 24-month overall survival rate (%): 55 vs. 55 vs. 43	(85)

Table III. Individual immune checkpoint inhibitors with the various outcomes affecting clinical decisions regarding their use.

ICC, investigator's choice of chemotherapy; NA, not available.

### 6. Biomarkers

The prognostic marker for melanoma traditionally used is the depth of invasion and the associated mitotic count of the affected cells. With advancements being made, newer prognostic markers have been found and used. Prognostic and predictive biomarkers have gained importance, particularly in the treatment of melanoma.

*Serum biomarkers*. The role of serum biomarkers in the early detection of melanoma is described in Table IV (88-106).

#### Tissue markers

*Prognostic markers*. i) Tumor infiltrating lymphocyte (TIL) patterns are often divided into grades, such as 'absent', which is no presence of any lymphocytes within the tumor, 'non-brisk', which suggest few foci of lymphocytes within the tumor, or 'brisk', which is a large diffuse infiltration of lymphocytes within the tumor (107). As demonstrated by Clark *et al* (107) in 1989, as well as by others, the presence of brisk TILs in a vertical growth pattern is often associated with a favorable disease-specific survival and overall survival rate after non-brisk and absent patterns of TILs (107,108).

ii) Histotype: The majority of melanoma histotypes are not considered prognostic when looked at individually from tumor thickness, and are therefore not included in the American Joint Committee on Cancer staging system (90,109,110). However, a nodular melanoma is an independent predictor which can be used for the measurement of recurrence and its association with mortality due to melanoma (111).

iii) Digital images trained from AI: New advancements have allowed for the development of deep learning-based biomarkers, which can help to stratify the stages of melanoma into risk groups, and thus associate disease-specific survival with two independent validating cohorts to accurately predict the prognosis of patients with early-stage melanoma (112).

iv) Melanoma cell adhesion molecule (MCAM): Expressed in 80% of metastatic tumors, MCAM is a cell adhesion marker (113). Those who are positive for MCAM have significantly worse 5-year survival rates than those who are negative for MCAM, and there is an inverse association between the amount of marker expressed and survival (114,115).

v) Ki-67: Ki-67 is a unique nuclear antigen that can function as a marker for cellular proliferation during the active phase of the cell cycle (116). For melanomas who have a thickness <1 mm, the risk of metastasis increases with the expression of

Table IV. Role o	Table IV. Role of serum biomarkers in melanoma.		
Biomarker	Use	Type	Information (Refs.)
LDH	Prognostic and predictive	Protein-based	The AJCC staging scheme includes blood LDH as an independent prognostic biomarker for metastatic melanoma. A univariate analysis revealed that an increased amount of LDH was associated with a lesser response to anti-PD-1 medications in patients who received anti-PD-1 monotherapy (88). It was previously found that patients with elevated LDH and NLR levels, despite receiving ICIs, had poor a OS and PFS (89). Elevated serum LDH levels increased the disease stage of patients with metastasis to stage IV M1c (1) metastatic disease, which included individuals with metastases to the lungs, as well as to other visceral sites anart from the central nervous system (90.91).
S100	Prognostic	Protein-based	S100B levels in serum are affected by the tumor load, and higher values of S100B may be used as a prognostic biomarker and a sign of disease progression (92-94). It was previously found that the discriminative ability with serum S100B levels for detecting disease relapse was much higher than that of serum LDH (95).
miRNA	Prognostic	Genomic	The role of miRNAs as post-transcriptional inhibitors of translation has been substantially investigated (96-100). It has been discovered that these small non-coding RNAs are detectable in exosomes in serum (101). It has been demonstrated that aberrant levels of expression of certain miRNAs are associated with the stage and recurrence of disease (102,103). miRNAs are crucial in the deregulation of a number of oncogenic pathways in uveal melanoma, which can facilitate the metastatic spread of disease. The study by Wróblewska <i>et al</i> (104) found that it may be beneficial to use differentially expressed miRNAs as a biomarker for the evaluation of the risk of metastasis in patients with uveal melanoma.
ctDNA	Prognostic	Genetic	It has been demonstrated that circulating tumor DNA testing may detect BRAF V600e mutations, which is helpful for determining how effectively the BRAF/MEK inhibitor therapy works. Imaging scan measurements of disease progression and treatment effectiveness have been found to be associated with the amount of circulating tumor DNA (105,106). Both univariate and multivariate analyses have revealed an association between ctDNA detectability and a shorter OS. Patients with distant metastases (79%) were more likely to have ctDNA detected than those with no distant metastases or only intracranial tumors (32%). Elevated protein S100 and CRP levels were more closely linked to detectable ctDNA than LDH (104).
AJCC, American survival; PFS, pro	AJCC, American Joint Committee on Cancer; LDH, lactate dehydrogenase; PD-1, programme survival; PFS, progression-free survival; miRNA, microRNA; ctDNA, circulating tumor DNA.	tate dehydrogenase; PD-1 RNA; ctDNA, circulating t	AJCC, American Joint Committee on Cancer; LDH, lactate dehydrogenase; PD-1, programmed cell death protein 1; NLR, neutrophil-lymphocyte ratio; ICI, immune checkpoint inhibitor; OS, overall survival; PFS, progression-free survival; miRNA, microRNA; ctDNA, circulating tumor DNA.

Ki-67 and an increased mitotic rate (117). However, with the increasing thickness of melanomas, Ki-67 can serve as a more effective prognostic marker than the mitotic rate, and is often associated with ulceration within the tumor, necrosis, higher level Clark's level of invasion, and even vascular invasion (118). In addition, with recurrent melanomas, higher values of Ki-67 exhibit an independent association with a decreased overall survival (119).

vi) Lymphatic invasion: In research on primary melanomas with a thickness >1 mm, D2-40 staining was assessed for lymphatic invasion, which is an antibody against sialoglycoprotein that selectively attaches on endothelial cells of lymphatic vessels and helps detect sentinel lymph node metastasis (120-122).

vii) Osteopontin: Overexpressed in numerous visceral malignancies, osteopontin is known as an integrin-binding protein and used as a biomarker to measure tumor progress and metastasis (123-125). It functions as an independent predictor for the prognosis of melanoma and was found to be associated with increased sentinel lymph node positivity in a cohort of 345 patients who had primary melanoma detected using immunohistochemical analysis (126).

viii) Driver mutations: It has been found that BRAF and NRAS are associated with a significantly lower melanoma-specific survival in high-risk tumors, such as a stage >2 (127). NF1 mutations has also been found to be associated with a lower disease-specific survival and overall survival (128). However, further research is required to identify patients with BRAF mutations and uncover the role of BRAF mutations in directing the treatment strategy.

*Predictive markers.* The role of predictive markers in melanoma and its clinical importance is summarized in Table V (61,129-151).

### 7. Factors affecting drug use

Therapy with nivolumab affects the frequencies of innate lymphoid cells (ILCs) in peripheral blood in patients with melanoma. The frequency, as well as the secretory activity of ILC subsets, particularly ILC2s, are affected by treatment. Albeit nivolumab was found to not effectively alter serum cytokine profiles, pro-inflammatory and angiogenic substances such as IL-1, IL-6, CCL2, CXCL8 and VEGF had levels outside the normal range in 7 of the 18 cytokines. In addition, the production of IL-5 and IL-13 was affected, which are released during parasite infections and allergic reactions (152). In malignant melanoma, type 3 ILC is suspected in tumor suppression (153). Serum levels of IL-6, CXCL8 and CCL2 in particular, surge during melanoma progression, while mature NKp44<sup>+</sup> ILC3s protect against melanoma (154).

As previously demonstrated, the advancement of melanoma was comparable with aging, although the treatment outcome did not differ significantly, and there was no significant change in the survival outcomes of elderly patients as compared to young ones. Moreover, it was recommended that both age groups should be treated in similar manner (155). Primary and secondary resistance are also a key factor affecting drug use (84,156). Combination therapy with ipilimumab and nivolumab, as approved by the FDA, has been proven to be efficient (157). There is an increased incidence of melanoma among women of reproductive age. As opposed to this, postmenopausal women have a relatively low incidence of the disease, thus raising the possibility that sex hormones such as estrogen may be involved in the growth of the disease (158). As a result, estrogen levels should be considered an important biomarker in advanced melanoma. Elderly patients aged  $\geq 65$ treated with combination therapy comprising of ipilimumab and nivolumab have not exhibited a considerable difference in overall mortality. When prior exposure to ipilimumab is considered, women have a 2.82-fold increased risk of mortality as compared to prior-exposed males with ipilimumab (158).

Moderate colitis which does not require the use of intravenous steroids is consistent with an improved overall survival of patients with stage IV melanoma when treated with a single anti-CTLA-4 drug, but not with combination drugs. This holds true even after the completion of therapy (159). Multiple nonrandomized studies have shown excellent results in patients who discontinue treatment after being treated for 1-2 years and disease progression is also uncommon in the following 2-5 years of treatment termination (160-162). This is in contrast to the progression of disease of patients with non-small cell lung cancer, for whom treatment continuation led to improved results compared to treatment termination (163).

### 8. Adverse effects

The use of PD-1 inhibitors, namely nivolumab and pembrolizumab, and the anti-CTLA-4 drug, ipilimumab, has been shown to be associated with a steady regression in malignancies, including metastatic melanoma (164).

PD-1 inhibitors function in the tumor setting, while CTLA-4 inhibitors act on lymphoid tissue, resulting in a wide and different set of adverse events (165). Combination therapies with nivolumab and ipilimumab have been proven to be more effective with a response rate of 59% as compared to when used alone, with response rate of 43% for nivolumab and 15-20% for ipilimumab. Moreover, an increased response rate is associated with an increase in adverse events, resulting in an overall increase in adverse events with the combination of nivolumab with ipilimumab, as compared to nivolumab or ipilimumab monotherapy (36,62).

A CTLA-4 blockade with or without anti-PD-1 antibody produces adverse events in a dose-dependent manner (166,167). Considering that older patients are more inclined to develop rheumatologic events and female patients are also at an increased risk, the toxicity profile may vary according to age and sex (168,169).

However, as these molecules are targeted, due to the resulting immune response, an increase in the incidence of autoimmune conditions is observed; these adverse events are known as immune-related adverse events (irAE). If severe irAEs occur with one of the drugs, then it is a safe practice to re-challenge the patient with a different class of drug (165). These drugs have the following on the following systems.

*Gastrointestinal tract*. Diarrhea is the most frequent irAE with incidences between 10 and 13% (164).

*Endocrine disorders*. In decreasing order, the first endocrine system that is most affected by ICIs is the thyroid gland

Table V. Role of predictive biomarkers and	and their clinical relevance.
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Туре	Biomarker	Clinical relevance (Refs.)
Tumor intrinsic	TMB/neoantigen profile	Tumors with a higher TMB are potentially more responsive to ICIs; a reason for this may be the fact that these tumors express more neoantigen and hence, may be recognized easily as a target by T-cells (129). With the increasing evidence of the ability of TMBs to predict the response to immunotherapy in melanoma, various studies have been conducted (130,131). In a previous study, 321 patients with melanoma were treated with ICIs and it was found that with a higher TMB, one could predict an increased survival following treatment (132).
	Driver mutations	Immunotherapy was found to be more effective in patients with activating NRAS genes than in those without such mutations. According to the CheckMate 067 study, those patients with BRAF-mutation in melanoma had a 4-year overall survival rate of 62% with combination therapy with ipilimumab and nivolumab, when compared to only 50% with nivolumab alone and 33% with ipilimumab alone (61,133).
Tumor microenvironment	МНС	Longer overall survival, a greater response rate, and an increased infiltration of CD4 <sup>+</sup> and CD8 <sup>+</sup> T-cells in the tumor microenvironment were all associated with elevated levels of MHC-II expression. Conversely, ipilimumab treatment was found to be associated with a higher risk of disease progression when MHC-I expression was lower (<30%), with a 100% negative predictive value (134-136).
	HLA supertypes	Two HLA supertypes were identified and linked to CTLA-4 inhibition therapeutic outcomes: HLA-B44 was associated with a longer survival, while HLA-B62 was associated with a shorter survival (137).
	PD-L1	PD-L1 is a complex biomarker to study since it is regulated by several pathways, which is vulnerable to high sampling error, and expressed on numerous immune cells in the microenvironment (138). Treatment with PD-1 inhibitors has been demonstrated to be successful in both patients with and without PD-L expression; however, the quality of immunohistochemical staining for PD-L1 is not dependable for clinical use (139,141).
	Immune metabolism	ICI-sensitive melanomas exhibited more oxidative phosphorylation and lipid metabolism than ICI-resistant tumors using high resolution mass spectrometry. In addition, elevated lipid metabolism increases antigen presentation, which may be the reason why ICI-sensitive cancers have a better response to therapy (142).
	TIL/TCF-7	It has been demonstrated that TCF-7, a transcription factor, promotes a central memory stem-like state, and that TCF7-expressing CD8 <sup>+</sup> cells have the ability to self-renew and differentiate into effector cells (143,144). TCF-7-positive cells appear to be the ones that multiply following anti-PD1 therapy (145). According to this finding, TCF-7 expression has been linked to successful clinical results in patients with melanoma receiving ICIs (146). In light of this, further research is required to establish the role of TCF7 expression as a predictive biomarker in patients with melanoma who are being treated with ICIs and its role as a therapeutic target.
	GEP	Research has been performed to identify a gene expression profile (GEP) that can predict the response to pembrolizumab. It was discovered that the 18-gene profile could identify aspects of the tumor microenvironment that are pertinent to predicting the clinical outcome of pembrolizumab that are independent of tumor type. This indicates that GEPs, such as the 18-gene profile, can be utilized to predict how different types of cancer will respond to PD-1/PD-L1 inhibitors (147-149).
	IPRES	The IPRES transcriptional signature is a peculiar pattern of gene expression that is present in tumors which are resistant to anti-PD-1 medications. Increased activity in genes is related to an increase in transition through the mesenchyme, helping in cell adhesion, extracellular matrix remodeling, increase in angiogenesis, and wound repair which are the hallmark of this pattern. The response rate to anti-PD-1 therapy can be enhanced with the identification of transcriptomic features that are associated with anti-PD-1 resistance, which may suggest the mitigation of IPRES-related biological process and eventually, an enhance response rate to treatment (147)

Туре	Biomarker	Clinical relevance (Refs.)	
Host factors	Gut microbiome	Patients with high-fiber diets were 5-fold more likely than those with low-fiber diets to respond to anti-PD-1 therapy. In addition, regardless of the type of antibiotic, indi viduals who had received antibiotics for >30 days prior to commencing ICI treatment suffered worse outcomes. A past but not concurrent antibiotic treatment has an impact on the gut microbiome's 'activating' response to ICIs (150).	
	Stress	A solid tumor line was grafted into mice, and they were then treated with an anti-PD- monoclonal antibody. Subsequently, they were made to experience social defeat stress and compared to the mice who did not experience this type of stress; the mice who were socially defeated with stress responded to PD-1 inhibition less effectively (151).	
Others	AI	According to recent research, pre-treatment computerized H&E slides can be used to artificially predict the probability of a response to ICIs (112).	

TMB, tumor mutational burden; ICI, immune checkpoint inhibitor; MHC, major histocompatibility complex; HLA, human leukocyte antigen; CTLA-4, cytotoxic T-lymphocyte-associated antigen-4; PD-L1, programmed cell death protein ligand 1; TCF-7, transcription factor 7; IPRES, innate anti-PD1 resistance; AI, artificial intelligence; H&E, hematoxylin and eosin.

(typically hypothyroidism observed following a transient thyroiditis-induced thyrotoxicosis) followed by the rest of the endocrine organs. The median time frame from the start of the treatment to the development of thyroid symptoms, most commonly hypothyroidism, is 6 weeks, followed by pituitary (hypophysitis), adrenals (primary adrenal insufficiency) and  $\beta$ -cells of the pancreas (insulin deficient diabetes, analogous to type 1 diabetes) (170). These are different from the side-effects brought on by conventional cytotoxic chemotherapy or even more recent molecular-targeted medicines, which infrequently result in endocrine dysfunction (171).

Skin disorders. Non-specific adverse events such as maculopapular rash, pruritus, psoriasiform, eczematous and lichenoid dermatosis are among the most prevalent (172,173). Compared to anti-PD-1 monotherapy, the maculopapular rash phenotype is more prevalent when CTLA-4 inhibition is implemented (21). Bullous pemphigoid, vitiligo-like skin hypopigmentation/depigmentation and alopecia are other less-common irCAEs (174,175). Although severe reactions, such as Stevens-Johnson syndrome, toxic epidermal necrolysis and drug reaction with eosinophilia and systemic symptoms are uncommon, cutaneous consequences are typically self-limiting (174-176). Early diagnosis and the administration of corticosteroids or antitumor necrosis factor-agents are the foundation of treatment algorithms for irCAEs (176,177). However, the use of corticosteroids before or after ICI initiation may result in a diminished antitumor efficacy. Anti-CTLA-4 and anti-PD1 therapy have both been associated with reports of vitiligo (178). The occurrence of skin hypopigmentation or depigmentation such as vitiligo has been linked to an extensive anticancer benefit from drug treatment in patients with melanoma. Vitiligo has been proven as a positive predictive factor in measuring the tumor response to treatment. In comparison with the general population, patients with melanoma have a 10-fold increased incidence of drug-related cutaneous hypopigmentation and depigmentation (179). Since the PD-L1:PD1 pathway mostly regulates the peripheral tolerance of melanosomal proteins (such as tyrosinase and TRP-2), the interference of PD-1 signaling may result in autoimmune vitiligo (180). This offers a reasonable explanation for the onset and durability of depigmentation in patients receiving immunotherapy.

*Lungs*. Case series studies have demonstrated that patients develop organizing pneumonia, diffuse alveolar damage, acute respiratory distress syndrome (ARDS) and non-specific interstitial pneumonia, which is then managed by intravenous and oral steroids (181-185).

*Liver and kidneys*. A previous meta-analysis revealed adverse effects associated with the use of anti-PD-1/PD-L1 monoclonal antibodies for malignancies with an increased incidence of pancreatitis, and increased levels of liver enzymes, such as aspartate aminotransferase and alanine transaminase, elevated creatinine levels, nephritis and renal failure (164).

### 9. Mechanisms of resistance

A myriad of ongoing clinical trials and practices have discovered multiple mechanisms leading to resistance to ICIs. More precisely, these include changes in the tumor microenvironment prohibiting T-cell interaction, tumor invasion and tumor cell destruction by effector mechanisms. The key to tumor cell destruction via effector T-cells is through the processing of tumor antigens to antigen-presenting cells. The failure of antigen-presenting components in this pathway is a major cause of resistance in melanoma (186). ß2 microglobulin is an key molecule responsible for the folding and transportation of major histocompatibility complex-1 to the surface of cells. Mutations in these molecules have been noted in patients with melanoma at the time of anti-PD1 treatment failure (187). Other mechanisms responsible for limiting T-cell trafficking in the tumor microenvironment include mutations in BRAF, and the inhibition of PTEN. This leads to the increased expression of immunosuppressive molecules, such as VEGF (188). It also inhibits the migration and trafficking of effector T-cells (189). In addition to these tumor-intrinsic mechanisms, various tumor-extrinsic mechanisms also play a role in the development of resistance to ICIs. These include the development of new inhibitory checkpoints, immunosuppressive cytokines and molecules in the tumor microenvironment suppressing immune cell function. One such example is the production of transforming growth factor  $\beta$  (TGF- $\beta$ ) by tumor cells. TGF- $\beta$  is an immunosuppressive cytokine that functions by stimulating Tregs and inhibiting the cytotoxicity of effector T-cells (190).

To summarize, understanding and investigating the potential mechanisms that lead to resistance to ICIs is crucial in developing effective strategies to guide therapy. Further studies are required to identify new mechanisms and develop targeted therapies to improve the clinical outcome of patients undertaking immunotherapy.

# **10.** Emerging newer therapeutic strategies: Targeting tumor metabolic dependencies

Tumor cells sustain themselves by utilizing altered metabolic pathways by using nutrients, such as glucose, tryptophan and arginine to produce toxic metabolites such as adenosine, lactate and kynurenine (191,192). Such toxic metabolites produce an unfavorable environment for the antitumor cells to function resulting in increased expression of immune checkpoints and expansion of Tregs (193).

The mechanism that tumor cells use is the mutation in the myelocytomatosis oncogene (MYC) and PI3K/AKT/mammalian target of rapamycin (mTOR) signaling pathways. The increased expression of hypoxia-inducible factor-1- $\alpha$  leads to the overexpression of the PI3K/AKT/mTOR pathway, as well as glucose transporters such as glucose transporter 1, leading to increased glucose consumption and acidification of the tumor microenvironment (194,195). As hypoxia is generated, glucose depletion occurs and increased toxic waste is produced within the tumor microenvironment, resulting in the inhibition of tumor antigen presentation by APCs (196). Thus, there is an overall decrease in the antitumor immune response by T-effector, macrophages or NK cells, while pro-tumor immune cells such as Tregs proliferate to increase the expression of inhibitory checkpoint ligand PD-1 on immune cells, inhibiting the antitumor immunity (197). With the advancement of technologies, newer therapeutic strategies that target the immunosuppressive tumor microenvironment generated by tumor cells may be developed to reprogram the behavior of immune cells, leading to an improved efficacy in terms of the treatment response.

One of the important T-cellular processes is the activation of the PI3K pathway, which plays a vital role in proliferation and differentiation. Monotherapy, which inhibits the PI3K pathway, has not yielded any significant results in the treatment of cancer; however, combining PI3K inhibitors and the PD-1-PDL1 blockade has shown some notable results (198). The loss of PTEN, which is a PI3K-inhibiting tumor suppressor often mutated in tumor cells, results in the uncontrolled growth of tumor cells and escapes the immune destruction imposed on it. As previously demonstrated, when mice with PTEN-null melanoma were treated *in vivo* with the PI3K $\beta$  inhibitor, GSK2636771, this resulted in a decreased AKT phosphorylation and the activation of mTOR targets. Additionally, when it was combined with an anti-PD1 antibody, it markedly improved the survival and increased immune response with reduced tumor cell mass (199). With such promising results, a number of newer anti-PI3K medicines are being developed and tested to increase efficacy (NCT01390818). Despite this, more novel promising approaches are needed to prove the success of combining anti-PI3K drugs with ICIs in the treatment of melanoma (200).

### 11. Conclusion and future perspectives

The development of ICIs and targeted therapies has played a crucial role in revolutionizing the management of melanomas by improving the overall and progression-free survival. Although both of these therapies have advantages and disadvantages, combination therapy (ICI + ICI, or ICI + targeted therapies) has been found to be more effective in improving patient outcomes. However, there is limited literature available regarding combination therapies and different types of potential combinations. There are also insufficient data on patients and their responses to draw sufficient conclusions. The development of drug-related adverse effects with the use of combination therapies is also a debatable question. However, when developing newer ICIs to achieve a more effective response, a focus should certainly be placed on the integration of nanotechnology or antibody engineering. Through these, one can increase drug delivery to a specific target and thus increase overall response. In addition, focusing on epigenetic modulation and developing ICIs that target those changes can enhance the responsiveness of ICIs.

There may be concerns regarding resistance to ICIs in patients with melanoma. Some patients may have resistance to certain ICIs from the beginning or may develop them as an acquired resistance with subsequent treatment after progression of a tumor with clinical benefit. Further treatment decisions shall be made on the basis of evaluation of the tumor and factors related to the patient, focusing on targeted therapeutic drugs, other immunotherapy drugs, cellular therapies, intralesional therapies, or chemotherapy. It is important to tailor ICI treatment based on an individual's genetic makeup and tumor characteristics to decrease the resistance. The early identification of tumor biomarkers can predict future responses to particular ICIs and may help to select a personalized treatment strategy. Furthermore, with the use of tumor metabolic pathway inhibitors in combination with ICIs, targeting signaling pathways and immune responses can be better used to overcome potential resistance to ICIs than when used alone.

Trials are being conducted on newer inhibitory immune checkpoint targets, as well as certain inhibitory targets beyond immune checkpoints. These include LAF-3, TIM-3, B7-H3 and B7-H4, CD73, etc. which are immune checkpoints, and CEACAM1, CEACAM5/6, CCL2/CCR2, etc. which are other inhibitory targets (201). It is essential to maintain enrollment in clinical trials so that newer ICIs, additional inhibitory treatments, combination therapy, and mechanisms of resistance and methods of overcoming the resistance can all be further investigated.

In conclusion, with the increasing incidence of melanoma over the past two decades, managing it with different treatment modalities has become cumbersome. With the limited effectiveness of the traditional approach using chemotherapy and immunotherapy, the role of newer treatment modalities should be given equal emphasis. Novel approaches using ICIs have been a revolution in the therapeutic approach by unleashing the immune system's ability to recognize and eliminate cancer cells. Ipilimumab, nivolumab and pembrolizumab have been shown to lead to a substantial improvement in the overall survival of patients with advanced melanoma, particularly in high-risk metastatic melanoma compared to traditional therapies. However, with ICIs, it is paramount to monitor any side-effects, and to ensure the optimal outcome is achieved using personalized treatment approaches.

### Acknowledgements

Not applicable.

### Funding

No funding was received.

### Availability of data and materials

Not applicable.

### Authors' contributions

VS, VP and AS were involved in the conceptualization of the study. VS, VP, AS, BV, SB and SA were involved in the curation of data from the literature for inclusion in the present review. VS, VP and AS were involved in selecting the relevant literature. VS, VP and AS were involved in project administration. VS supervised the study. VS, VP, AS, BV, SB and SA were involved in the writing of the original draft. VS, VP and AS were involved in the writing, review and editing of the manuscript. All authors have read and approved the final manuscript. Data authentication is not applicable.

### Ethics approval and consent to participate

Not applicable.

#### Patient consent for publication

Not applicable.

### **Competing interests**

The authors declare that they have no competing interests.

### References

- 1. Finn L, Markovic SN and Joseph RW: Therapy for metastatic melanoma: The past, present, and future. BMC Med 10: 23, 2012.
- Arnold M, Singh D, Laversanne M, Vignat J, Vaccarella S, Meheus F, Cust AE, de Vries E, Whiteman DC and Bray F: Global burden of cutaneous melanoma in 2020 and projections to 2040. JAMA Dermatol 158: 495-503, 2022.
- 3. Miller AJ and Mihm MC Jr: Melanoma. N Engl J Med 355: 51-65, 2006.

- 4. Rosenberg SA, Lotze MT, Yang JC, Topalian SL, Chang AE, Schwartzentruber DJ, Aebersold P, Leitman S, Linehan WM, Seipp CA, *et al*: Prospective randomized trial of high-dose interleukin-2 alone or in conjunction with lymphokine-activated killer cells for the treatment of patients with advanced cancer. J Natl Cancer Inst 85: 622-632, 1993.
- 5. Bronte V and Mocellin S: Suppressive influences in the immune response to cancer. J Immunother 32: 1-11, 2009.
- 6. Mellman I, Coukos G and Dranoff G: Cancer immunotherapy comes of age. Nature 480: 480-489, 2001.
- 7. Pardoll DM: The blockade of immune checkpoints in cancer immunotherapy. Nat Rev Cancer 12: 252-264, 2012.
- Okazaki T, Okazaki IM, Wang J, Sugiura D, Nakaki F, Yoshida T, Kato Y, Fagarasan S, Muramatsu M, Eto T, *et al*: PD-1 and LAG-3 inhibitory co-receptors act synergistically to prevent autoimmunity in mice. J Exp Med 208: 395-407, 2011.
- Fourcade J, Sun Z, Pagliano O, Chauvin JM, Sander C, Janjic B, Tarhini AA, Tawbi HA, Kirkwood JM, Moschos S, *et al*: PD-1 and Tim-3 regulate the expansion of tumor antigen-specific CD8<sup>+</sup> T cells induced by melanoma vaccines. Cancer Res 74: 1045-1055, 2014.
- Lines JL, Pantazi E, Mak J, Sempere LF, Wang L, O'Connell S, Ceeraz S, Suriawinata AA, Yan S, Ernstoff MS and Noelle R: VISTA is an immune checkpoint molecule for human T-cells. Cancer Res 74: 1924-1932, 2014.
- 11. Hanaizi Z, van Zwieten-Boot B, Calvo G, Lopez AS, van Dartel M, Camarero J, Abadie E and Pignatti F: The European medicines agency review of ipilimumab (Yervoy) for the treatment of advanced (unresectable or metastatic) melanoma in adults who have received prior therapy: Summary of the scientific assessment of the committee for medicinal products for human use. Eur J Cancer 48: 237-242, 2012.
- 12. Tarhini AA: Tremelimumab: A review of development to date in solid tumors. Immunotherapy 5: 215-229, 2013.
- Wang D, Wang T, Liu J, Yu H, Jiao S, Feng B, Zhou F, Fu Y, Yin Q, Zhang P, *et al*: Acid-activatable versatile micelleplexes for PD-L1 blockade-enhanced cancer photodynamic immunotherapy. Nano Lett 16: 5503-5513, 2016.
- Ottaviano M, De Placido S and Ascierto PA: Recent success and limitations of immune checkpoint inhibitors for cancer: A lesson from melanoma. Virchows Arch 474: 421-432, 2019.
- Chambers CA, Sullivan TJ and Allison JP: Lymphoproliferation in CTLA-4-deficient mice is mediated by costimulationdependent activation of CD4<sup>+</sup> T-cells. Immunity 7: 885-895, 1997.
- 16. Tivol EA, Borriello F, Schweitzer AN, Lynch WP, Bluestone JA and Sharpe AH: Loss of CTLA-4 leads to massive lymphoproliferation and fatal multiorgan tissue destruction, revealing a critical negative regulatory role of CTLA-4. Immunity 3: 541-547, 1995.
- Waterhouse P, Penninger JM, Timms E, Wakeham A, Shahinian A, Lee KP, Thompson CB, Griesser H and Mak TW: Lymphoproliferative disorders with early lethality in mice deficient in Ctla-4. Science 270: 985-988, 1995.
- Walker LSK and Sansom DM: The emerging role of CTLA4 as a cell-extrinsic regulator of T cell responses. Nat Rev Immunol 11: 852-863, 2011.
- Ménard C, Ghiringhelli F, Roux S, Chaput N, Mateus C, Grohmann U, Caillat-Zucman S, Zitvogel L and Robert C: Ctla-4 blockade confers lymphocyte resistance to regulatory T-cells in advanced melanoma: Surrogate marker of efficacy of tremelimumab? Clin Cancer Res 14: 5242-5249, 2008.
- 20. Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, Gonzalez R, Robert C, Schadendorf D, Hassel JC, *et al*: Improved survival with ipilimumab in patients with metastatic melanoma. N Engl J Med 363: 711-723, 2010.
- with metastatic melanoma. N Engl J Med 363: 711-723, 2010.
  21. Phan GQ, Yang JC, Sherry RM, Hwu P, Topalian SL, Schwartzentruber DJ, Restifo NP, Haworth LR, Seipp CA, Freezer LJ, *et al*: Cancer regression and autoimmunity induced by cytotoxic T lymphocyte-associated antigen 4 blockade in patients with metastatic melanoma. Proc Natl Acad Sci USA 100: 8372-8377, 2003.
- 22. Malek TR and Castro I: Interleukin-2 receptor signaling: At the interface between tolerance and immunity. Immunity 33: 153-165, 2010.
- 23. Reuben JM, Lee BN, Li C, Gomez-Navarro J, Bozon VA, Parker CA, Hernandez IM, Gutierrez C, Lopez-Berestein G and Camacho LH: Biologic and immunomodulatory events after CTLA-4 blockade with ticilimumab in patients with advanced malignant melanoma. Cancer 106: 2437-2444, 2006.

- 24. Ribas A, Comin-Anduix B, Economou JS, Donahue TR, de la Rocha P, Morris LF, Jalil J, Dissette VB, Shintaku IP, Glaspy JA, *et al*: Intratumoral immune cell infiltrates, FoxP3, and indoleamine 2,3-dioxygenase in patients with melanoma undergoing CTLA4 blockade. Clin Cancer Res 15: 390-399, 2009.
- 25. Robert C, Thomas L, Bondarenko I, O'Day S, Weber J, Garbe C, Lebbe C, Baurain JF, Testori A, Grob JJ, et al: Ipilimumab plus dacarbazine for previously untreated metastatic melanoma. N Engl J Med 364: 2517-2526, 2011.
- Buchbinder EI and Desai A: CTLA-4 and PD-1 pathways: Similarities, differences, and implications of their inhibition. Am J Clin Oncol 39: 98-106, 2016.
- 27. Amarnath S, Mangus CW, Wang JCM, Wei F, He A, Kapoor V, Foley JE, Massey PR, Felizardo TC, Riley JL, *et al*: The PDL1-PD1 axis converts human TH1 cells into regulatory T-cells. Sci Transl Med 3: 111ra120, 2011.
- 28. Spranger S, Spaapen RM, Zha Y, Williams J, Meng Y, Ha TT and Gajewski TF: Up-regulation of PD-L1, IDO, and T(regs) in the melanoma tumor microenvironment is driven by CD8(+) T cells. Sci Transl Med 5: 200ra116, 2013.
- 29. Sun Z, Fourcade J, Pagliano O, Chauvin JM, Sander C, Kirkwood JM and Zarour HM: IL10 and PD-1 cooperate to limit the activity of tumor-specific CD8<sup>+</sup> T cells. Cancer Res 75: 1635-1644, 2015.
- 30. Zou W and Chen L: Inhibitory B7-family molecules in the tumour microenvironment. Nat Rev Immunol 8: 467-477, 2008.
- Kinter AL, Godbout EJ, McNally JP, Sereti I, Roby GA, O'Shea MA and Fauci AS: The common gamma-chain cytokines IL-2, IL-7, IL-15, and IL-21 induce the expression of programmed death-1 and its ligands. J Immunol 181: 6738-6746, 2008.
- 32. Yang J, Riella LV, Chock S, Liu T, Zhao X, Yuan X, Paterson AM, Watanabe T, Vanguri V, Yagita H, *et al*: The novel costimulatory programmed death ligand 1/B7.1 pathway is functional in inhibiting alloimmune responses in vivo. J Immunol 187: 1113-1119, 2011.
- Krönig H, Julia Falchner K, Odendahl M, Brackertz B, Conrad H, Muck D, Hein R, Blank C, Peschel C, Haller B, *et al*: PD-1 expression on Melan-A-reactive T cells increases during progression to metastatic disease. Int J Cancer 130: 2327-2336, 2012.
   Hamid O, Robert C, Daud A, Hodi FS, Hwu WJ, Kefford R,
- 34. Hamid O, Robert C, Daud A, Hodi FS, Hwu WJ, Kefford R, Wolchok JD, Hersey P, Joseph RW, Weber JS, *et al*: Safety and tumor responses with lambrolizumab (anti-PD-1) in melanoma. N Engl J Med 369: 134-144, 2013.
- Francisco LM, Salinas VH, Brown KE, Vanguri VK, Freeman GJ, Kuchroo VK and Sharpe AH: PD-L1 regulates the development, maintenance, and function of induced regulatory T cells. J Exp Med 206: 3015-3029, 2009.
- 36. Larkin J, Chiarion-Sileni V, Gonzalez R, Grob JJ, Cowey CL, Lao CD, Schadendorf D, Dummer R, Smylie M, Rutkowski P, *et al*: Combined nivolumab and ipilimumab or monotherapy in untreated melanoma. N Engl J Med 373: 23-34, 2015.
- 37. Wolchok JD, Chiarion-Sileni V, Gonzalez R, Rutkowski P, Grob JJ, Cowey CL, Lao CD, Wagstaff J, Schadendorf D, Ferrucci PF, *et al*: Overall survival with combined nivolumab and ipilimumab in advanced melanoma. N Engl J Med 377: 1345-1356, 2017.
- Marconcini R, Spagnolo F, Stucci LS, Ribero S, Marra E, Rosa F, Picasso V, Di Guardo L, Cimminiello C, Cavalieri S, *et al*: Current status and perspectives in immunotherapy for metastatic melanoma. Oncotarget 9: 12452-12470, 2018.
- Watanabe N, Gavrieli M, Sedy JR, Yang J, Fallarino F, Loftin SK, Hurchla MA, Zimmerman N, Sim J, Zang X, *et al*: BTLA is a lymphocyte inhibitory receptor with similarities to CTLA-4 and PD-1. Nat Immunol 4: 670-679, 2003.
- Murphy KM, Nelson CA and Sedý JR: Balancing co-stimulation and inhibition with BTLA and HVEM. Nat Rev Immunol 6: 671-681, 2006.
- 41. Fourcade J, Sun Z, Pagliano O, Guillaume P, Luescher IF, Sander C, Kirkwood JM, Olive D, Kuchroo V and Zarour HM: CD8(+) T cells specific for tumor antigens can be rendered dysfunctional by the tumor microenvironment through upregulation of the inhibitory receptors BTLA and PD-1. Cancer Res 72: 887-896, 2012.
- 42. Le Mercier I, Chen W, Lines JL, Day M, Li J, Sergent P, Noelle RJ and Wang L: VISTA regulates the development of protective antitumor immunity. Cancer Res 74: 1933-1944, 2014.
- 43. Monney L, Sabatos CA, Gaglia JL, Ryu A, Waldner H, Chernova T, Manning S, Greenfield EA, Coyle AJ, Sobel RA, *et al*: Th1-specific cell surface protein Tim-3 regulates macrophage activation and severity of an autoimmune disease. Nature 415: 536-541, 2002.

- 44. Anderson AC, Anderson DE, Bregoli L, Hastings WD, Kassam N, Lei C, Chandwaskar R, Karman J, Su EW, Hirashima M, *et al*: Promotion of tissue inflammation by the immune receptor Tim-3 expressed on innate immune cells. Science 318: 1141-1143, 2007.
- 45. Zhu C, Anderson AC, Schubart A, Xiong H, Imitola J, Khoury SJ, Zheng XX, Strom TB and Kuchroo VK: The Tim-3 ligand galectin-9 negatively regulates T helper type 1 immunity. Nat Immunol 6: 1245-1252, 2005.
- 46. Sabatos CA, Chakravarti S, Cha E, Schubart A, Sánchez-Fueyo A, Zheng XX, Coyle AJ, Strom TB, Freeman GJ and Kuchroo VK: Interaction of Tim-3 and Tim-3 ligand regulates T helper type 1 responses and induction of peripheral tolerance. Nat Immunol 4: 1102-1110, 2003.
- 47. Ngiow SF, von Scheidt B, Akiba H, Yagita H, Teng MWL and Smyth MJ: Anti-TIM3 antibody promotes T cell IFN-γ-mediated antitumor immunity and suppresses established tumors. Cancer Res 71: 3540-3551, 2011.
- 48. Advani R, Flinn I, Popplewell L, Forero A, Bartlett NL, Ghosh N, Kline J, Roschewski M, LaCasce A, Collins GP, et al: CD47 blockade by Hu5F9-G4 and rituximab in non-Hodgkin's lymphoma. N Engl J Med 379: 1711-1721, 2018.
- 49. Ascierto PA, Melero I, Bhatia S, Bono P, Sanborn RE, Lipson EJ, Callahan MK, Gajewski T, Gomez-Roca CA, Hodi FS, *et al*: Initial efficacy of anti-lymphocyte activation gene-3 (anti-LAG-3; BMS-986016) in combination with nivolumab (nivo) in pts with melanoma (MEL) previously treated with anti-PD-1/PD-L1 therapy. J Clin Orthod 35 (15 Suppl): S9520, 2017.
- 50. Wei SC, Levine JH, Cogdill AP, Zhao Y, Anang NAS, Andrews MC, Sharma P, Wang J, Wargo JA, Pe'er D and Allison JP: Distinct cellular mechanisms underlie anti-CTLA-4 and anti-PD-1 checkpoint blockade. Cell 170: 1120-1133.e17, 2017.
- 51. Rotte A, Jin JY and Lemaire V: Mechanistic overview of immune checkpoints to support the rational design of their combinations in cancer immunotherapy. Ann Oncol 29: 71-83, 2018.
- 52. Tarhini A: Immune-mediated adverse events associated with ipilimumab ctla-4 blockade therapy: The underlying mechanisms and clinical management. Scientifica (Cairo) 2013: 857519, 2013.
- 53. Michot JM, Bigenwald C, Champiat S, Collins M, Carbonnel F, Postel-Vinay S, Berdelou A, Varga A, Bahleda R, Hollebecque A, *et al*: Immune-related adverse events with immune checkpoint blockade: A comprehensive review. Eur J Cancer 54: 139-148, 2016.
- 54. Tawbi HA, Schadendorf D, Lipson EJ, Ascierto PA, Matamala L, Castillo Gutiérrez E, Rutkowski P, Gogas HJ, Lao CD, De Menezes JJ, *et al*: Relatlimab and nivolumab versus nivolumab in untreated advanced melanoma. N Engl J Med 386: 24-34, 2022.
- 55. Olson DJ, Eroglu Z, Brockstein B, Poklepovic AS, Bajaj M, Babu S, Hallmeyer S, Velasco M, Lutzky J, Higgs E, *et al*: Pembrolizumab plus ipilimumab following anti-PD-1/L1 failure in melanoma. J Clin Oncol 39: 2647-2655, 2021.
- 56. Weber JS, Gibney G, Sullivan RJ, Sosman JA, Slingluff CL Jr, Lawrence DP, Logan TF, Schuchter LM, Nair S, Fecher L, *et al*: Sequential administration of nivolumab and ipilimumab with a planned switch in patients with advanced melanoma (CheckMate 064): An open-label, randomised, phase 2 trial. Lancet Oncol 17: 943-955, 2016.
- 57. Shoushtari AN, Wagstaff J, Ascierto PA, Butler MO, Lao CD, Marquez-Rodas I, Chiarion-Sileni V, Dummer R, Ferrucci PF, Lorigan P, *et al*: CheckMate 067: Long-term outcomes in patients with mucosal melanoma. J Clin Orthod 38 (15 Suppl): S10019, 2020.
- 58. Pradeep J, Win TT, Aye SN and Sreeramareddy CT: Efficacy and safety of immune checkpoint inhibitors for advanced malignant melanoma: A meta-analysis on monotherapy vs combination therapy. J Cancer 13: 3091-3102, 2022.
- 59. Amaria RN, Reddy SM, Tawbi HA, Davies MA, Ross MI, Glitza IC, Cormier JN, Lewis C, Hwu WJ, Hanna E, *et al*: Neoadjuvant immune checkpoint blockade in high-risk resectable melanoma. Nat Med 24: 1649-1654, 2018.
- able melanoma. Nat Med 24: 1649-1654, 2018.
  60. Hodi FS, Chesney J, Pavlick AC, Robert C, Grossmann KF, McDermott DF, Linette GP, Meyer N, Giguere JK, Agarwala SS, *et al*: Combined nivolumab and ipilimumab versus ipilimumab alone in patients with advanced melanoma: 2-Year overall survival outcomes in a multicentre, randomised, controlled, phase 2 trial. Lancet Oncol 17: 1558-1568, 2016.

- 61. Hodi FS, Chiarion-Sileni V, Gonzalez R, Grob JJ, Rutkowski P, Cowey CL, Lao CD, Schadendorf D, Wagstaff J, Dummer R, *et al*: Nivolumab plus ipilimumab or nivolumab alone versus ipilimumab alone in advanced melanoma (CheckMate 067): 4-Year outcomes of a multicentre, randomised, phase 3 trial. Lancet Oncol 19: 1480-1492, 2018.
- 62. Larkin J, Chiarion-Sileni V, Gonzalez R, Grob JJ, Rutkowski P, Lao CD, Cowey CL, Schadendorf D, Wagstaff J, Dummer R, *et al*: Five-year survival with combined nivolumab and ipilimumab in advanced melanoma. N Engl J Med 381: 1535-1546, 2019.
- 63. Postow MA, Chesney J, Pavlick AC, Robert C, Grossmann K, McDermott D, Linette GP, Meyer N, Giguere JK, Agarwala SS, *et al*: Nivolumab and ipilimumab versus ipilimumab in untreated melanoma. N Engl J Med 372: 2006-2017, 2015.
- 64. Long GV, Atkinson V, Lo S, Sandhu S, Guminski AD, Brown MP, Wilmott JS, Edwards J, Gonzalez M, Scolyer RA, *et al*: Combination nivolumab and ipilimumab or nivolumab alone in melanoma brain metastases: A multicentre randomised phase 2 study. Lancet Oncol 19: 672-681, 2018.
- 65. Wagle N, Emery C, Berger MF, Davis MJ, Sawyer A, Pochanard P, Kehoe SM, Johannessen CM, Macconaill LE, Hahn WC, *et al*: Dissecting therapeutic resistance to RAF inhibition in melanoma by tumor genomic profiling. J Clin Oncol 29: 3085-3096, 2011.
- 66. Gorre ME, Mohammed M, Ellwood K, Hsu N, Paquette R, Rao PN and Sawyers CL: Clinical resistance to STI-571 cancer therapy caused by BCR-ABL gene mutation or amplification. Science 293: 876-880, 2001.
- 67. Ellis LM and Hicklin DJ: Resistance to targeted therapies: Refining anticancer therapy in the era of molecular oncology. Clin Cancer Res 15: 7471-7478, 2009.
- 68. Nazarian R, Shi H, Wang Q, Kong X, Koya RC, Lee H, Chen Z, Lee MK, Attar N, Sazegar H, *et al*: Melanomas acquire resistance to B-RAF(V600E) inhibition by RTK or N-RAS upregulation. Nature 468: 973-977, 2010.
- 69. Johannessen CM, Boehm JS, Kim SY, Thomas SR, Wardwell L, Johnson LA, Emery CM, Stransky N, Cogdill AP, Barretina J, *et al*: COT drives resistance to RAF inhibition through MAP kinase pathway reactivation. Nature 468: 968-972, 2010.
- Montagut C, Sharma SV, Shioda T, McDermott U, Ulman M, Ulkus LE, Dias-Santagata D, Stubbs H, Lee DY, Singh A, *et al*: Elevated CRAF as a potential mechanism of acquired resistance to BRAF inhibition in melanoma. Cancer Res 68: 4853-4861, 2008.
- 71. Villanueva J, Vultur A, Lee JT, Somasundaram R, Fukunaga-Kalabis M, Cipolla AK, Wubbenhorst B, Xu X, Gimotty PA, Kee D, *et al*: Acquired resistance to BRAF inhibitors mediated by a RAF kinase switch in melanoma can be overcome by cotargeting MEK and IGF-1R/PI3K. Cancer Cell 18: 683-695, 2010.
- 72. Turke AB, Zejnullahu K, Wu YL, Song Y, Dias-Santagata D, Lifshits E, Toschi L, Rogers A, Mok T, Sequist L, *et al*: Preexistence and clonal selection of MET amplification in EGFR mutant NSCLC. Cancer Cell 17: 77-88, 2010.
- Fingelman JA, Zejnullahu K, Mitsudomi T, Song Y, Hyland C, Park JO, Lindeman N, Gale CM, Zhao X, Christensen J, *et al*: MET amplification leads to gefitinib resistance in lung cancer by activating ERBB3 signaling. Science 316: 1039-1043, 2007.
   Guix M, Faber AC, Wang SE, Olivares MG, Song Y, Qu S,
- 74. Guix M, Faber AC, Wang SE, Olivares MG, Song Y, Qu S, Rinehart C, Seidel B, Yee D, Arteaga CL and Engelman JA: Acquired resistance to EGFR tyrosine kinase inhibitors in cancer cells is mediated by loss of IGF-binding proteins. J Clin Invest 118: 2609-2619, 2008.
- 75. Paraiso KHT, Xiang Y, Rebecca VW, Abel EV, Chen YA, Munko AC, Wood E, Fedorenko IV, Sondak VK, Anderson AR, *et al*: PTEN loss confers BRAF inhibitor resistance to melanoma cells through the suppression of BIM expression. Cancer Res 71: 2750-2760, 2011.
- 76. Maio M, Grob JJ, Aamdal S, Bondarenko I, Robert C, Thomas L, Garbe C, Chiarion-Sileni V, Testori A, Chen TT, *et al*: Five-year survival rates for treatment-naive patients with advanced melanoma who received ipilimumab plus dacarbazine in a phase III trial. J Clin Oncol 33: 1191-1196, 2015.
- 77. Eggermont AMM, Chiarion-Sileni V, Grob JJ, Dummer R, Wolchok JD, Schmidt H, Hamid O, Robert C, Ascierto PA, Richards JM, *et al*: Prolonged survival in stage III melanoma with ipilimumab adjuvant therapy. N Engl J Med 375: 1845-1855, 2016.

- 78. Ascierto PA, Del Vecchio M, Robert C, Mackiewicz A, Chiarion-Sileni V, Arance A, Lebbé C, Bastholt L, Hamid O, Rutkowski P, *et al*: Ipilimumab 10 mg/kg versus ipilimumab 3 mg/kg in patients with unresectable or metastatic melanoma: A randomised, double-blind, multicentre, phase 3 trial. Lancet Oncol 18: 611-622, 2017.
- 79. Robert C, Long GV, Brady B, Dutriaux C, Maio M, Mortier L, Hassel JC, Rutkowski P, McNeil C, Kalinka-Warzocha E, *et al*: Nivolumab in previously untreated melanoma without BRAF mutation. N Engl J Med 372: 320-330, 2015.
- 80. Weber JS, D'Angelo SP, Minor D, Hodi FS, Gutzmer R, Neyns B, Hoeller C, Khushalani NI, Miller WH Jr, Lao CD, et al: Nivolumab versus chemotherapy in patients with advanced melanoma who progressed after anti-CTLA-4 treatment (CheckMate 037): A randomised, controlled, open-label, phase 3 trial. Lancet Oncol 16: 375-384, 2015.
- 81. Larkin J, Minor D, D'Angelo S, Neyns B, Smylie M, Miller WH Jr, Gutzmer R, Linette G, Chmielowski B, Lao CD, et al: Overall survival in patients with advanced melanoma who received nivolumab versus investigator's choice chemotherapy in CheckMate 037: A randomized, controlled, open-label phase III trial. J Clin Oncol 36: 383-390, 2018.
- 82. Weber J, Mandala M, Del Vecchio M, Gogas HJ, Arance AM, Cowey CL, Dalle S, Schenker M, Chiarion-Sileni V, Marquez-Rodas I, *et al*: Adjuvant nivolumab versus ipilimumab in resected stage III or IV melanoma. N Engl J Med 377: 1824-1835, 2017.
- 83. Ribas A, Puzanov I, Dummer R, Schadendorf D, Hamid O, Robert C, Hodi FS, Schachter J, Pavlick AC, Lewis KD, et al: Pembrolizumab versus investigator-choice chemotherapy for ipilimumab-refractory melanoma (KEYNOTE-002): A randomised, controlled, phase 2 trial. Lancet Oncol 16: 908-918, 2015.
- 84. Robert C, Ribas A, Schachter J, Arance A, Grob JJ, Mortier L, Daud A, Carlino MS, McNeil CM, Lotem M, et al: Pembrolizumab versus ipilimumab in advanced melanoma (KEYNOTE-006): Post-hoc 5-year results from an open-label, multicentre, randomised, controlled, phase 3 study. Lancet Oncol 20: 1239-1251, 2019.
- 85. Schachter J, Ribas A, Long GV, Arance A, Grob JJ, Mortier L, Daud A, Carlino MS, McNeil C, Lotem M, et al: Pembrolizumab versus ipilimumab for advanced melanoma: Final overall survival results of a multicentre, randomised, open-label phase 3 study (KEYNOTE-006). Lancet 390: 1853-1862, 2017.
- 86. Long GV, Atkinson V, Ascierto PA, Robert C, Hassel JC, Rutkowski P, Savage KJ, Taylor F, Coon C, Gilloteau I, et al: Effect of nivolumab on health-related quality of life in patients with treatment-naïve advanced melanoma: Results from the phase III CheckMate 066 study. Ann Oncol 27: 1940-1946, 2016.
- 87. Schadendorf D, Dummer R, Hauschild A, Robert C, Hamid O, Daud A, van den Eertwegh A, Cranmer L, O'Day S, Puzanov I, *et al*: Health-related quality of life in the randomised KEYNOTE-002 study of pembrolizumab versus chemotherapy in patients with ipilimumab-refractory melanoma. Eur J Cancer 67: 46-54, 2016.
- 88. Nosrati A, Tsai KK, Goldinger SM, Tumeh P, Grimes B, Loo K, Algazi AP, Nguyen-Kim TDL, Levesque M, Dummer R, *et al*: Evaluation of clinicopathological factors in PD-1 response: derivation and validation of a prediction scale for response to PD-1 monotherapy. Br J Cancer 116: 1141-1147, 2017.
- Zhang Y, Liu B, Kotenko S and Li W: Prognostic value of neutrophil-lymphocyte ratio and lactate dehydrogenase in melanoma patients treated with immune checkpoint inhibitors: A systematic review and meta-analysis. Medicine (Baltimore) 101: e29536, 2022.
- 90. Gershenwald JE, Scolyer RA, Hess KR, Sondak VK, Long GV, Ross MI, Lazar AJ, Faries MB, Kirkwood JM, McArthur GA, *et al*: Melanoma staging: Evidence-based changes in the American joint committee on cancer eighth edition cancer staging manual. CA Cancer J Clin 67: 472-492, 2017.
- Balch CM, Gershenwald JE, Soong SJ, Soong SJ, Thompson JF, Atkins MB, Byrd DR, Buzaid AC, Cochran AJ, Coit DG, et al: Final version of 2009 AJCC melanoma staging and classification. J Clin Oncol 27: 6199-6206, 2009.
- 92. Hauschild A, Engel G, Brenner W, Gläser R, Mönig H, Henze E and Christophers E: S100B protein detection in serum is a significant prognostic factor in metastatic melanoma. Oncology 56: 338-344, 1999.

- 93. Jury CS, McAllister EJ and MacKie RM: Rising levels of serum S100 protein precede other evidence of disease progression in patients with malignant melanoma. Br J Dermatol 143: 269-274, 2000.
- 94. Mårtenson ED, Hansson LO, Nilsson B, von Schoultz E, Månsson Brahme E, Ringborg U and Hansson J: Serum S-100b protein as a prognostic marker in malignant cutaneous melanoma. J Clin Oncol 19: 824-831, 2001.
- 95. Janka EA, Várvölgyi T, Sipos Z, Soós A, Hegyi P, Kiss S, Dembrovszky F, Csupor D, Kéringer P, Pécsi D, *et al*: Predictive performance of serum S100B versus LDH in melanoma patients: A systematic review and meta-analysis. Front Oncol 11: 772165, 2021.
- 96.Friedman RC, Farh KKH, Burge CB and Bartel DP: Most mammalian mRNAs are conserved targets of microRNAs. Genome Res 19: 92-105, 2009.
- 97. Lim LP, Glasner ME, Yekta S, Burge CB and Bartel DP: Vertebrate microRNA genes. Science 299: 1540, 2003.
- 98. Lagos-Quintana M, Rauhut R, Lendeckel W and Tuschl T: Identification of novel genes coding for small expressed RNAs. Science 294: 853-858, 2001.
  99. Lau NC, Lim LP, Weinstein EG and Bartel DP: An abundant class
- 99. Lau NC, Lim LP, Weinstein EG and Bartel DP: An abundant class of tiny RNAs with probable regulatory roles in Caenorhabditis elegans. Science 294: 858-862, 2001.
- 100. Lee RC and Ambros V: An extensive class of small RNAs in Caenorhabditis elegans. Science 294: 862-864, 2001.
- 101. Pfeffer SR, Grossmann KF, Cassidy PB, Yang CH, Fan M, Kopelovich L, Leachman SA and Pfeffer LM: Detection of exosomal miRNAs in the plasma of melanoma patients. J Clin Med Res 4: 2012-2027, 2015.
- 102. Lin N, Zhou Y, Lian X and Tu Y: Expression of microRNA-106b and its clinical significance in cutaneous melanoma. Genet Mol Res 14: 16379-16385, 2015.
- 103. Friedman EB, Shang S, de Miera EVS, Fog JU, Teilum MW, Ma MW, Berman RS, Shapiro RL, Pavlick AC, Hernando E, *et al*: Serum microRNAs as biomarkers for recurrence in melanoma. J Transl Med 10: 155, 2012.
- 104. Wróblewska JP, Lach MS, Ustaszewski A, Kulcenty K, Ibbs M, Jagiełło I, Suchorska WM and Marszałek A: The potential role of selected miRNA in uveal melanoma primary tumors as early biomarkers of disease progression. Genes (Basel) 11: 271, 2020.
- 105. Tsao SCH, Weiss J, Hudson C, Christophi C, Cebon J, Behren A and Dobrovic A: Monitoring response to therapy in melanoma by quantifying circulating tumour DNA with droplet digital PCR for BRAF and NRAS mutations. Sci Rep 5: 11198, 2015.
- 106. Girotti MR, Gremel G, Lee R, Galvani E, Rothwell D, Viros A, Mandal AK, Lim KH, Saturno G, Furney SJ, *et al*: Application of sequencing, liquid biopsies, and patient-derived xenografts for personalized medicine in melanoma. Cancer Discov 6: 286-299, 2016.
- 107. Clark WH Jr, Elder DE, Guerry D IV, Braitman LE, Trock BJ, Schultz D, Synnestvedt M and Halpern AC: Model predicting survival in stage I melanoma based on tumor progression. J Natl Cancer Inst 81: 1893-1904, 1989.
- 108. Clemente CG, Mihm MC Jr, Bufalino R, Zurrida S, Collini P and Cascinelli N: Prognostic value of tumor infiltrating lymphocytes in the vertical growth phase of primary cutaneous melanoma. Cancer 77: 1303-1310, 1996.
- 109. Mandalà M and Massi D: Tissue prognostic biomarkers in primary cutaneous melanoma. Virchows Arch 464: 265-281, 2014.
- 110. Balch CM, Murad TM, Soong SJ, Ingalls AL, Halpern NB and Maddox WA: A multifactorial analysis of melanoma: Prognostic histopathological features comparing Clark's and Breslow's staging methods. Ann Surg 188: 732-742, 1978.
- 111. Lattanzi M, Lee Y, Simpson D, Moran U, Darvishian F, Kim RH, Hernando E, Polsky D, Hanniford D, Shapiro R, *et al*: Primary melanoma histologic subtype: Impact on survival and response to therapy. J Natl Cancer Inst 111: 180-188, 2019.
- 112. Robinson E, Kulkarni PM, Pradhan JS, Gartrell RD, Yang C, Acs B, Rohr B, Phelps R, Ferringer T, Horst B, *et al*: Prediction of distant melanoma recurrence from primary tumor digital H&E images using deep learning. J Clin Orthod 37 (15 Suppl): S9577, 2019.
- 113. Lehmann JM, Holzmann B, Breitbart EW, Schmiegelow P, Riethmüller G and Johnson JP: Discrimination between benign and malignant cells of melanocytic lineage by two novel antigens, a glycoprotein with a molecular weight of 113,000 and a protein with a molecular weight of 76,000. Cancer Res 47: 841-845, 1987.

- 114. Lei X, Guan CW, Song Y and Wang H: The multifaceted role of CD146/MCAM in the promotion of melanoma progression. Cancer Cell Int 15: 3, 2015.
- 115. Pacifico MD, Grover R, Richman PI, Daley FM, Buffa F and Wilson GD: Development of a tissue array for primary melanoma with long-term follow-up: Discovering melanoma cell adhesion molecule as an important prognostic marker. Plast Reconstr Surg 115: 367-375, 2005.
- 116. Weinstein D, Leininger J, Hamby C and Safai B: Diagnostic and prognostic biomarkers in melanoma. J Clin Aesthet Dermatol 7: 13-24, 2014.
- 117. Gimotty PA, Van Belle P, Elder DE, Murry T, Montone KT, Xu X, Hotz S, Raines S, Ming ME, Wahl P and Guerry D: Biologic and prognostic significance of dermal Ki67 expression, mitoses, and tumorigenicity in thin invasive cutaneous melanoma. J Clin Oncol 23: 8048-8056, 2005.
- 118. Ladstein RG, Bachmann IM, Straume O and Akslen LA: Ki-67 expression is superior to mitotic count and novel proliferation markers PHH3, MCM4 and mitosin as a prognostic factor in thick cutaneous melanoma. BMC Cancer 10: 140, 2010.
- 119. Tu TJ, Ma MW, Monni S, Rose AE, Yee H, Darvishian F, Polsky D, Berman RS, Shapiro RL, Pavlick AC, *et al*: A high proliferative index of recurrent melanoma is associated with worse survival. Oncology 80: 181-187, 2011.
- 120. Kahn HJ, Bailey D and Marks A: Monoclonal antibody D2-40, a new marker of lymphatic endothelium, reacts with Kaposi's sarcoma and a subset of angiosarcomas. Mod Pathol 15: 434-440, 2002.
- 121. Kahn HJ and Marks A: A new monoclonal antibody, D2-40, for detection of lymphatic invasion in primary tumors. Lab Invest 82: 1255-1257, 2002.
- 122. Niakosari F, Kahn HJ, McCready D, Ghazarian D, Rotstein LE, Marks A, Kiss A and From L: Lymphatic invasion identified by monoclonal antibody D2-40, younger age, and ulceration: Predictors of sentinel lymph node involvement in primary cutaneous melanoma. Arch Dermatol 144: 462-467, 2008.
- 123. Rittling SR and Chambers AF: Role of osteopontin in tumour progression. Br J Cancer 90: 1877-1881, 2004.
- 124. Rudland PS, Platt-Higgins A, El-Tanani M, Silva Rudland S, Barraclough R, Winstanley JH, Howitt R and West CR: Prognostic significance of the metastasis-associated protein osteopontin in human breast cancer. Cancer Res 62: 3417-3427, 2002.
- 125. Pan HW, Ou YH, Peng SY, Liu SH, Lai PL, Lee PH, Sheu JC, Chen CL and Hsu HC: Overexpression of osteopontin is associated with intrahepatic metastasis, early recurrence, and poorer prognosis of surgically resected hepatocellular carcinoma. Cancer 98: 119-127, 2003.
- 126. Rangel J, Nosrati M, Torabian S, Shaikh L, Leong SP, Haqq C, Miller JR II, Sagebiel RW and Kashani-Sabet M: Osteopontin as a molecular prognostic marker for melanoma. Cancer 112: 144-150, 2008.
- 127. Thomas NE, Edmiston SN, Alexander A, Groben PA, Parrish E, Kricker A, Armstrong BK, Anton-Culver H, Gruber SB, From L, *et al*: Association between NRAS and BRAF mutational status and melanoma-specific survival among patients with higher-risk primary melanoma. JAMA Oncol 1: 359-368, 2015.
- 128. Cirenajwis H, Lauss M, Ekedahl H, Törngren T, Kvist A, Saal LH, Olsson H, Staaf J, Carneiro A, Ingvar C, et al: NF1-mutated melanoma tumors harbor distinct clinical and biological characteristics. Mol Oncol 11: 438-451, 2017.
- 129. Schumacher TN and Schreiber RD: Neoantigens in cancer immunotherapy. Science 348: 69-74, 2015.
  130. Van Allen EM, Miao D, Schilling B, Shukla SA,
- 130. Van Allen EM, Miao D, Schilling B, Shukla SA, Blank C, Zimmer L, Sucker A, Hillen U, Foppen MHG, Goldinger SM, *et al*: Genomic correlates of response to CTLA-4 blockade in metastatic melanoma. Science 350: 207-211, 2015.
- 131. Snyder A, Makarov V, Merghoub T, Yuan J, Zaretsky JM, Desrichard A, Walsh LA, Postow MA, Wong P, Ho TS, *et al*: Genetic basis for clinical response to CTLA-4 blockade in melanoma. N Engl J Med 371: 2189-2199, 2014.
- 132. Samstein RM, Lee CH, Shoushtari AN, Hellmann MD, Shen R, Janjigian YY, Barron DA, Zehir A, Jordan EJ, Omuro A, *et al*: Tumor mutational load predicts survival after immunotherapy across multiple cancer types. Nat Genet 51: 202-206, 2019.
- 133. Johnson DB, Lovly CM, Flavin M, Panageas KS, Ayers GD, Zhao Z, Iams WT, Colgan M, DeNoble S, Terry CR, et al: Impact of NRAS mutations for patients with advanced melanoma treated with immune therapies. Cancer Immunol Res 3: 288-295, 2015.

- 134. Johnson DB, Bordeaux J, Kim JY, Vaupel C, Rimm DL, Ho TH, Joseph RW, Daud AI, Conry RM, Gaughan EM, et al: Quantitative spatial profiling of PD-1/PD-L1 interaction and HLA-DR/IDO-1 predicts improved outcomes of anti-PD-1 therapies in metastatic melanoma. Clin Cancer Res 24: 5250-2560, 2018.
- 135. Johnson DB, Estrada MV, Salgado R, Sanchez V, Doxie DB, Opalenik SR, Vilgelm AE, Feld E, Johnson AS, Greenplate AR, et al: Melanoma-specific MHC-II expression represents a tumour-autonomous phenotype and predicts response to anti-PD-1/PD-L1 therapy. Nat Commun 7: 10582, 2016.
- 136. Rodig SJ, Gusenleitner D, Jackson DG, Gjini E, Giobbie-Hurder A, Jin C, Chang H, Lovitch SB, Horak C, Weber JS, *et al*: MHC proteins confer differential sensitivity to CTLA-4 and PD-1 blockade in untreated metastatic melanoma. Sci Transl Med 10: eaar3342, 2018.
- Chowell D, Morris LGT, Grigg CM, Weber JK, Samstein RM, Makarov V, Kuo F, Kendall SM, Requena D, Riaz N, *et al*: Patient HLA class I genotype influences cancer response to checkpoint blockade immunotherapy. Science 359: 582-587, 2018.
   Sanlorenzo M, Vujic I, Floris A, Novelli M, Gammaitoni L,
- 138. Sanlorenzo M, Vujic I, Floris A, Novelli M, Gammaitoni L, Giraudo L, Macagno M, Leuci V, Rotolo R, Donini C, et al: BRAF and MEK inhibitors increase PD-1-positive melanoma cells leading to a potential lymphocyte-independent synergism with anti-PD-1 antibody. Clin Cancer Res 24: 3377-3385, 2018.
- 139. Eggermont AMM, Blank CU, Mandala M, Long GV, Atkinson V, Dalle S, Haydon A, Lichinitser M, Khattak A, Carlino MS, *et al*: Adjuvant pembrolizumab versus placebo in resected stage III melanoma. N Engl J Med 378: 1789-1801, 2018.
- 140. Rimm DL, Han G, Taube JM, Yi ES, Bridge JA, Flieder DB, Homer R, West WW, Wu H, Roden AC, *et al*: A prospective, multi-institutional, pathologist-based assessment of 4 immunohistochemistry assays for PD-L1 expression in non-small cell lung cancer. JAMA Oncol 3: 1051-1058, 2017.
- 141. Rizk EM, Gartrell RD, Barker LW, Esancy CL, Finkel GG, Bordbar DD and Saenger YM: Prognostic and predictive immunohistochemistry-based biomarkers in cancer and immunotherapy. Hematol Oncol Clin North Am 33: 291-299, 2019.
- 142. Harel M, Ortenberg R, Varanasi SK, Mangalhara KC, Mardamshina M, Markovits E, Baruch EN, Tripple V, Arama-Chayoth M, Greenberg E, *et al*: Proteomics of melanoma response to immunotherapy reveals mitochondrial dependence. Cell 179: 236-250.e18, 2019.
- 143. Zhou X, Yu S, Zhao DM, Harty JT, Badovinac VP and Xue HH: Differentiation and persistence of memory CD8(+) T cells depend on T cell factor 1. Immunity 33: 229-240, 2010.
- 144. Kratchmarov R, Magun AM and Reiner SL: TCF1 expression marks self-renewing human CD8<sup>+</sup> T cells. Blood Adv 2: 1685-1690, 2018.
- 145. Im SJ, Hashimoto M, Gerner MY, Lee J, Kissick HT, Burger MC, Shan Q, Hale JS, Lee J, Nasti TH, *et al*: Defining CD8+ T cells that provide the proliferative burst after PD-1 therapy. Nature 537: 417-421, 2016.
- 146. Sade-Feldman M, Yizhak K, Bjorgaard SL, Ray JP, de Boer CG, Jenkins RW, Lieb DJ, Chen JH, Frederick DT, Barzily-Rokni M, *et al*: Defining T cell states associated with response to checkpoint immunotherapy in melanoma. Cell 175: 998-1013.e20, 2018.
  147. Hugo W, Zaretsky JM, Sun L, Song C, Moreno BH,
- 147. Hugo W, Zaretsky JM, Sun L, Song C, Moreno BH, Hu-Lieskovan S, Berent-Maoz B, Pang J, Chmielowski B, Cherry G, et al: Genomic and transcriptomic features of response to anti-PD-1 therapy in metastatic melanoma. Cell 165: 35-44, 2016.
- 148. Ott PÁ, Bang YJ, Piha-Paul SA, Razak ARA, Bennouna J, Soria JC, Rugo HS, Cohen RB, O'Neil BH, Mehnert JM, *et al*: T-cell-inflamed gene-expression profile, programmed death ligand 1 expression, and tumor mutational burden predict efficacy in patients treated with pembrolizumab across 20 cancers: KEYNOTE-028. J Clin Oncol 37: 318-327, 2019.
- 149. Ayers M, Lunceford J, Nebozhyn M, Murphy E, Loboda A, Kaufman DR, Albright A, Cheng JD, Kang SP, Shankaran V, *et al*: IFN-γ-related mRNA profile predicts clinical response to PD-1 blockade. J Clin Invest 127: 2930-2940, 2017.
- 150. Pinato DJ, Howlett S, Ottaviani D, Urus H, Patel A, Mineo T, Brock C, Power D, Hatcher O, Falconer A, *et al*: Association of prior antibiotic treatment with survival and response to immune checkpoint inhibitor therapy in patients with cancer. JAMA Oncol 5: 1774-1778, 2019.

- 151. Yang H, Xia L, Chen J, Zhang S, Martin V, Li Q, Lin S, Chen J, Calmette J, Lu M, *et al*: Stress-glucocorticoid-TSC22D3 axis compromises therapy-induced antitumor immunity. Nat Med 25: 1428-1441, 2019.
- 152. Altan-Bonnet G and Mukherjee R: Cytokine-mediated communication: A quantitative appraisal of immune complexity. Nat Rev Immunol 19: 205-217, 2019.
- 153. Eisenring M, vom Berg J, Kristiansen G, Saller E and Becher B: IL-12 initiates tumor rejection via lymphoid tissue-inducer cells bearing the natural cytotoxicity receptor NKp46. Nat Immunol 11: 1030-1038, 2010.
- 154. Cristiani CM, Capone M, Garofalo C, Madonna G, Mallardo D, Tuffanelli M, Vanella V, Greco M, Foti DP, Viglietto G, *et al*: Altered frequencies and functions of innate lymphoid cells in melanoma patients are modulated by immune checkpoints inhibitors. Front Immunol 13: 811131, 2002.
- 155. Joshi K, Atwal D, Ravilla R, Pandey Y, Yarlagadda N, Kakadia S, Makhoul I, Hutchins L and Mahmoud F: Immunotherapy outcomes in advanced melanoma in relation to age. Perm J 24: 19.093, 2020.
- 156. Seidel JA, Otsuka A and Kabashima K: Anti-PD-1 and anti-CTLA-4 therapies in cancer: Mechanisms of action, efficacy, and limitations. Front Oncol 8: 86, 2018.
- 157. Wei SC, Duffy CR and Allison JP: Fundamental mechanisms of immune checkpoint blockade therapy. Cancer Discov 8: 1069-1086, 2018.
- 158. Jang SR, Nikita N, Banks J, Keith SW, Johnson JM, Wilson M and Lu-Yao G: Association between sex and immune checkpoint inhibitor outcomes for patients with melanoma. JAMA Netw Open 4: e2136823, 2021.
- 159. Anstadt EJ, Chu B, Yegya-Raman N, Han X, Doucette A, Poirier K, Mohiuddin JJ, Maity A, Facciabene A, Amaravadi RK, *et al*: Moderate colitis not requiring intravenous steroids is associated with improved survival in stage IV melanoma after anti-CTLA4 monotherapy, but not combination therapy. Oncologist 27: 799-808, 2022.
- 160. Jansen YJL, Rozeman EA, Mason R, Goldinger SM, Geukes Foppen MH, Hoejberg L, Schmidt H, van Thienen JV, Haanen JBAG, Tiainen L, *et al*: Discontinuation of anti-PD-1 antibody therapy in the absence of disease progression or treatment limiting toxicity: Clinical outcomes in advanced melanoma. Ann Oncol 30: 1154-1161, 2019.
- 161. Robert C, Ribas A, Hamid O, Daud A, Wolchok JD, Joshua AM, Hwu WJ, Weber JS, Gangadhar TC, Joseph RW, *et al*: Durable complete response after discontinuation of pembrolizumab in patients with metastatic melanoma. J Clin Orthod 36: 1668-1674, 2018.
- 162. Betof Warner A, Palmer JS, Shoushtari AN, Goldman DA, Panageas KS, Hayes SA, Bajwa R, Momtaz P, Callahan MK, Wolchok JD, *et al*: Long-term outcomes and responses to retreatment in patients with melanoma treated with PD-1 blockade. J Clin Oncol 38: 1655-1663, 2020.
- 163. Waterhouse DM, Garon EB, Chandler J, McCleod M, Hussein M, Jotte R, Horn L, Daniel DB, Keogh G, Creelan B, *et al*: Continuous versus 1-year fixed-duration nivolumab in previously treated advanced non-small-cell lung cancer: CheckMate 153. J Clin Orthod 38: 3863-3873, 2020.
- 164. Wang PF, Chen Y, Song SY, Wang TJ, Ji WJ, Li SW, Liu N and Yan CX: Immune-related adverse events associated with anti-PD-1/PD-L1 treatment for malignancies: A meta-analysis. Front Pharmacol 8: 730, 2017.
- 165. Santini FC, Rizvi H, Plodkowski AJ, Ni A, Lacouture ME, Gambarin-Gelwan M, Wilkins O, Panora E, Halpenny DF, Long NM, *et al*: Safety and efficacy of re-treating with immunotherapy after immune-related adverse events in patients with NSCLC. Cancer Immunol Res 6: 1093-1099, 2018.
- 166. Lebbé C, Meyer N, Mortier L, Marquez-Rodas I, Robert C, Rutkowski P, Menzies AM, Eigentler T, Ascierto PA, Smylie M, et al: Evaluation of two dosing regimens for nivolumab in combination with ipilimumab in patients with advanced melanoma: Results from the phase IIIb/IV CheckMate 511 trial. J Clin Oncol 37: 867-875, 2019.
- 167. Tarhini AA, Lee SJ, Hodi FS, Rao UNM, Cohen GI, Hamid O, Hutchins LF, Sosman JA, Kluger HM, Eroglu Z, *et al*: Phase III study of adjuvant ipilimumab (3 or 10 mg/kg) versus high-dose interferon alfa-2b for resected high-risk melanoma: North American intergroup E1609. J Clin Oncol 38: 567-575, 2020.
- 168. Betof AS, Nipp RD, Giobbie-Hurder A, Johnpulle RAN, Rubin K, Rubinstein SM, Flaherty KT, Lawrence DP, Johnson DB and Sullivan RJ: Impact of age on outcomes with immunotherapy for patients with melanoma. Oncologist 22: 963-971, 2017.

- 169. Zamami Y, Niimura T, Okada N, Koyama T, Fukushima K, Izawa-Ishizawa Y and Ishizawa K: Factors associated with immune checkpoint inhibitor-related myocarditis. JAMA Oncol 5: 1635-1637, 2019.
- 170. Tan MH, Iyengar R, Mizokami-Stout K, Yentz S, MacEachern MP, Shen LY, Redman B and Gianchandani R: Spectrum of immune checkpoint inhibitors-induced endocrinopathies in cancer patients: A scoping review of case reports. Clin Diabetes Endocrinol 5: 1, 2019.
- Wright JJ, Powers AC and Johnson DB: Endocrine toxicities of immune checkpoint inhibitors. Nat Rev Endocrinol 17: 389-399, 2021.
- 172. Minkis K, Garden BC, Wu S, Pulitzer MP and Lacouture ME: The risk of rash associated with ipilimumab in patients with cancer: A systematic review of the literature and meta-analysis. J Am Acad Dermatol 69: e121-e128, 2013.
- 173. Coleman E, Ko C, Dai F, Tomayko MM, Kluger H and Leventhal JS: Inflammatory eruptions associated with immune checkpoint inhibitor therapy: A single-institution retrospective analysis with stratification of reactions by toxicity and implications for management. J Am Acad Dermatol 80: 990-997, 2019.
- 174. Sibaud V, Meyer N, Lamant L, Vigarios E, Mazieres J and Delord JP: Dermatologic complications of anti-PD-1/PD-L1 immune checkpoint antibodies. Curr Opin Oncol 28: 254-263, 2016.
- 175. Sibaud V: Dermatologic reactions to immune checkpoint inhibitors: Skin toxicities and immunotherapy. Am J Clin Dermatol 19: 345-361, 2018.
- 176. Inno A, Metro G, Bironzo P, Grimaldi AM, Grego E, Di Nunno V, Picasso V, Massari F and Gori S: Pathogenesis, clinical manifestations and management of immune checkpoint inhibitors toxicity. Tumori 103: 405-421, 2017.
- 177. Kumar V, Chaudhary N, Garg M, Floudas CS, Soni P and Chandra AB: Current diagnosis and management of immune related adverse events (irAEs) Induced by immune checkpoint inhibitor therapy. Front Pharmacol 8: 49, 2017.
- 178. Bryce J and Boers-Doets CB: Non-rash dermatologic adverse events related to targeted therapies. Semin Oncol Nurs 30: 155-168, 2014.
- 179. Geisler AN, Phillips GS, Barrios DM, Wu J, Leung DYM, Moy AP, Kern JA and Lacouture ME: Immune checkpoint inhibitor-related dermatologic adverse events. J Am Acad Dermatol 83: 1255-1268, 2020.
- 180. Tewalt EF, Cohen JN, Rouhani SJ, Guidi CJ, Qiao H, Fahl SP, Conaway MR, Bender TP, Tung KS, Vella AT, *et al*: Lymphatic endothelial cells induce tolerance via PD-L1 and lack of costimulation leading to high-level PD-1 expression on CD8 T cells. Blood 120: 4772-4782, 2012.
- 181. Sano T, Uhara H, Mikoshiba Y, Kobayashi A, Uchiyama R, Tateishi K, Yamamoto H and Okuyama R: Nivolumab-induced organizing pneumonia in a melanoma patient. Jpn J Clin Oncol 46: 270-272, 2016.
- 182. Nakashima K, Naito T, Omori S, Yoshikawa S, Endo M, Kiyohara Y and Takahashi T: Organizing pneumonia induced by nivolumab in a patient with metastatic melanoma. J Thorac Oncol 11: 432-433, 2016.
- 183. Koelzer VH, Rothschild SI, Zihler D, Wicki A, Willi B, Willi N, Voegeli M, Cathomas G, Zippelius A and Mertz KD: Systemic inflammation in a melanoma patient treated with immune checkpoint inhibitors-an autopsy study. J Immunother Cancer 4: 13, 2016.

- 184. Watanabe S, Kimura H, Takato H, Waseda Y, Hara J, Sone T, Abo M, Maeda S, Matsushita T and Kasahara K: Severe pneumonitis after nivolumab treatment in a patient with melanoma. Allergol Int 65: 487-489, 2016.
- 185. Nishino M, Sholl LM, Hatabu H, Ramaiya NH and Hodi FS: Anti-PD-1-related pneumonitis during cancer immunotherapy. N Engl J Med 373: 288-290, 2015.
- 186. Mir MA: T-cell costimulation and its applications in diseases. Dev Costimulatory Mol Immunother Dis 29: 255-292, 2015.
- 187. Tarazona R, Duran E and Solana R: Natural killer cell recognition of melanoma: New clues for a more effective immunotherapy. Front Immunol 6: 649, 2015.
- 188. Rieth J and Subramanian S: Mechanisms of intrinsic tumor resistance to immunotherapy. Int J Mol Sci 19: 1340, 2018.
- 189. Fisher DT, Appenheimer MM and Evans SS: The two faces of IL-6 in the tumor microenvironment. Semin Immunol 26: 38-47, 2014.
- 190. O'Donnell JS, Long GV, Scolyer RA, Teng MWL and Smyth MJ: Resistance to PDI/PDL1 checkpoint inhibition. Cancer Treat Rev 52: 71-81, 2017.
- 191. Vander Heiden MG, Cantley LC and Thompson CB: Understanding the Warburg effect: The metabolic requirements of cell proliferation. Science 324: 1029-1033, 2009.
- 192. Ahn CS and Metallo CM: Mitochondria as biosynthetic factories for cancer proliferation. Cancer Metab 3: 1, 2015.
- 193. O'Donnell JS, Teng MWL and Smyth MJ: Cancer immunoediting and resistance to T cell-based immunotherapy. Nat Rev Clin Oncol 16: 151-167, 2019.
- 194. DeBerardinis RJ and Chandel NS: Fundamentals of cancer metabolism. Sci Adv 2: e1600200, 2016.
- 195. Robey IF, Lien AD, Welsh SJ, Baggett BK and Gillies RJ: Hypoxia-inducible factor-1alpha and the glycolytic phenotype in tumors. Neoplasia 7: 324-330, 2005.
- 196. Franco F, Jaccard A, Romero P, Yu YR and Ho PC: Metabolic and epigenetic regulation of T-cell exhaustion. Nat Metab 2: 1001-1012, 2020.
- 197. Jiang Y, Li Y and Zhu B: T-cell exhaustion in the tumor microenvironment. Cell Death Dis 6: e1792, 2015.
- 198. Sirico M, D'Angelo A, Gianni C, Casadei C, Merloni F and De Giorgi U: Current state and future challenges for PI3K inhibitors in cancer therapy. Cancers (Basel) 15: 703, 2023.
- 199. Peng W, Chen JQ, Liu C, Malu S, Creasy C, Tetzlaff MT, Xu C, McKenzie JA, Zhang C, Liang X, *et al*: Loss of PTEN promotes resistance to T cell-mediated immunotherapy. Cancer Discov 6: 202-216, 2016.
- 200. Li X, Wenes M, Romero P, Huang SCC, Fendt SM and Ho PC: Navigating metabolic pathways to enhance antitumour immunity and immunotherapy. Nat Rev Clin Oncol 16: 425-441, 2019.
- 201. Marin-Acevedo JA, Kimbrough EO and Lou Y: Next generation of immune checkpoint inhibitors and beyond. J Hematol Oncol 14: 45, 2021.



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