

Involvement of calmodulin and actin in directed differentiation of rat cortical neural stem cells into neurons

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Abstract. Calmodulin (CaM) is a multifunctional receptor of intercellular Ca^{2+} , which under different physiological conditions or at different developmental stages plays different roles in different tissues and cells. This study aimed to investigate the involvement of spatial expression and coexistence of CaM and actin in directed differentiation of rat cortical neural stem cells (NSCs) into neurons. Immunohistochemistry, RT-PCR and immunofluorescence dual-labeling technology was conducted to investigate the temporal and spatial pattern of CaM and actin proteins in neuron-oriented cortical NSC differentiation. Confocal laser scanning microscopy (CLSM) was used to observe changes of the coexistence in protein expression. Our results showed that cortical NSCs expressed and showed localized CaM and actin in a well-defined temporal order. In the process of rat neuron-oriented cerebral cortical NSC differentiation, CaM displayed a similar expression pattern with actin in the development of neurons, that is, both proteins extended into neurites following the sprouting and growth of neurons. These results suggest that the temporal and spatial pattern of CaM and actin expression is comparable to the growth of cell processes in differentiating NSCs. Therefore, both CaM and actin may jointly participate in the development and maturation of neurites, and this provides a theoretical basis for further study of the biological features of neuron-oriented NSC differentiation.

Introduction

As a multi-functional receptor of intercellular Ca^{2+} , calmodulin (CaM) mediates many physiological processes regulated by

Ca^{2+} . CaM is extensively localized in almost all tissues of vertebrates, plants and prokaryotes. This protein is especially rich in the brain and some endocrine organs (1). CaM has been shown to play different roles in different tissues and cells, under different physiological conditions or at different developmental stages. In addition, actin is a ubiquitous cytoskeletal protein, which is mainly involved in the formation of microfilaments and maintains normal cellular functions with the assistance of intermediate filaments and microtubules (2-4). As a cytoskeletal protein, actin plays an important role in cell growth and development, which not only closely correlates with cell morphology, but also participates in multiple functions such as cell movement, substance transport, as well as cell division, differentiation and gene expression (3,5).

It has been shown that alteration of the intracellular Ca^{2+} concentration is closely related to the function of actin (6). Therefore, CaM, a Ca^{2+} receptor should also affect the function of actin (7). However, it is not clear whether this correlation exists in the directed differentiation of rat cerebral cortical neural stem cells (NSCs) into neurons. In this study, immunohistochemistry, RT-PCR and image analysis technology were conducted to detect the spatial expression of CaM and its action during the process of rat neuron-oriented cerebral cortical NSC differentiation. Immunofluorescence dual-labeling technology was used to label CaM and actin. Confocal laser scanning microscopy (CLSM) was used to observe whether there is coexistence in the protein expression and correlation between CaM and actin. The main objective of this study was to provide the theoretical basis for further study of the biological characteristics of neuron-oriented directed differentiation of NSCs.

Materials and methods

Experimental animals. Wistar rats born within 24 h were provided by the Department of Laboratory Animals of the China Medical University. All procedures were approved by the Institutional Animal Care and Use Committee of the China Medical University, Shenyang, China.

Culture and differentiation of NSCs. Cerebral cortices from neonatal rats were taken out in a sterilized environment, placed in Hank's solution, rinsed, cut into small pieces and

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transferred into a centrifuge tube; 0.25% trypsin was added followed by incubation at 37°C for 25 min to digest; then DMEM/F12 serum-free medium was added to terminate the digestion. Mechanical force was used to disperse the cells which were then filtered by 4 layers of sterile gauze; cells were then centrifuged at 1,600 rpm/min for 5 min; the supernatant was discarded and DMEM/F12 serum-free medium containing 20 ng/ml bFGF, 20 ng/ml EGF, 1% N₂ supplement (all from Gibco, USA), 10,000 U/ml penicillin, 10,000 U/ml streptomycin (North China Pharmaceutical Company, Ltd., China) and 0.0014 ml/l diflucan (Pfizer, France) was added. Trypan blue staining was used to evaluate cell viability and the cell density in suspension was adjusted to 1-5x10⁶/ml, plated in 75 ml culture flasks and placed at 37°C in a 5% CO₂ incubator. The culture medium was changed by half every two days. The cell morphology was observed using an inverted phase contrast microscope and the cell images were captured. Neurospheres formed after 7 days of primary culture with culture medium were collected into a centrifuge tube, centrifuged at 400 rpm for 5 min and resuspended into a single cell suspension by the above culture medium. Cells were again plated into culture flasks at the same density. Cells were passaged for a total of 4 generations. To induce differentiation of NSCs, a single cell suspension of NSCs was plated onto poly-L-ornithine coated dishes at a density of 2x10⁴ cell/ml in the basic neurosphere medium with 10% FCS and 2 ng/ml BDNF and cultured for 7 days.

Histological analysis. Immunohistochemistry was performed to identify NSCs and neuron-oriented differentiation of NSCs. After NSCs were cultured for 7 days, the actively growing neurosphere suspension was dropped onto polylysine-treated coverslips and air-dried. To assess the expression of CaM and actin, the coverslips were taken out of culture capsules on Day 1, 3, 5 and 7 of the differentiation. Cells were then fixed by 4% paraformaldehyde for 20 min, washed with PBS solution containing 0.03% Triton X-100 for 10 min, rinsed by PBS again, immersed in 3% H₂O₂ for 10 min, washed with PBS, blocked by normal goat serum and incubated at 4°C overnight with one of the following primary antibodies: rabbit anti-rat nestin polyclonal antibody (1:200, Boster, Wuhan, China) for NSCs, rabbit anti-rat neuron-specific enolase (NSE) polyclonal antibody (1:400, Boster) for neurons, anti-rat CaM monoclonal antibody (1:2,000, Sigma, USA) and rabbit anti-rat actin polyclonal antibody (1:100, Boster). Then, the biotinylated secondary antibody and ABC complex (Boster) were added in sequence followed by development with DAB. The reaction time was controlled under a microscope. Samples were washed again by distilled water, dehydrated by series of alcohols, processed by xylene and sealed by Balsam neutral. The control group used PBS to replace the primary antibody with all other staining steps same as above. The Olympus optical microscope was used to observe the cells and to capture images.

Immunofluorescence double-labeling of CaM and actin. Coverslips were taken out from the culture dishes on Day 1, 3, 5 and 7 of differentiation, fixed with 4% paraformaldehyde for 20 min, washed with PBS solution containing 0.03% Triton X-100, rinsed with PBS, blocked with normal goat serum and incubated with CaM (1:2,000) and actin (1:300)

Table I. The primers of calmodulin (CaM) and β -actin.

Name	Primers	Product size (bp)
CaM	Forward 5'-ggcatcctgcttagcctgag-3'	328
	Reverse 5'-acatgctatccctctcgtgtga-3'	
β -actin	Forward 5'-atcaccacactgtgcccatc-3'	542
	Reverse 5'-acagagtactgcgctcagga-3'	

mixed antibodies at 4°C overnight, followed by addition of Cy3-labeled anti-mouse IgG antibody (1:100, Gibco) and FITC-labeled anti-rabbit IgG antibody (1:50, Boster) mixed secondary antibodies for 1 h, rinsed by PBS and sealed by 50% glycerol. Fluorescent signals were detected using a Nikon compound fluorescent microscope and a Nikon C1 Plus confocal microscope (Nikon, Japan).

Reverse transcription-polymerase chain reaction to detect CaM. Total-mRNA was extracted from differentiation of NSCs according to the instructions of the TRIzol kit. The primers designed by Shenggong Biotech Company (Shanghai, China) according to the serial numbers from GenBank are shown in Table I. PCR was performed using Taq Dynazyme (Takara, Dalian, China) under standard conditions. The reaction was started at 94°C for 4 min and amplification for CaM occurred for 32 cycles of 94°C for 30 sec, 58°C for 30 sec, 72°C for 40 sec, followed by 7 min final extension at 72°C. β -actin mRNA was co-amplified with CaM mRNA as an internal control. The products were observed after electrophoresis on 1.2% agarose gel, and the density of each band was analyzed on the Gel Image Analysis System. The level of CaM mRNA was determined by calculating the density ratio of CaM mRNA/ β -actin mRNA.

Statistical analysis. The positive products of CaM and actin in directed differentiation of NSCs into neurons were analyzed with the Motic Instruments Images Advanced 3.0 image analysis system. Under the microscope (x400), 50 cells were quantified in each group, and the average integral optical density (IOD) was determined. Differences between mean values of CaM or actin for different time points were analyzed using one-way ANOVA, and independent sample t-test was performed to detect the difference between mean values of CaM and actin at the same time points. The data are expressed as mean \pm SD and the difference was considered significant at $P < 0.05$.

Results

Identification of NSCs and directed differentiation of NSCs into neuron. On Day 7 of culture, each colony included up to dozens to hundreds of cells. Colony forming units appeared as spherical, oval or morular shapes and grew in suspension (Fig. 1A) with good vitality. These cell colonies showed strong immunological reaction signals by nestin immunohistochemical staining (Fig. 1B).

NSCs were dispersed by mechanical force and cultured in medium containing serum and induction factors. After 4 h,

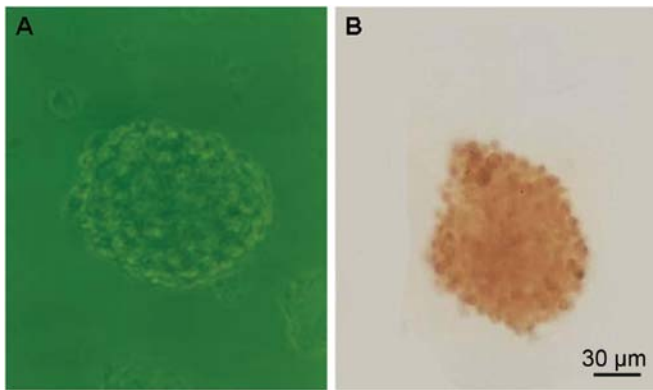


Figure 1. Culture and identification of neonatal rat cortical NSCs. (A) NSCs proliferating into neurospheres. (B) Immunohistochemical staining showed that neurospheres were nestin positive.

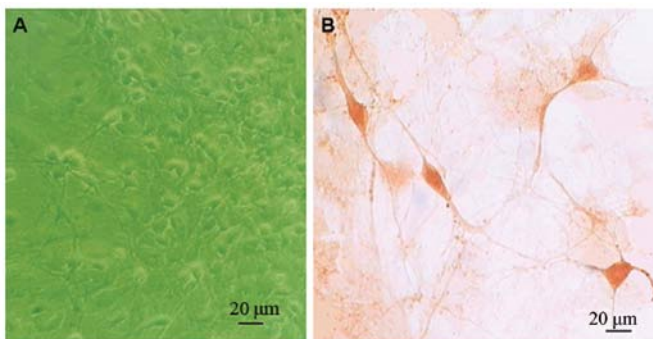


Figure 2. Differentiation of neonatal rat cortical NSCs. (A) On Day 7 of differentiation of neonatal rat cortical NSCs, various morphological cells were observed under inverted phase contrast microscopy. (B) Immunohistochemical staining showed that some of these differentiating cells were NSE-positive, which were part of the directed differentiation of neonatal rat cortical NSCs into neurons.

cells started to attach and some cells differentiated to possess neurites on Day 2. Among these differentiated cells, there were three types: some were larger cells, attached flat on the coverslip, with many thick neurites; these cells had a large quantity. The second type of cells were smaller, with fewer but very long neurites, highly refractive and a part of which sat upon the larger cells; there was a small quantity of these cells. The last type of cells were the smallest and fewest, with relatively short and thin neurites (Fig. 2A). Immunohistochemical staining showed that some of these directed NSCs were NSE-positive (Fig. 2B).

CaM immunocytochemical staining. During the directed differentiation of cerebral cortical NSCs into neurons, on the first day, cell bodies were small, and neurites were short. CaM proteins were mainly located in and around nucleus (Fig. 3A). On the Days 3, 5 and 7 of differentiation, as the cells grew, cell bodies became bigger, and neurites became longer and more plentiful. A positive expression of CaM proteins was observed in both the cytoplasm and in neurites, which increased as the neurites extended. We could see segmental positive expression products extending to the extremities of the neurites. However, the positive reaction of the nucleus was weaker than that on the

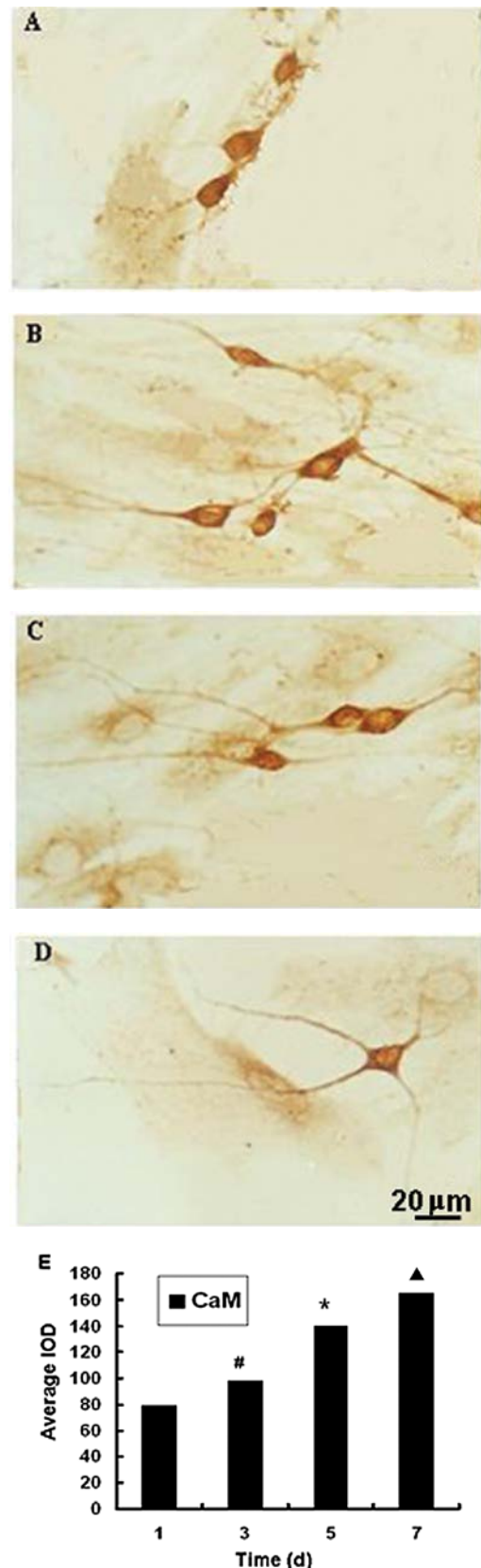


Figure 3. CaM immunohistochemical staining during the directed differentiation of neonatal rat cortical NSCs into neurons. (A) At the early stage of differentiation, cell bodies were small, and a brown positive reaction was shown in and around the nucleus. (B-D) As cells differentiated, positive reaction products extended segmentally into the cytoplasm and neurites. (E) Average integral optical density value of CaM during different stages of the directed differentiation of cerebral cortical neural stem cells into neurons. The data are expressed as mean \pm SD, # P <0.01 vs. Day 1; * P <0.01 vs. Day 3; ▲ P <0.01 vs. Day 5.

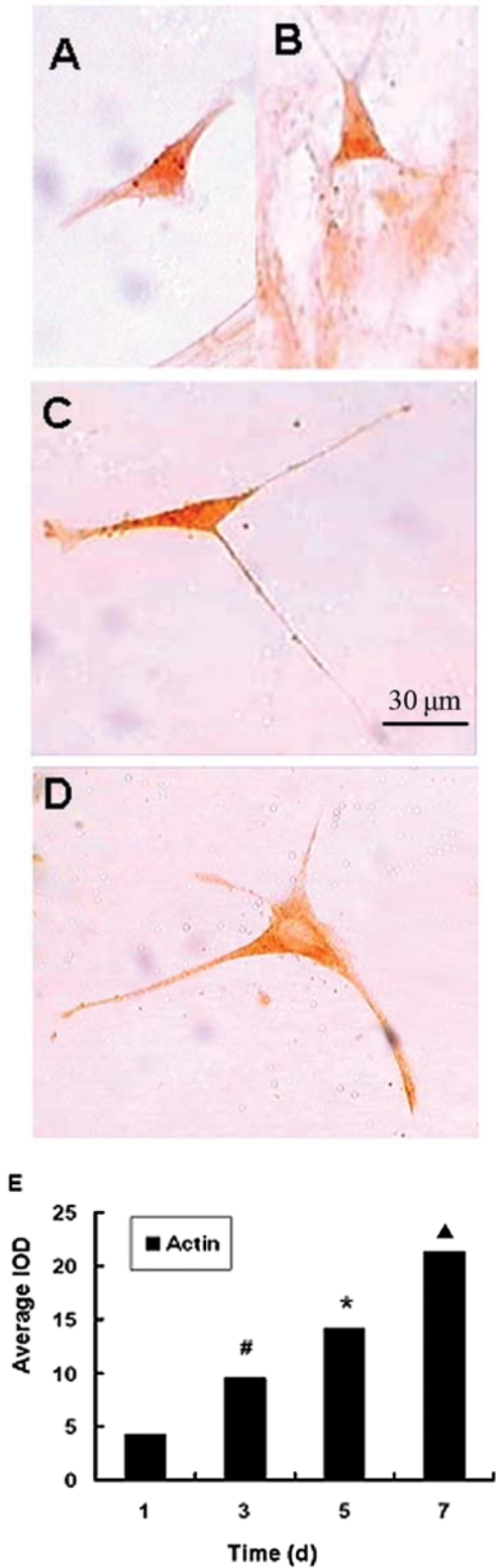


Figure 4. Actin immunohistochemical staining during the directed differentiation of neonatal rat cortical NSCs into neurons. (A) On the first day of differentiation, actin was mainly expressed around the nucleus and pinosomes were seen in the cytoplasm. (B and C) On Day 3 and 5 of differentiation, actin was expressed both in the cytoplasm and in neurites. Byssoid brown actin positive products could be seen. (D) On Day 7 of differentiation, filiform cytoskeleton was seen clearly in cytoplasm and neurites, which was arranged in parallel to the neurites and extended with them. (E) The average integral optical density value of actin during the different stages of directional differentiation of cerebral cortical NSCs into neurons. The data are expressed as the mean \pm SD; #*P*<0.01 vs. Day 1; **P*<0.01 vs. Day 3; ▲*P*<0.01 vs. Day 5.

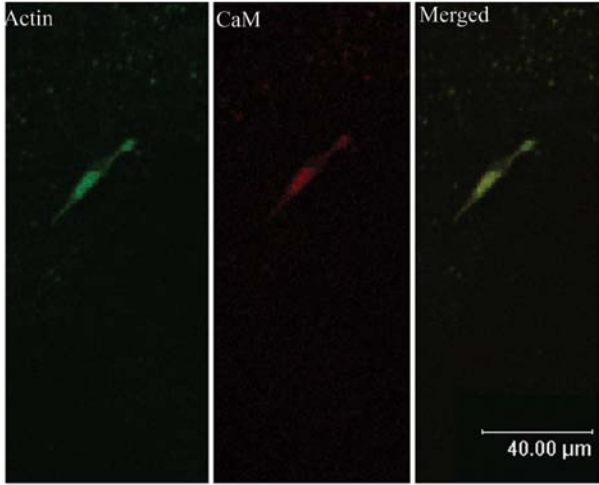


Figure 5. Observation of fluorescence double-labeled CaM and actin on the first day of differentiation. A smaller cell body with short neurites was observed (actin, green; CaM, red; merged image, yellow). Distribution patterns of CaM and actin were largely similar.

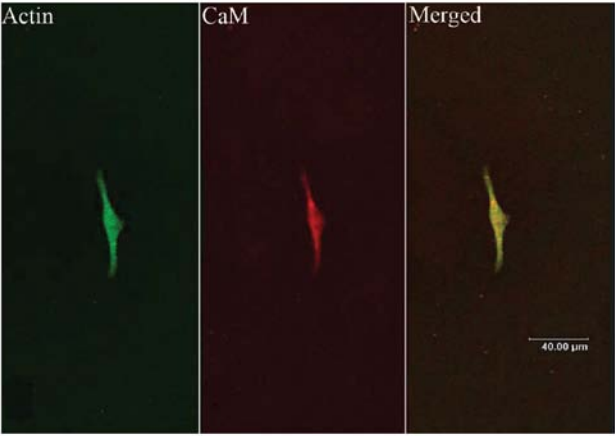


Figure 6. Observation of fluorescence double-labeled CaM and actin on Day 3 of differentiation. Cell bodies were enlarged and neurites were elongated. CaM and actin had similar distribution patterns and gradually extended into the neurites. Orange fluorescence appeared around the nucleus.

first day (Fig. 3B-D). The average integral optical density value of CaM during the different stages of directed differentiation of cerebral cortical NSCs into neurons is shown in Fig. 3E.

Actin immunocytochemical staining. Actin positive staining shown as brown granulations were located in the cytoplasm of neurons, especially around the nucleus. However, there was no expression of actin in the nucleus. On the first day after differentiation, actin was expressed mainly around the nucleus, and pinosomes were seen in the cytoplasm (Fig. 4A). However, with maturation of cells, on Day 3 and 5, actin was expressed both in the cytoplasm and in neurites. It was more strongly expressed around the nucleus, and its distribution near the base of neurites appeared to be reticular (Fig. 4B). On Day 7 after differentiation, cells were nearly mature. The expression of actin was even more increased around the nucleus, and the brown positive reaction was darker. The filiform cytoskeleton was arranged in parallel in the cytoplasm and extended into

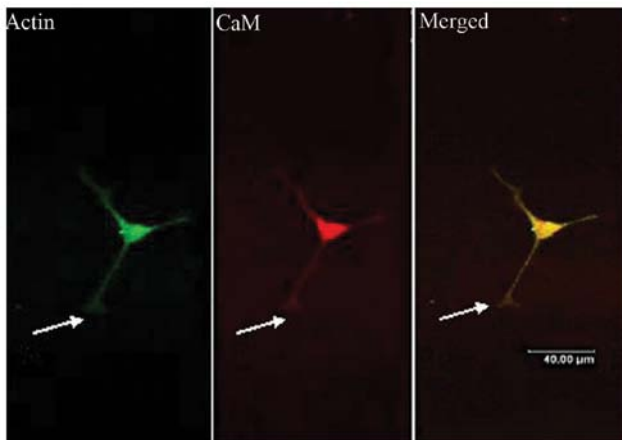


Figure 7. Observation of fluorescence double-labeled CaM and actin on Day 5 of differentiation. Cell bodies further enlarged and neurites elongated in continuously differentiating neurons. Expression of CaM and actin extended like a duck web into the extremities of neurites.

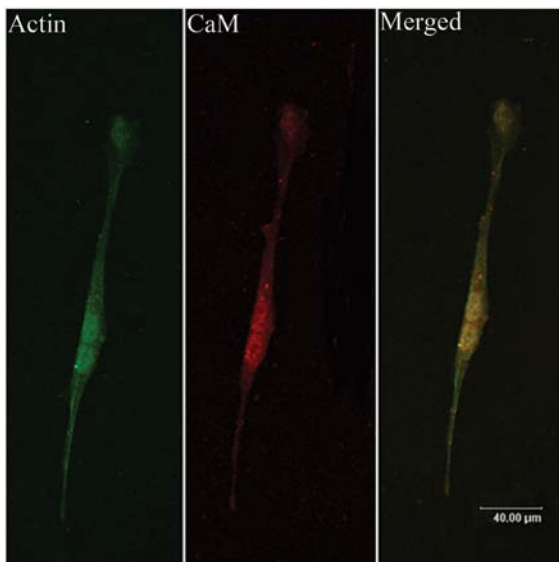


Figure 8. Observation of fluorescence double-labeled CaM and actin on Day 7 of differentiation. Orange fluorescence appeared both around the nucleus and at the neurite terminus. However, they also showed independent existence without overlapping within part of the process.

neurites (Fig. 4C and D). On Days 1, 3, 5 and 7, the quantity of expression gradually increased, and there were significant differences in actin protein expression ($P<0.01$) (Fig. 4E).

Co-expression of CaM and actin in differentiating neurons. In the same sample, excitation by different wavelengths of mono-fluorescence can reveal the existence and distribution pattern of red fluorescent CaM (excitation wavelength of 543 nm) and green fluorescent actin (excitation wavelength of 488 nm) in neurons. They showed a similar localization distribution. Specifically, at the early stage of differentiation, these two proteins concentrated inside the nucleus and at the vicinity of the nucleus (Fig. 5). With the maturation of neurons, the fluorescence gradually extended into the cytoplasm and neurites of the continuously differentiating neurons (Figs. 6-8). These two fluorescence signals largely co-localized in the cytoplasm

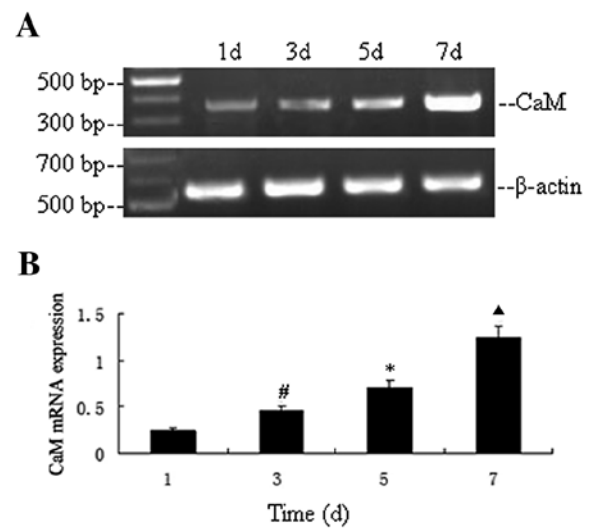


Figure 9. Expression of CaM mRNA. (A) The representative gel pattern of CaM and β -actin of cDNA bands. (B) Densitometric analysis of CaM of the semi-quantitative RT-PCR. Data are expressed as the mean \pm SD * $P<0.05$ vs. Day 1; # $P<0.05$ vs. Day 3; ▲ $P<0.05$ vs. Day 5.

and neurites and overlapped with each other. However, in some cases, they showed independent existence without overlapping, suggesting the correlation between CaM and actin.

Expression of CaM and actin mRNA. The representative gel pattern of CaM assessed the level of CaM mRNA normalized to the β -actin mRNA level. The level of CaM mRNA showed a significant up-regulation on Day 3, 5 and 7 compared with that on the first day after differentiation (Fig. 9).

Discussion

NSCs are multipotent with the ability of self-renewal. They can differentiate into neurons, astrocytes and oligodendrocytes (8). Currently, the regular methodology is to use a combination of FGF-2 and EGF co-factors to culture NSCs *in vitro* (9-12). In this study, we used the same method to culture NSCs and confirmed the cell identify using immunohistochemical staining for nestin, a protein that disappears along with the differentiation and maturation of the neural epithelium (13-16).

CaM has been proven to be a versatile Ca^{2+} -binding protein and exists in neural cells in a large quantity (17-19). In the nervous system, CaM participates in many important biological functions while multiple activities of neural cells are subject to the regulation by Ca^{2+} -CaM signaling (17). Previous studies have proven that CaM plays important roles in many biological processes, such as early development of neurites, regulation of microtubule formation, neuron metabolism, axoplasmic transport, synaptic transmission and neurotransmitter synthesis, release and transport (20-23).

As a cytoskeletal protein, actin plays a key role in maintaining the mobility of the neuronal growth cones (24-27). Our results confirmed that in the process of neuron-oriented rat cerebral cortical NSC differentiation, the expressions of CaM and actin were in accordance with each other, they extended into the neurites and even in the duck-web like lamellipodia corresponding to the growth and development of

neurons, which was shown both under CLSM and by immunohistochemistry. These results suggest that the role of actin in neurite development is modulated by CaM. At the early stage of neuron-oriented cerebral cortical NSC differentiation, the relationship between CaM and actin was closely related to the robust neuron functional activities. At the early developmental stage of neural cells, an increase in intracellular free Ca^{2+} concentration affected the initiation of cell migration and the connection between cells, which maintained cell morphology of the neuron and promoted the extension of neurites. However, those functions executed by Ca^{2+} needed the involvement of CaM (28). In addition, Ca^{2+} was a very effective regulator in which it could significantly affect the structure and function of actin (29). Ca^{2+} -CaM have been shown to be related to the assembly and disassembly of tubulin, thereby affecting the polymerization/depolymerization of microfilaments, the dynamic change and balance which are important regulatory factors for cell movement, attachment and the cell division cycle (28). Therefore, actin determined the active movement of lamellipodia and filopodia in neuronal growth cones subjected to the influence of Ca^{2+} -CaM, so that filopodia and lamellipodia were able to extend leading to the growth and extension of axons (30). This study only observed the co-expression of actin and CaM in the early stage of differentiating neural cells. However, how CaM affects the impact of actin on growth cones, as well as whether this similar expression pattern exists at late stages of neuronal differentiation, are still subject to study.

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References

1. Kitamura Y, Arima T, Imaizumi R, Sato T and Nomura Y: Inhibition of constitutive nitric oxide synthase in the brain by pentamidine, a calmodulin antagonist. *Eur J Pharmacol* 289: 299-304, 1995.
2. Schaloske RH, Lusche DF, Bezares-Roder K, Happel K, Malchow D and Schlatterer C: Ca^{2+} regulation in the absence of the iPLA gene product in *Dictyostelium discoideum*. *BMC Cell Biol* 6: 13, 2005.
3. Dabrowska R, Kulikova N and Gagola M: Nonmuscle caldesmon: its distribution and involvement in various cellular processes. *Protoplasma* 224: 1-13, 2004.
4. Levinson H, Moyer KE, Siggers GC and Ehrlich HP: Calmodulin-myosin light chain kinase inhibition changes fibroblast-populated collagen lattice contraction, cell migration, focal adhesion formation, and wound contraction. *Wound Repair Regen* 12: 505-511, 2004.
5. Larsson C: Protein kinase C and the regulation of the actin cytoskeleton. *Cell Signal* 18: 276-284, 2006.
6. Furukawa R, Maselli A, Thomson SA, Lim RW, Stokes JV and Fechtmeier M: Calcium regulation of actin crosslinking is important for function of the actin cytoskeleton in *Dictyostelium*. *J Cell Sci* 116: 187-196, 2003.
7. Lieto-Trivedi A and Coluccio LM: Calcium, nucleotide, and actin affect the interaction of mammalian Myo1c with its light chain calmodulin. *Biochemistry* 47: 10218-10226, 2008.
8. Temple S and Alvarez BA: Stem cells in the adult mammalian central nervous system. *Curr Opin Neurobiol* 9: 135-141, 1999.
9. Gritti A, Coval L, Parati EA, Galli R and Vescovi AL: Basic fibroblast growth factor supports the proliferation of epidermal growth factor-generated neuron precursor cells of the adult mouse CNS. *Neurosci Lett* 185: 151-154, 1995.
10. Ciccolini F and Svendsen CN: Fibroblast growth factor 2 (FGF-2) promotes acquisition of epidermal growth factor (EGF) responsiveness in mouse striatal precursor cells: identification of neural precursors responding to both EGF and FGF-2. *J Neurosci* 18: 7869-7880, 1998.
11. Takahashi T, Palmer TD and Gage FH: Retinoic acid and neurotrophins collaborate to regulate neurogenesis in adult-derived neural stem cell cultures. *J Neurobiol* 38: 65-81, 1999.
12. Tatebayashi T, Iqbal K and Grundke-Iqbal I: Dynamic regulation of expression and phosphorylation of tau by fibroblast growth factor-2 in neural progenitor cells adult rat hippocampus. *J Neurosci* 19: 5245-5254, 1999.
13. Pérez Castillo A, Aguilar-Morante D, Morales-García JA and Dorado J: Cancer stem cells and brain tumors. *Clin Transl Oncol* 10: 262-267, 2008.
14. Gilyarov AV: Nestin in central nervous system cells. *Neurosci Behav Physiol* 38: 165-169, 2008.
15. Gilyarov AV: Nestin in the cells of the central nervous system. *Morfologiya* 131: 85-90, 2007 (In Russian).
16. Daniel C, Albrecht H, Lüdke A and Hugo C: Nestin expression in repopulating mesangial cells promotes their proliferation. *Lab Invest* 88: 387-397, 2008.
17. Thomas R, Bill C and Debra B: Cellular signaling through multifunctional Ca^{2+} /Calmodulin-dependent protein kinase II. *Biol Chem* 276: 3719-3722, 2001.
18. Yamasaki TR, Blurton-Jones M and Morrisette DA: NSCs improve memory in an inducible mouse model of neuronal loss. *J Neurosci* 27: 11925-11933, 2007.
19. Jori FP, Napolitano MA, Melone MA, Cipollaro M, Cascino A, Altucci L, Peluso G, Giordano A and Galderisi U: Molecular pathways involved in neural in vitro differentiation of marrow stromal stem cells. *J Cell Biochem* 94: 645-655, 2005.
20. Deisseroth K, Heist EK and Tsien RW: Translocation of calmodulin to the nucleus supports CREB phosphorylation in hippocampal neurons. *Nature* 392: 198-202, 1998.
21. Corcoran EE and Means AR: Defining Ca^{2+} /calmodulin-dependent protein kinase cascades in transcriptional regulation. *J Biol Chem* 276: 2975-2978, 2001.
22. Laporte SA, Oakley RH and Caron MG: Signal transduction. Bringing channels closer to the action. *Science* 293: 62-63, 2001.
23. Kasahara J, Fukunaga K and Miyamoto E: Activation of calcium/calmodulin-dependent protein kinase IV in long term potentiation in the rat hippocampal CA1 region. *J Biol Chem* 276: 24044-24050, 2001.
24. Wang G, Krishnamurthy K, Chiang YW, Dasgupta S and Bieberich E: Regulation of neural progenitor cell motility by ceramide and potential implications for mouse brain development. *J Neurochem* 106: 718-733, 2008.
25. Dore-Duffy P, Katyshev A, Wang X and Van Buren E: CNS microvascular pericytes exhibit multipotential stem cell activity. *J Cereb Blood Flow Metab* 26: 613-624, 2006.
26. Lo Iacono L and Gross C: Alpha- Ca^{2+} /calmodulin-dependent protein kinase II contributes to the developmental programming of anxiety in serotonin receptor 1A knock-out mice. *J Neurosci* 28: 6250-6257, 2008.
27. Wei J, Zhang M, Zhu Y and Wang JH: Ca^{2+} -calmodulin signalling pathway up-regulates GABA synaptic transmission through cytoskeleton-mediated mechanisms. *Neuroscience* 127: 637-647, 2004.
28. Nebl T and Fisher PR: Intracellular Ca^{2+} signals in *Dictyostelium* chemotaxis are mediated exclusively by Ca^{2+} influx. *J Cell Sci* 110: 2845-2853, 1997.
29. Zuo H, Cui B, She X and Wu M: Changes in Guinea pig cochlear hair cells after sound conditioning and noise exposure. *J Occup Health* 50: 373-379, 2008.
30. Papp S, Szabo E, Kim H, McCulloch CA and Opas M: Kinase-dependent adhesion to fibronectin: regulation by calreticulin. *Exp Cell Res* 314: 1313-1326, 2008.