

Oncolytic potential of an E4-deficient adenovirus that can recognize the stabilization of AU-rich element containing mRNA in cancer cells

AYA YANAGAWA-MATSUDA¹, YOHEI MIKAWA^{1,2}, UMMA HABIBA¹, TETSUYA KITAMURA¹,
MOTOAKI YASUDA³, MOHAMMAD TOWFIK-ALAM^{1,4}, YOSHIMASA KITAGAWA²,
KAZUYUKI MINOWA⁴, MASANOBU SHINDOH¹ and FUMIHIRO HIGASHINO^{1,5}

Departments of ¹Oral Pathology and Biology, ²Oral Diagnosis and Medicine, ³Oral Molecular Microbiology and
⁴Dental Radiology, Hokkaido University Faculty of Dental Medicine and Graduate School of Dental Medicine,
Sapporo 060-8586; ⁵Department of Molecular Oncology, Hokkaido University Faculty of Dental Medicine
and Graduate School of Biomedical Science and Engineering, Sapporo 060-8638, Japan

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Abstract. AU-rich elements (AREs) are RNA elements that enhance the rapid decay of mRNA. The fate of ARE-mRNA is controlled by ARE-binding proteins. HuR, a member of the embryonic lethal abnormal vision (ELAV) family of RNA-binding proteins, is involved in the export and stabilization of ARE-mRNA. In the vast majority of cancer cells, HuR constitutively relocates to the cytoplasm, resulting in the stabilization of ARE-mRNA. Previously, we described that the adenovirus gene product, E4orf6, which is necessary for virus replication, participates in ARE-mRNA export and stabilization. In the present study, we showed the oncolytic potential of E4orf6-deleted adenovirus dl355, which is expected to be replicated selectively in cancer cells. Virus production and cytolytic activity of dl355 were higher in cancer cells than in normal cells. HuR-depletion downregulated dl355 replication, demonstrating that ARE-mRNA stabilization is required for the production of this virus. Tumor growth was inhibited in nude mice by an intra-tumoral injection of dl355. Furthermore, dl355 had a stronger oncolytic effect than E1B55k-deleted adenovirus. These results indicate that dl355 has potential as an oncolytic adenovirus for a large number of cancers where ARE-mRNA is stabilized.

Introduction

For adenovirus replication, several early gene products, such as E1A, E1B and E4, are necessary to change the environment

within host cells (1). The E4 region of adenoviruses encodes multiple proteins and these proteins are required for DNA replication, late gene expression and host cell shutoff (2). The largest protein encoded in the E4 region is E4orf6, which is required for productive adenovirus replication (1). E4orf6 forms a complex with another early gene product, E1B55k (3), and this complex associates with cellular proteins such as cullin 5, elongins B and C and Rbx, to form E3 ubiquitin ligase (4,5). This ligase targets p53 (4-7), the MRN complex (8), DNA ligase IV (9) and integrin $\alpha 3$ (10) for proteasomal degradation. This ubiquitin ligase activity is known to be required for the nuclear export of viral late mRNAs (11,12).

Several viral proteins encoded in the E4 region, such as E4orf1, E4orf3 and E4orf6 are known to have oncogenic activities (13-17). E4orf1 of adenovirus type 9, which belongs to adenovirus subgroup D, has been shown to be involved in mammary tumorigenesis (14,18). E4orf3 and E4orf6 proteins of subgroup C adenovirus type 5 have the potential to transform cells in cooperation with E1A and E1B proteins and enhance the growth of tumors transplanted in nude mice (15-17). In a previous report, we demonstrated that in cells transformed with adenovirus type 5 E4orf6, cellular pp32 protein associates with E4orf6 and AU-rich element (ARE)-containing mRNAs are exported to the cytoplasm in a chromosome region maintenance 1 (CRM1)-independent manner (19). Furthermore, the exported ARE-mRNAs are stabilized and those mRNAs acquire the potential to transform cells (20).

AREs usually exist in the 3'-untranslated region (UTR) of certain mRNAs encoding early response genes or growth-related genes, such as proto-oncogenes and growth factors (21,22). AREs are targets for rapid degradation of mRNA (21,23) and the fate of ARE-mRNA is controlled by several RNA-binding proteins, such as AUF1, tristetraprolin (TTP) and HuR (24). HuR, which is a member of the embryonic lethal abnormal vision (ELAV) family of RNA-binding proteins, binds to AREs in order to protect ARE-mRNA from rapid degradation (23,24). HuR is able to shuttle between the nucleus and cytoplasm, whereas it is mainly located in the

Correspondence to: Dr Fumihiko Higashino, Department of Oral Pathology and Biology, Hokkaido University Graduate School of Dental Medicine, North 13 West 7, Kita-ku, Sapporo 060-8586, Japan
E-mail: fhigashi@den.hokudai.ac.jp

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nucleus. HuR-mediated stabilization of ARE-mRNA depends on HuR localization in the cytoplasm (24,25). In normal cells, HuR transiently relocates to the cytoplasm under conditions of stress. On the other hand, HuR constitutively accumulates in the cytoplasm of cancer cells and the cytoplasmic expression of HuR is thought to be involved in malignant transformation of cancer cells (25,26).

These facts suggest that the E4orf6-deleted mutant adenovirus is able to proliferate selectively in cancer cells in which ARE-mRNA is stabilized. In the present study, we examined the oncolytic activity of the adenovirus E4orf6-deleted mutant, dl355. The ability of this virus to replicate was markedly increased in cancer cells compared with that in normal cells. dl355 showed cytolytic activity for cancer cells *in vitro* and *in vivo*. The propagation and cytolytic activity of this virus were higher than those of an E1B55k-deleted adenovirus. These findings indicate that dl355 is a potential oncolytic virus.

Materials and methods

Cell lines, viruses and antibodies. The human lung cancer cell lines, A549 and H1299; cervical carcinoma cell lines HeLa, HeLa S3 and C33A; African green monkey kidney (Vero) cells carrying an integrated copy of the Ad5 E4 region, W162; human embryonal kidney cell line 293 (transformed by the adenovirus E1 gene); human foreskin fibroblast cell line, BJ; and normal human lung primary cell line, WI38 were used in the present study. All cells were obtained from the American Type Culture Collection (ATCC; Manassas, VA, USA) and cultured in Dulbecco's modified Eagle's medium (DMEM; Sigma-Aldrich; Merck KGaA, Darmstadt, Germany) containing 10% fetal bovine serum (FBS; Biowest, Nuaille, France) with antibiotics at 37°C in a 5% CO₂ atmosphere under humidified conditions.

Wild-type adenovirus type 5 (WT300), E4orf6-deleted mutant adenovirus (dl355) (generous gift from Dr T. Shenk; Princeton University) and E1B55k-deleted mutant adenovirus (dl1520) (generous gift from Dr A.J. Berk; University of California) were used in the present study.

A western blot analysis was performed as previously described (27) using antibodies specific to E1A (M73; generous gift from Dr T. Shenk, Princeton University), E4orf6 (RSA#3; generous gift from Dr T. Shenk; Princeton University), actin (dilution 1:1,000; cat. no. sc-1616; Santa Cruz Biotechnology, Dallas, TX, USA) and HuR (dilution 1:2,500; cat. no. sc-5261; Santa Cruz Biotechnology) and β -tubulin (dilution 1:1,000; cat. no. 05-661; EMD Millipore Corp., Darmstadt, Germany) as primary antibodies. The secondary antibody was horseradish peroxidase-conjugated anti-goat IgG (dilution 1:5,000; cat. no. 805-035-180; Jackson ImmunoResearch Laboratories, West Grove, PA, USA) and horseradish peroxidase-conjugated anti-mouse IgG (dilution 1:5,000; cat. no. 115-035-062; Jackson ImmunoResearch Laboratories). Antibody binding was visualized using SuperSignal West Femto Maximum Sensitivity Substrate (Thermo Fisher Scientific, Inc., Waltham, MA, USA).

Preparation of dl355 and dl1520 lysates. To prepare virus lysates, dl355-infected W162 cells and dl1520 or WT300-infected 293 cells were subjected to three cycles of freezing and

thawing. Virus concentrations [virus particles (vp)/ml] were then determined by a QuickTiter Adenovirus Quantitation kit (Cell Biolabs, San Diego, CA, USA). Viral titers [infectious units (ifu)/ml] were determined using the Adeno-X™ Rapid Titer kit (Clontech Laboratories, Inc., Mountain View, CA, USA) according to the manufacturer's instructions. To use a virus in *in vivo* experiments, its extract was purified using a Fast-Trap Adenovirus Purification and Concentration kit (Millipore, Billerica, MA, USA) according to the manufacturer's protocols.

Cytopathic effect assay and cell viability assay. Human cancer and normal cells were plated on 24-well plates (5×10⁴ cells/well). Twenty-four hours later, the cells were infected with dl355 at a multiplicity of infection (MOI) of 0.1, 0.5, 1, 10 or 100 vp/cell and maintained for an additional 7 days. Cells were then fixed and stained with Coomassie brilliant blue.

A 2-3-bis [2-methoxy-4-nitro-5-sulphophenyl]-2H-tetrazolium-5-carboxanilide inner salt assay was used to examine cell metabolic activity. Cancer and normal cells were seeded on 96-well plates at a density of 3.0×10³ cells/well. Twenty-four hours later, the cells were infected with dl355, dl1520 or WT300 at an MOI of 100 vp/cell. Cell metabolic activity was determined using an XTT assay on days 1, 3, 5 and 7 with the Cell Proliferation kit II (Roche Diagnostics, Basel, Switzerland) according to the manufacturer's protocol.

***In vitro* virus proliferation assay.** Cancer and normal cells were seeded at 5.0×10⁴ cells/well 24 h before the infection. Cells were infected with dl355, dl1520, or WT300 at an MOI of 1 vp/cell. These cells were incubated at 37°C for 48 h, after which cells were collected and a virus lysate was prepared as described above. Viral titers (ifu/ml) were determined using the Adeno-X Rapid Titer kit (Clontech Laboratories).

HuR depletion. For RNA interference analysis, Lipofectamine RNAiMAX (Invitrogen; Thermo Fisher Scientific) was used to transfect HeLa cells with 20 nM of each small interfering RNA (siRNA) targeting HuR (5'-TTCGTAAGTTATTTCTTTAATT-3') or with a negative control siRNA (5'-TCTTAATCGCGTATAAG GCTT-3'; Qiagen, Hilden, Germany). After 48 h of transfection, HeLa cells were infected with dl355. After 24 h of infection, all cells were collected and the virus lysate was prepared using three freeze-thaw cycles. Viral titers were determined using the Adeno-X Rapid Titer kit (Clontech Laboratories) and 293 cells.

For heat shock treatment, HeLa cells were incubated at 43°C for 2 h immediately after dl355 infection, and the infected cells were heat shocked (2 h) again at 24 h after infection. Cells were harvested at 28 h after infection and the viral titers were determined as described above.

***In vivo* human tumor model.** Female BALB/c nu/nu mice (purchased from Hokudo, Sapporo, Japan) were housed under specific pathogen-free conditions. The temperature was 26–28°C, and the light/dark cycle was 10-h/14-h cycle, food and water were taken *ad libitum*. HeLa S3 cells (1.0×10⁶ cells/mouse) were injected subcutaneously into the flanks of mice (5 week old and 20–24 g) and permitted to grow to ~5–6 mm in diameter. The mice were randomly

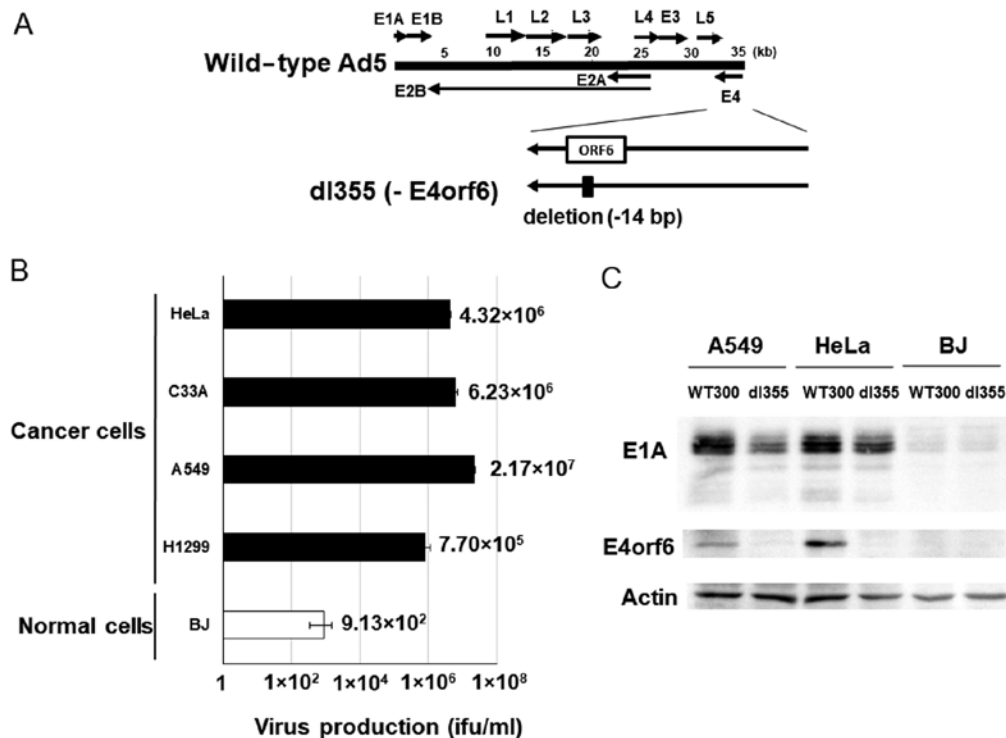


Figure 1. The structure of dl355 and its production efficiency. (A) Schematic representation of dl355 with a 14 base-pair (bp) deletion in the open reading frame 6 gene of the wild-type adenovirus type 5 (Ad5) E4 region. Early (E1-4) and late (L1-5) genes are indicated by arrows. (B) Cancer (HeLa, C33A, A549 and H1299) and normal (BJ) cells were infected with dl355 at an MOI of 1 vp/cells and virus production was determined 48 h after infection by hexon staining. Each titer (ifu/ml) is indicated on the graph. Data are shown as the mean \pm standard deviation of three independent experiments. (C) Expression of virus protein in dl355 and WT300- infected HeLa cells. Both viruses were infected with A549, HeLa and BJ cells, then the expression of E1A, E4orf6 and actin were estimated by western blot analysis. MOI, multiplicity of infection; ifu, infectious units; WT300, wild-type adenovirus type 5; dl355, E4orf6-deleted mutant adenovirus.

divided into two groups (5 per group) and 10^9 vp (100 μ l) of dl355 or the same volume of PBS was injected twice (days 0 and 3) directly into the tumors. The perpendicular diameters of the tumors were measured every 3 or 4 days and tumor volumes were calculated using the following equation: Volume (mm^3) = $A \times B^2 \times 0.5$ (A is the longest diameter, B is the shortest diameter). The mice were sacrificed by cervical dislocation after 30 days of injection of virus. All procedures performed in this study involving animals were in accordance with the ethical standards of the Animal Care and Use Committee of the Hokkaido University (Sapporo, Japan).

Results

Selective dl355 replication in cancer cells. dl355 (Fig. 1A) has a 14-bp deletion in the *E4orf6* gene and was constructed in 1985 to investigate the functions of genes located in the E4 region in adenovirus-infected host cells. This virus showed deficient virus DNA replication, accumulation of late viral mRNAs, and shutoff of host cell mRNAs compared to wild-type adenovirus type 5 (Ad5) (2). To examine the productive efficiency of dl355, cancer cells (HeLa, C33A, A549 and H1299) and normal cells (BJ) were infected with dl355 at an MOI of 1 virus particle (vp)/cell and virus titers generated after 48 h were detected by staining the hexon protein of virus particles in 293 cells. In these cancer cells, the propagation of dl355 was very high, with titers from 7.70×10^5 to 2.17×10^7 ifu/ml. On the other hand, the titer of dl355 in

normal cells (BJ; foreskin fibroblasts) was 3 to 5 logs lower (9.13×10^2 ifu/ml) than in cancer cells (Fig. 1B). We examined the expression of E1A protein, which is expressed first after infection. The amount of E1A protein was at a high level in dl355-infected A549 and HeLa cells, although the level was low in normal BJ cells (Fig. 1C). These results suggest that dl355 is selectively produced in cancer cells.

Since dl355 was thought to propagate in cancer cells in which ARE-mRNA is stabilized, we examined whether the ARE-mRNA stabilization system was required for dl355 replication. To evaluate this, we confirmed adenovirus production in HuR-depleted cells, because decreased HuR inhibits ARE-mRNA stabilization (28). Heat shock (HS) treatment is known to downregulate HuR by ubiquitin-mediated proteolysis and HuR-targeted mRNA is also decreased in HS-treated cells (29). If dl355 replicates using the ARE-mRNA stabilization system, the virus titer is expected to reduce with HS treatment. HeLa cells were subjected to HS, as described in Materials and methods, and were then examined for HuR protein and virus production. As expected, a 2-h HS treatment of HeLa cells resulted in reduced expression of HuR protein in the cytoplasm of HeLa cells (Fig. 2A). Furthermore, HS-treated cells showed a significant reduction in virus production compared to the non-treated cells ($\sim 1/28.6$; Fig. 2B).

Since HS treatment affects have many influences other than HuR proteolysis in cells, we confirmed the HuR-depletion effect by HuR knockdown (KD). The virus production in siRNA for HuR introduced cells (Fig. 2C) was much less

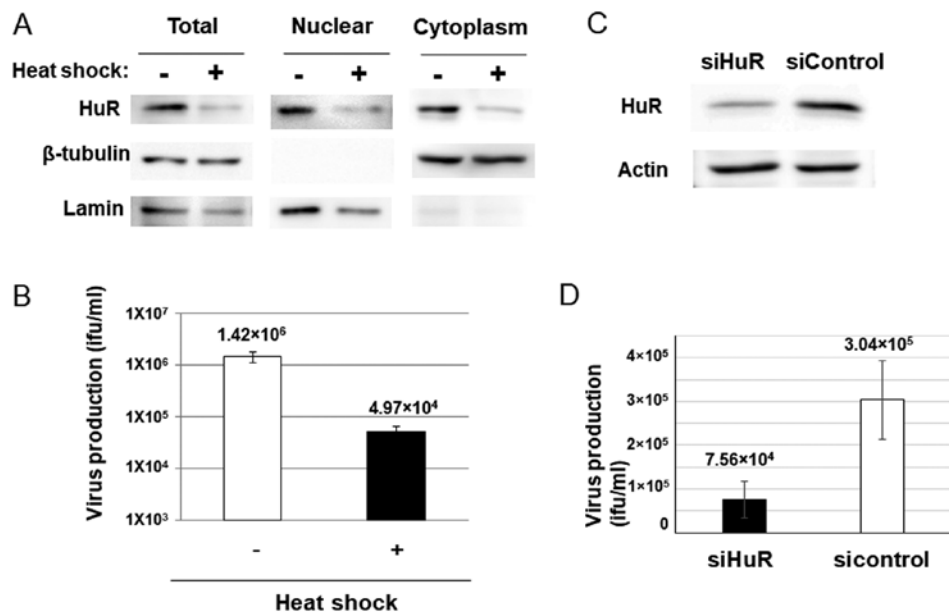


Figure 2. Effect of heat shock and HuR knockdown (KD) on dl355 propagation. (A) HeLa cells were heat shocked at 43°C for 2 h and the amount of HuR in the total, nuclear and cytoplasmic fraction was determined using western blotting. β-tubulin and lamin expression was also determined as the markers of the cytoplasm and nuclear fractions. (B) HeLa cells were heat shocked at 43°C for 2 h immediately after WT300 infection and the infected cells were heat shocked again 24 h after the infection. Virus production was then evaluated and the values were compared to the non-treated cells. Data are shown as the mean ± standard deviation of three independent experiments. (C) siRNA targeting HuR and a negative control siRNA were introduced into HeLa cells then the expression of HuR was estimated by western blot analysis. (D) HuR KD-HeLa cells were infected with dl355 and viral titers were determined as described in Materials and methods after 24 h of infection. ifu, infectious units; WT300, wild-type adenovirus type 5; dl355, E4orf6-deleted mutant adenovirus.

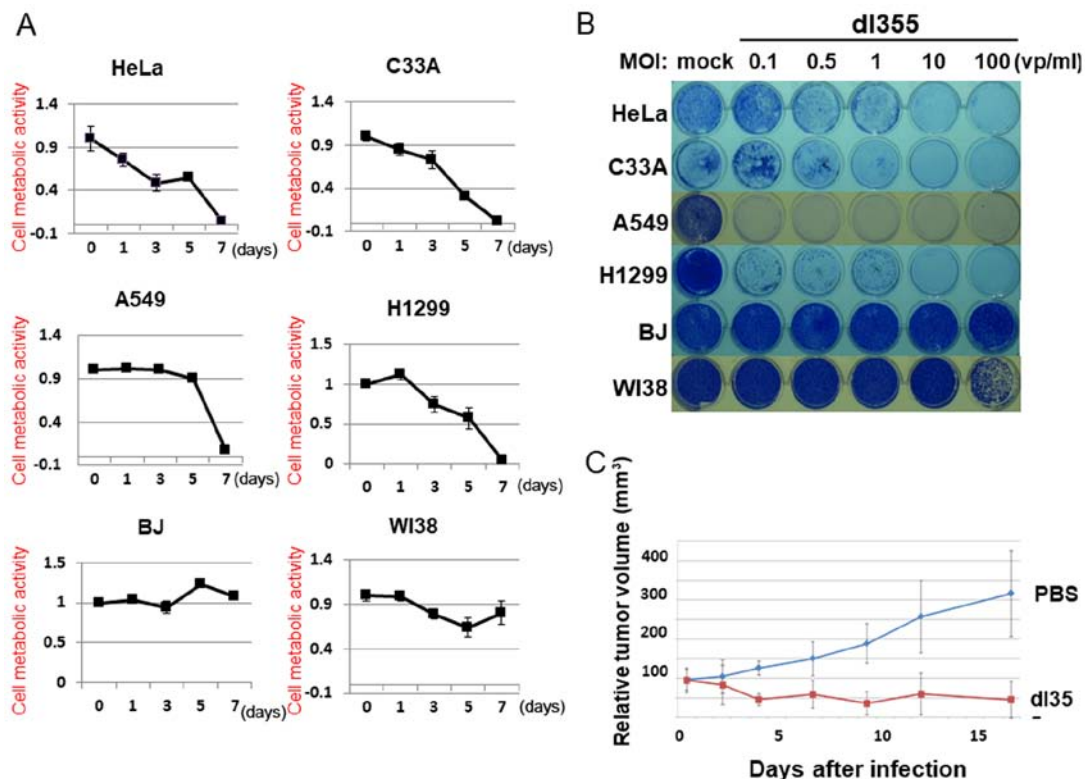


Figure 3. *In vitro* and *in vivo* cell lysis activity of dl355. (A) Cell metabolic activities of dl355-infected cells was measured using the XTT assay. Cancer (HeLa, A549, C33A and H1299) and normal (BJ and WI38) cells were infected with the virus at an MOI of 100 vp/cell and cell viabilities were estimated 1, 3, 5 and 7 days after infection. (B) The same cancer and normal cells were infected with dl355 at the MOIs indicated. Cells were stained with Coomassie brilliant blue 7 days after infection. Living cells stained blue. Data are shown as the mean ± standard deviation of three independent experiments. (C) *In vivo* antitumor effects of intratumorally injected dl355 were evaluated in HeLa xenograft nude mice. HeLa S3 cells were injected subcutaneously into nude mice to form a tumor with a diameter of ~5 mm (after ~3 weeks). dl355 [10^9 vp (100 μl)] was injected intratumorally into the tumor on days 0 and 3. The same volume of PBS was injected as a control. At least five mice were used for each group. Tumor volumes were measured twice a week and the results obtained are shown as the mean of relative volumes ± standard deviation. MOI, multiplicity of infection; vp, virus particles; WT300, wild-type adenovirus type 5; dl355, E4orf6-deleted mutant adenovirus; PBS, phosphate-buffered saline.

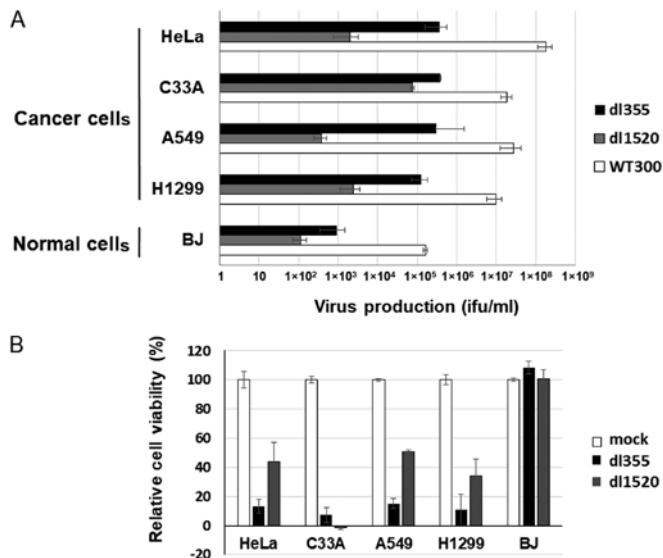


Figure 4. Comparison of virus production and cell lysis activity between dl355 and dl1520. (A) Cancer (HeLa, C33A, A549 and H1299) and normal (BJ) cells were infected with dl355, dl1520, or WT300 at an MOI of 1 vp/cell and virus production was determined as described in Materials and methods. Each titer (vp/ml) is indicated on the graph. Data are shown as the mean \pm standard deviation of three independent experiments. (B) The same cancer and normal cells were infected with dl355 or dl1520 at an MOI of 100 vp/cell and cell viabilities were estimated 7 days after infection. Data are shown as the mean \pm standard deviation of three independent experiments. WT300, wild-type adenovirus type 5; dl355, E4orf6-deleted mutant adenovirus; dl1520, E1B55k-deleted mutant adenovirus; MOI, multiplicity of infection; vp, virus particles; ifu, infectious units.

than that of the cells with control siRNA transfected cells (~1/4; Fig. 2D). These results suggest that HuR plays an important role in dl355 replication.

In vitro and in vivo cytolytic potential of dl355. To estimate the cell lysis activity of dl355, we examined cell viability using the XTT assay. HeLa, A549, C33A, H1299, BJ and WI38 cells were infected with dl355 at an MOI of 100 (vp/cells) and the XTT assay was performed 1, 3, 5 and 7 days after infection (Fig. 3A). In the case of HeLa cells, dl355 infection resulted in reduced cell viability 3 days after infection and most cells died after 7 days. HeLa cells showed the earliest effects, but in other cancer cells, nearly all cells died on day 7. In contrast, most normal cells infected with dl355 did not die even after 7 days.

To assess the cell lysis activity of dl355 further, we examined the infected cells using a cytopathic effects (CPE) assay. Four different cancer cell types (HeLa, C33A, A549 and H1299) and two normal cell types (BJ and WI38) were infected with dl355 at MOIs of 0.1, 0.5, 1, 10 or 100. Cytotoxicity was estimated by staining the remaining cells with Coomassie brilliant blue 7 days after infection (Fig. 3B). Although several cell lines survived with low MOIs, all cancer cells were killed by dl355 in a dose-dependent manner. On the other hand, most normal cells survived in all infections. Taken together, these results demonstrate the *in vitro*-selective cytolytic activity of dl355.

We estimated the therapeutic effect of dl355 for human cancers using a tumor xenograft model. In order to examine this, HeLa S3 cells were implanted into the hind flanks of 5-week-old female BALB/c nu/nu mice. When the tumors

grew to ~5-6 mm in diameter, 10⁹ vp dl355 (100 μ l) or the same volume of phosphate-buffered saline (PBS) (as a control) were injected twice (days 0 and 3) directly into the tumor. Tumor growth was significantly suppressed by injection with dl355 whereas the PBS-injected tumors grew almost three times as large in 18 days (Fig. 3C). Thus, dl355 exerted significant effects on human tumor xenografts in nude mice.

Comparison of the oncolytic effects of dl355 and dl1520. Since an E1B-55k gene deleted-adenovirus has been developed as an oncolytic adenovirus (30) and is already applied clinically, we next compared the oncolytic activity of dl355 with that of the E1B-55k-deleted adenovirus, dl1520. Using cancer cells, HeLa, C33A, A549 and H1299, and normal BJ cells, dl355, dl1520 and WT300 as a control of infectivity were infected at an MOI of 1 and virus titers were estimated 48 h after infection. Since different cell lines (293 and W162 cells) were used to determine the virus production rate (ifu/ml) of each virus, we compared the productive efficiencies of both viruses with viral particles (vp/ml). As shown in Fig. 4A, dl355 virus production was significantly higher (1-3 logs higher) than dl1520 virus production in all cancer cells.

To compare cell lysis activity, both viruses (MOI 100) were infected into the same cancer and normal cells and the XTT assay was performed 7 days after infection. Except for C33A case, dl355 showed stronger cell death activity than dl1520 in cancer cells (Fig. 4B). However, in normal cells, both viruses showed almost no cell death effect. Taken together, these data indicate that dl355 has a stronger oncolytic effect than dl1520 in at least several cancer cell lines.

Discussion

In the present study, we described the oncolytic potential of the E4orf6-deleted adenovirus, dl355. The productive efficiencies of this virus with cancer cells were approximately 10² to 10³ times higher from that of normal cells (Fig. 1B). It showed a high level of cytolytic activity for cervical and lung cancer cells *in vitro* and the same effect was evident in a tumor xenograft model. Furthermore, dl355 has oncolytic activity that may be superior to dl1520, which is currently used clinically. These findings indicate that dl355 is a potential oncolytic virus.

Many types of conditionally replicative adenoviruses (CRAds) targeted to cancer cells are being developed and several viruses are currently in clinical trials (31). Oncolytic adenoviruses can be divided into at least two types (32), one of which has mutations in genes required for viral replication. For example, the *E1A* or *E1B-55k* gene deleted-virus has been developed as a CRAd and it has oncolytic effects for pRB- or p53-deficient tumor cells (30,33). The other group consists of viruses that possess a cancer-specific transcription system in the virus genes required for replication such as *E1A*. For example, promoters of the telomerase gene (34) or prostate-specific antigen gene (35) are inserted into the 5'-untranslated region (UTR) of the *E1A* gene to produce CRAds, which are specifically activated in cancer cells. There are few reports describing oncolytic viruses with tumor selectivity based on the level of mRNA stability. Thus, dl355 is a rare type of oncolytic virus that its replication is controlled by RNA modulation.

As we have shown in a previous study (28), HuR KD attenuates the export and stabilization of ARE-mRNA. This is due to the fact that HuR is the only protein that binds directly to ARE, so the ARE-mRNA failed to be exported to the cytoplasm if HuR disappears. In this study, virus proliferation was downregulated under HuR HS or HuR KD conditions. These results suggest that the export and stabilization of ARE-mRNA is essential for the growth of dl355. In a previous study (36), it was clarified that various stimuli promote the relocalization of HuR to the cytoplasm and the stabilization of ARE-mRNA. Therefore, if stimulation is added to enhance export and stabilization of ARE-mRNA, the production efficiency of dl355 increases, indicating the possibility of obtaining a stronger tumor lysis activity.

We used various types of cancer cells in this study and it is expected that the infection efficiency of the virus was different in each cell line. We examined the expression of E1A protein, which is expressed first after infection, to evaluate the infectivity of dl355. As the amount of E1A protein was almost the same between dl355-infected A549 and HeLa cells (Fig. 1C), the infectivity can be considered to be equivalent. Furthermore, as shown in Fig. 4A, the virus production of the WT300 infected in different types of cancer cells was not significantly different, thus the infection efficiency was not so different. These data showed the validity of comparing the production efficiency of dl355 and dl1520.

In the vast majority of cancer cells, ARE-mRNAs relocate to the cytoplasm with HuR and are constitutively stabilized (26). We showed that dl355 replication depends on the ARE-mRNA stabilization system, since the replication of dl355 is downregulated in HuR-depleted cancer cells (Fig. 2). These facts suggest that dl355 has the potential to be effective for many types of cancer cells.

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Availability of data and materials

The datasets used during the present study are available from the corresponding author upon reasonable request.

Authors' contributions

FH, AYM and YM conceived and designed the research and contributed to the writing of the paper. AYM and YM mainly conducted the research. UH, TK and MTA conducted the *in vivo* analysis. MY and UH contributed to the production of the virus. FH, MY, YK, KM and MS assisted with statistical

analysis and also analyzed the data. TK, YK and MS proofread the paper. KM and MTA revised the manuscript. All authors read and approved the manuscript and agree to be accountable for all aspects of the research in ensuring that the accuracy or integrity of any part of the study are appropriately investigated and resolved.

Ethics approval and consent to participate

All the animal experiments performed in this study were in accordance with the ethical standards of the Animal Care and Use Committee of Hokkaido University (Sapporo, Hokkaido, Japan).

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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