

# Expression of p-STAT3 in human gastric carcinoma: Significant correlation in tumour invasion and prognosis

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**Abstract.** Signal transducers and activators of transcription (STATs) belong to a family of transcription factors activated in response to cytokines and growth factors. Constitutive activation of STAT3 has been observed in a growing number of tumour-derived cell lines, as well as in tumor specimens from human cancers. Our aim was to determine the relationship between the expression of phosphorylated STAT3 (p-STAT3) and clinicopathological factors in human gastric adenocarcinoma. One-hundred and eleven cases of surgically resected human gastric adenocarcinoma were studied by immunohistochemistry. Fifty-five (49.5%) cases showed positive staining for the p-STAT3 proteins. The expression of p-STAT3 was significantly correlated with several clinicopathological factors. Moreover, expression of p-STAT3 was detected by Western blot analysis in 2 different kinds of cultured human gastric carcinoma cell lines and 4 cases of gastric carcinoma tissue obtained at surgery. Western blot analysis confirmed the presence of p-STAT3 in these specimens. In univariate survival analysis, p-STAT3 expression was associated with inferior survival ( $p < 0.05$ ). Constitutive activation of STAT3 may play an important role in the tumorigenesis of gastric adenocarcinoma, and the detailed mechanism of STAT3 signaling pathway in gastric adenocarcinoma deserves further investigation.

## Introduction

Gastric cancer remains one of the most common cancer types in the world today, despite the fact that the incidence has gradually declined in many countries (1). The occurrence and progression of cancer is related to a series of genetic events

affecting the structure and/or expression of a number of oncogenes, anti-oncogenes and growth factors (2,3). Potentially deeply invasive carcinomas, such as gastric carcinoma, have relatively high rates of lymph duct and venous invasion and lymph node metastasis (4). However, the mechanisms of invasion and metastasis of gastric carcinoma have not been fully elucidated.

Signal transducer and activator of transcription (STAT) proteins belong to a family of transcription factors that are normally inactive within the cytoplasm of cells and become activated by tyrosine phosphorylation in response to cytokines and growth factors. In normal cells, STAT activation is transient whereas, in a large number of primary tumors and cancer-derived cell lines, STAT proteins (in particular STAT3) remain activated by persistently activated tyrosine kinases and/or a decrease in the negative regulators of STAT dephosphorylation (5,6). The binding of a cytokine to its cognate receptor rapidly induces the tyrosine phosphorylation of the receptor by Jak kinases. Activated STATs rapidly translocate into the nucleus, bind to recognition sequences in the promoter region of target genes, and regulate their transcription. Recent studies have demonstrated the essential roles of STAT proteins in modulating the process of cell proliferation, differentiation and apoptosis (7-9).

Constitutively activated STAT proteins have been observed in a wide variety of human tumour cell lines and primary tumours including leukemia, multiple myeloma, breast cancer, prostate cancer, and other cancers (10-17). STAT3 has been classified as an oncogene because constitutively activated STAT3 was found to mediate oncogenic transformation in cultured cells and tumour formation in nude mice (18,19). STAT3 activation may not only provide a growth advantage, but also confer resistance to conventional therapies that depend on the mechanism of apoptosis to eliminate tumour cells (20). Previous studies showed that constitutive activation of STAT3 correlated with cyclin D1 expression, possibly providing a prognosis marker in head and neck cancer (21,22). These findings suggest that constitutive activation of STAT3 participates in the development of different human malignancies.

An antibody specific for the activated (phospho-Tyr705) form of STAT3 (p-STAT3) has become available (23). In prostate tissue, the activated form of STAT3 localized predominantly to the nuclei of malignant glands (24). This

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activated form may be a better probe for function than total STAT3, but it has not yet been evaluated in gastric cancer tissues. We evaluated the expression and subcellular localization of phospho-STAT3 (Tyr705) by immunohistochemistry in human gastric cancer.

### Materials and methods

We studied 10 cases of gastric adenoma and 111 cases of primary human gastric carcinoma by immunohistochemistry. Of the 111 patients with gastric carcinoma, there were 63 men and 48 women. The median age was 68.9 years (range, 38-89). All tumors were obtained from patients who had undergone surgery or endoscopic resection at Nagasaki University Hospital and Shunkaikai Inoue Hospital between 2000 and 2005. Fifteen specimens of gastric tissue obtained from a normal area adjacent to the tumour specimens (5-10 cm away) were taken and evaluated as normal controls. All samples were snap-frozen within 15 min after surgical removal to ensure preservation of STAT3 activities.

Each tumour was assigned a histological type and a depth grading of infiltration according to the Japanese Classification of Gastric Carcinoma by the Japanese Research Society for Gastric Cancer (25), based on the World Health Organization classification (WHO). Histologically, of the 111 primary human gastric adenocarcinoma cases, 6 were papillary, 20 were tubular of the well-differentiated type, 37 were tubular of the moderately differentiated type, 10 were poorly differentiated adenocarcinoma of the solid type, 16 were poorly differentiated of the nonsolid type, 17 were signet-ring cell carcinoma, and 5 were mucinous adenocarcinoma. A total of 15 normal gastric mucosal tissues were evaluated as normal controls.

Lymphatic and venous invasion was studied on routine hematoxylin and eosin-stained slides. In addition, the Elastica van Gieson stain was used in all cases. Each parameter was defined as 'present' only when invasion was identified with certainty, but defined as 'absent' when not seen at all or not seen with certainty (26,27). Lymph node metastasis was defined as 'present' only when confirmed histologically. The diagnosis was established by two independent pathologists (Y. Yakata and T. Nakayama) and cases with a questionable diagnosis were excluded from the study.

**Immunohistochemistry.** Formalin-fixed and paraffin-embedded tissues, deparaffinized in xylene, and dehydrated in phosphate-buffered saline. Deparaffinized sections were preincubated with an optimal dilution (0.1  $\mu$ g/ml) of a primary polyclonal goat antibody against human phosphorylated STAT3 (p-STAT3; Santa Cruz Biotechnology, Inc., Santa Cruz, CA, USA). The slides were sequentially incubated with a biotinylated and alkaline phosphatase-conjugated goat anti-rabbit immunoglobulin G antibody, and the reaction products were visualized using diaminobenzidine (DAB; Dako, Carpinteria, CA, USA) with methyl green as a counterstain. Prostatic cancer tissue served as the positive control (24). Two investigators analyzed the immunohistochemical staining (Y. Yakata and T. Nakayama). p-STAT3 expression was classified into two categories, depending on the percentage of cells stained: 0 to 10% positive cells were considered negative (-) and more than 10% positive cells were considered positive (+).

**Cell culture.** Two human gastric carcinoma cell lines, MKN-28 and SCH, were incubated at 37°C in a humidified atmosphere of 5% CO<sub>2</sub>. MKN-28 and SCH were maintained in RPMI-1640 medium (Invitrogen Corp. NY, USA) supplemented with 10% fetal calf serum. All of these cells were provided by the Japanese Collection of Research Bioresources (Tokyo, Japan). For the induction of p-STAT3, two human gastric carcinoma cell lines were stimulated by 100 ng/ml of interferon- $\alpha$  (IFN- $\alpha$ ; Cell Signaling Technology, Beverly, MA, USA) (28).

**Immunoblotting.** Specimens and cells were resuspended in ice-cold radioimmunoprecipitation (RIPA) buffer and one tablet of complete proteinase inhibitor mixture (Roche Applied Science, Indianapolis, IN, USA) per 50 ml for 10 min, sonicated on ice, and centrifuged (12,000 rpm, 15 min at 4°C). Protein concentration of the supernatant (protein fraction) was determined by the Bradford protein assay (Bio-Rad, Hercules, CA, USA). An aliquot of 10  $\mu$ g of protein was mixed with an equivalent volume of 2X protein loading buffer containing 2- $\beta$ -mercaptoethanol and boiled for 5 min before loading onto a 15% SDS-polyacrylamide gel. After electrophoresis, proteins were transferred onto nitrocellulose membranes using ECL (Amersham Biosciences, Piscataway, NJ, USA) and blocked in TBST containing 5% non-fat dry milk powder. Protein immunoblots were performed using specific antibodies to  $\beta$ -actin (clone JLA20; Calbiochem, San Diego, CA, USA), phosphotyrosine (Tyr<sup>705</sup>) STAT3 (Cell Signaling Technology), and STAT3 (Santa Cruz Biotechnology Inc.). The membranes were further incubated with peroxidase-conjugated secondary antibodies, and protein bands were visualized using a commercial chemiluminescence detection kit (ECL Plus; Amersham Biosciences) as described by the manufacturer.

**Statistical analysis.** The Stat View II program (Abacus Concepts, Inc., Berkeley, CA, USA) was used for statistical analyses. Analyses comparing the expression of p-STAT3 were performed by the Chi-square test for independence and the Mann-Whitney U test. Survival durations were calculated using the Kaplan-Meier method. The log-rank test was used to compare the cumulative survival of patient groups.

### Results

The results from immunohistochemical staining are summarized in Table I. Expressed p-STAT3 was heterogeneous in the cytoplasm and nucleus of carcinoma cells. p-STAT3 immunoreactivity in relation to histological type is shown in Table I. Poorly differentiated adenocarcinoma of the solid and nonsolid type and tubular adenocarcinoma of the moderately differentiated type gastric carcinoma stained for p-STAT3. However, tubular adenocarcinoma of the well-differentiated type expressed relatively weak p-STAT3. In signet-ring cell carcinoma, most cases were negative for p-STAT3 (4 of 17, 23.5%). All cases of 5 mucinous carcinoma were negative group-STAT3. All 10 cases of adenoma and all 15 cases of normal mucosa were negative for p-STAT3. There was a statistical difference between total carcinoma and adenoma (p<0.05).

Table I. p-STAT3 expression in gastric carcinoma.

|                              | n   | -           | +          | p-value              |
|------------------------------|-----|-------------|------------|----------------------|
| Tubular adenoma              | 10  | 10 (100.0%) | 0 (0.0%)   | p<0.005 <sup>a</sup> |
| Total carcinoma              | 111 | 56 (50.5%)  | 55 (49.5%) |                      |
| Histological differentiation |     |             |            | n.s.                 |
| papillary                    | 6   | 2 (33.3%)   | 4 (66.7%)  |                      |
| tub/well                     | 20  | 14 (70.0%)  | 6 (30.0%)  |                      |
| tub/moderate                 | 37  | 14 (37.8%)  | 23 (62.1%) |                      |
| poor/solid                   | 10  | 3 (30.0%)   | 7 (70.0%)  |                      |
| poor/nonsolid                | 16  | 5 (31.3%)   | 11 (68.8%) |                      |
| sig                          | 17  | 13 (76.5%)  | 4 (23.5%)  |                      |
| muc                          | 5   | 5 (100.0%)  | 0 (0.0%)   |                      |
| Lauren's classification      |     |             |            | n.s.                 |
| Intestinal                   | 63  | 30 (47.6%)  | 33 (52.4%) |                      |
| Diffuse                      | 48  | 26 (54.2%)  | 22 (45.8%) |                      |
| Depth of invasion            |     |             |            | p<0.001 <sup>b</sup> |
| m                            | 26  | 24 (92.3%)  | 2 (7.7%)   |                      |
| sm                           | 37  | 21 (56.8%)  | 16 (43.2%) |                      |
| mp                           | 12  | 4 (33.3%)   | 8 (66.7%)  |                      |
| ss                           | 11  | 2 (18.2%)   | 9 (81.8%)  |                      |
| se                           | 25  | 5 (28.0%)   | 20 (80.0%) |                      |
| Lymph node metastasis        |     |             |            | p<0.005              |
| Present                      | 32  | 8 (25.0%)   | 24 (75.0%) |                      |
| Absent                       | 79  | 51 (64.6%)  | 28 (35.4%) |                      |

<sup>a</sup>Significant difference between adenoma and total carcinoma in the expression of p-STAT3. <sup>b</sup>Significant difference between mucosal carcinomas and other carcinomas. n.s., not significant; tub, tubular adenocarcinoma; sig, signet ring-cell carcinoma; muc, mucinous carcinoma.

Fig. 1 shows a representative example of strong immunohistochemical p-STAT3 staining in a carcinoma invading the subserosa. p-STAT3 protein was detected in both the cytoplasm and the nucleus of almost all carcinomas (Fig. 1B). Fig. 2 shows a mucosal carcinoma with negative staining for p-STAT3.

The presence of immunoreactivity appears to be correlated with the degree of tumour infiltration (Table I). Staining for p-STAT3 was mainly negative for tumours with mucosal infiltration (only 2 of 26 were positive, 7.7%), whereas 43.2% of submucosal infiltrative tumours were p-STAT3 immunoreactive (16 of 37). In tumours infiltrating the proprial muscle layer, the subserosa and the serosal surface, p-STAT3 was frequently expressed (37 of 48, 77.1%). Statistical analysis showed significant differences between mucosal carcinomas and the other carcinomas (p<0.001; Table I). In the invasive carcinomas, the staining intensity in signet-ring cell carcinomas and mucinous adenocarcinomas was statistically lower than in the other type of carcinoma. Also, the invasive front and/or the peripheral parts of the primary tumour were intensely stained compared with superficial and central parts of the tumour in almost all cases.

p-STAT3 expression in primary carcinomas correlated with the presence of lymph node metastasis (p<0.005; Table I). Moreover, p-STAT3 expression in primary carcinoma

correlated with the presence of venous invasion (p<0.0001) and lymphatic invasion (p<0.0005) (Table II). There was no correlation between p-STAT3 expression and the differentiation or the pattern of tumour infiltration of primary carcinomas.

The Western blot analyses of p-STAT3 expression in MKN-28 and SCH, human gastric carcinoma cell lines, are shown in Fig. 3. STAT3 expression was detected in both cell lines. P-STAT3 expression after IFN- $\alpha$  stimulation also was up-regulated in both cell lines, whereas a weak expression of p-STAT3 was observed without IFN- $\alpha$  stimulation.

The results of Western blot analysis of p-STAT3 expression in surgical specimens of human gastric cancer are shown in Fig. 4. The STAT3 non-activated type was expressed in all samples. p-STAT3 expression was detected in normal mucosa (N), whereas strong expression of p-STAT3 was observed in the gastric carcinoma tissues (T).

The respective Kaplan-Meier curves are given in Fig. 5. The corresponding log-rank test shows that there is a significant difference between p-STAT3 positive and negative subgroups in gastric cancer patients (p<0.05).

## Discussion

We investigated the expression of p-STAT3 in gastric carcinoma using immunohistochemical techniques and

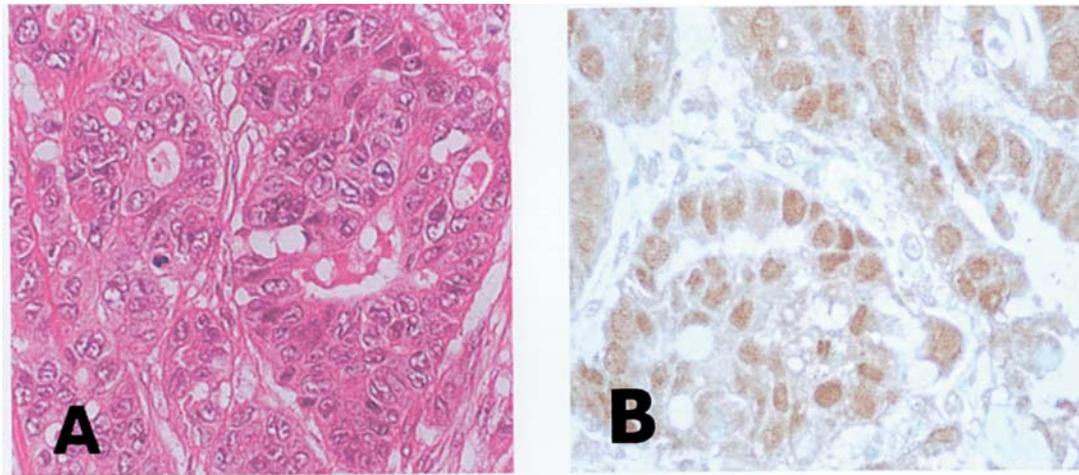


Figure 1. (A) Invasive gastric adenocarcinoma to subserosa (hematoxylin and eosin staining, x400). (B) p-STAT3 reveals positive staining in the cytoplasm and nucleus of carcinoma cells (DAB staining, x400).

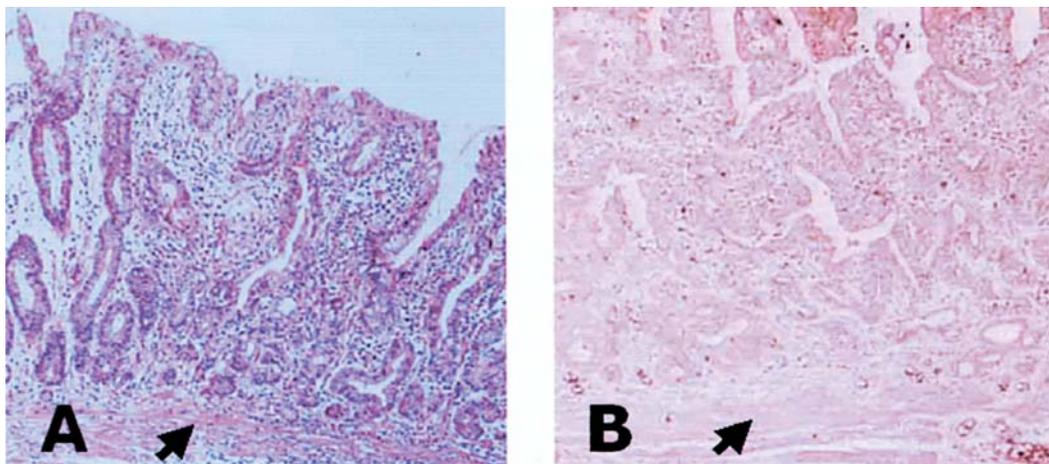


Figure 2. (A) Mucosal gastric adenocarcinoma. Arrow shows the muscularis mucosa (hematoxylin and eosin staining, x100). (B) Carcinoma cells are negative for p-STAT3. Arrow shows the muscularis mucosa (DAB staining, x100).

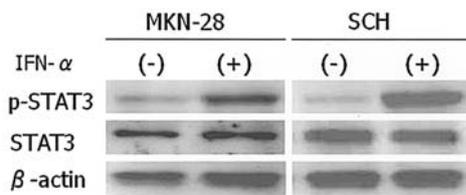


Figure 3. Expression of STAT3 and p-STAT3 in human gastric adenocarcinoma cell lines (MKN28, SCH). Cells were stimulated with 100 ng/ml of IFN- $\alpha$ .  $\beta$ -actin represents the internal protein control. Two cell lines demonstrate elevated levels of p-STAT3.

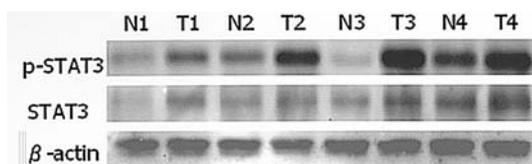


Figure 4. Expression of STAT3 and p-STAT3 in human gastric carcinoma.  $\beta$ -actin represents the internal protein control. Elevated levels of p-STAT3 in tumour (T) tissues were compared to adjacent normal mucosa (N).

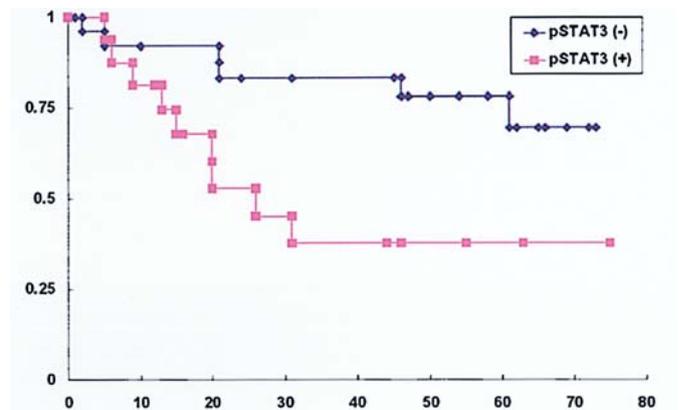


Figure 5. Kaplan-Meier survival curves. The patients with p-STAT3 expressed in carcinoma tissue had significantly worse prognoses ( $^*p < 0.05$ , log-rank test).

Western blot analysis. Statistical analyses of our data showed no correlation between p-STAT3 expression and histological differentiation, but the expression of p-STAT3 correlated

Table II. p-STAT3 expression correlated with vascular invasion in invasive carcinoma (85 cases).

|                    | n  | -          | +          | p-value  |
|--------------------|----|------------|------------|----------|
| Invasive carcinoma | 85 | 35 (43.8%) | 50 (58.8%) |          |
| Venous invasion    |    |            |            | p<0.0001 |
| Present            | 45 | 9 (20.0%)  | 36 (80.0%) |          |
| Absent             | 40 | 26 (65.0%) | 14 (21.5%) |          |
| Lymphatic invasion |    |            |            | p<0.0005 |
| Present            | 62 | 18 (29.0%) | 44 (71.0%) |          |
| Absent             | 23 | 17 (73.9%) | 6 (26.1%)  |          |

with the degree of tumour invasion, lymphatic and venous invasion and the presence of lymph node metastasis.

Activation of STAT3 signaling is reported to up-regulate the expression of various genes involved in cell survival and proliferation, such as the bcl-2 family, cyclin D1 and c-myc (18,20,29,30). However, induction of apoptosis elicited by STAT3 inhibition was not associated with down-regulation of bcl-2 family expression in gastric cancer cell lines. Thus, bcl-2 family genes might not always be involved in the STAT3-induced inhibition of apoptosis in gastric cancer cells (31). Otherwise, in colorectal carcinoma, protein levels of p-STAT3, cyclin D1 and Bcl-XL were increased (32). In the present study, we did not clarify the downstream components of STAT3 activation. In most tumours, the cause of persistent activation of STAT3 is not known. In some myeloma cells and prostate cancer cells, however, constitutively activated STAT3 signaling is due to abnormally regulated IL6-JAK signaling in an autocrine or paracrine manner (10,33,34). Further studies are needed to clarify the cause of constitutive activation of STAT3 signaling in gastric cancer cells.

In the present study, p-STAT3 showed significantly greater expression in advanced cancer than in 'early' cancer. Our results therefore lend support to previous reports of a correlation between p-STAT3 expression and prognosis. Moreover, our results have shown a correlation between p-STAT3 expression and venous and lymphatic invasion and lymph node metastasis. Notably, there is a significant correlation between p-STAT3 expression and venous invasion. Tumour angiogenesis has been considered the most important predictor of overall survival in gastric cancer (35,36).

The prognosis of patients with gastric carcinoma has conventionally been determined by the extent of the primary tumour and the presence or absence of metastasis (25). However, the mechanism of invasion and metastasis of gastric carcinoma has not been fully elucidated. Some reports have shown that the expression of p-STAT3 is correlated with tumour invasion and prognosis (14,31,32,37,38). The role of the JAK/STAT pathway in human gastric carcinoma has not been fully clarified, particularly the relationship between expression of p-STAT3 and the clinicopathological features of gastric carcinoma. Unlike the situation in normal cells and tissues, constitutively activated STATs are detected in a wide variety of human tumours, including carcinomas of the colorectum (14), ovary (22), prostate (24), breast (39) and

leukemia (40). Such aberrant activation of STATs, especially STAT3, is often associated with cell survival, proliferation, and transformation (18,22,41). Thus, as in other types of cancer cells, constitutive activation of STAT3 appears to have an important role in the survival of some kinds of gastric cancer cells by suppressing apoptosis and enhancing cell proliferation (31).

In conclusion, STAT3 is constitutively activated in certain subtypes of human gastric cancer and activation of STAT3 plays an important role in tumour invasion and patient prognosis.

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#### References

1. Neugut AI, Hayek M and Howe G: Epidemiology of gastric cancer. *Semin Oncol* 23: 281-291, 1996.
2. Vogelstein B, Fearon ER, Hamilton SR, Kern SE, Preisinger AC, Leppert M, Nakamura Y, White R, Smits AM and Bos JL: Genetic alterations during colorectal-tumor development. *N Engl J Med* 319: 525-532, 1988.
3. Ming SC and Goldman H: Pathology of the gastrointestinal tract. 2nd edition. Williams & Wilkins, Baltimore, pp632-633, 1998.
4. Goseki N, Koike M and Yoshida M: Histopathologic characteristics of early stage esophageal carcinoma. A comparative study with gastric carcinoma. *Cancer* 69: 1088-1093, 1992.
5. Darnell JE Jr: STATs and gene regulation. *Science* 277: 1630-1635, 1997.
6. Bromberg J: Stat proteins and oncogenesis. *J Clin Invest* 109: 1139-1142, 2002.
7. Levy DE and Darnell JE Jr: Stats: transcriptional control and biological impact. *Nat Rev Mol Cell Biol* 3: 651-662, 2000.
8. Smithgall TE, Briggs SD, Schreiner S, Lerner EC, Cheng H and Wilson MB: Control of myeloid differentiation and survival by Stats. *Oncogene* 19: 2612-2618, 2000.
9. Mora LB, Buettner R, Seigne J, Diaz J, Ahmad N, Garcia R, Bowman T, Falcone R, Fairclough R, Cantor A, Muro-Cacho C, Livingston S, Karras J, Pow-Sang J and Jove R: Constitutive activation of Stat3 in human prostate tumors and cell lines: direct inhibition of Stat3 signaling induces apoptosis of prostate cancer cells. *Cancer Res* 62: 6659-6666, 2002.
10. Catlett-Falcone R, Landowski TH, Oshiro MM, Turkson J, Levitzki A, Savino R, Ciliberto G, Moscinski L, Fernandez-Luna JL, Nunez G, Dalton WS and Jove R: Constitutive activation of Stat3 signaling confers resistance to apoptosis in human U266 myeloma cells. *Immunity* 10: 105-115, 1999.

11. Spiekermann K, Biethahn S, Wilde S, Hiddemann W and Alves F: Constitutive activation of STAT transcription factors in acute myelogenous leukemia. *Eur J Haematol* 67: 63-71, 2001.
12. Dhir R, Ni Z, Lou W, Demiguel F, Grandis JR and Gao AC: Stat3 activation in prostatic carcinomas. *Prostate* 51: 241-246, 2002.
13. Grandis JR, Drenning SD, Zeng Q, Watkins SC, Melhem MF, Endo S, Johnson DE, Huang L, He Y and Kim JD: Constitutive activation of Stat3 signaling abrogates apoptosis in squamous cell carcinogenesis *in vivo*. *Proc Natl Acad Sci USA* 97: 4227-4232, 2000.
14. Kusaba T, Nakayama T, Yamazumi K, Yakata Y, Yoshizaki A, Nagayasu T and Sekine I: Expression of p-STAT3 in human colorectal adenocarcinoma and adenoma; correlation with clinicopathological factors. *J Clin Pathol* 58: 833-838, 2005.
15. Feng DY, Zheng H, Tan Y and Cheng RX: Effect of phosphorylation of MAPK and Stat3 and expression of c-fos and c-jun proteins on hepatocarcinogenesis and their clinical significance. *World J Gastroenterol* 7: 33-36, 2001.
16. Bromberg JF, Horvath CM, Besser D, Lathem WW and Darnell JE Jr: Stat3 activation is required for cellular transformation by v-src. *Mol Cell Biol* 18: 2553-2558, 1998.
17. Bowman T, Broome MA, Sinibaldi D, Wharton W, Pledger WJ, Sedivy JM, Irby R, Yeatman T, Coutneidge SA and Jove R: Stat3-mediated Myc expression is required for Src transformation and PDGF-induced mitogenesis. *Proc Natl Acad Sci USA* 98: 7319-7324, 2001.
18. Bromberg JF, Wrzeszczynska MH, Devgan G, Zhao Y, Pestell RG, Albanese C and Darnell JE Jr: Stat3 as an oncogene. *Cell* 98: 295-303, 1999.
19. Bowman T, Garcia R, Turkson J and Jove R: STATs in oncogenesis. *Oncogene* 19: 2474-2488, 2000.
20. Real PJ, Sierra A, De Juan A, Segovia JC, Lopez-Vega JM and Fernandez-Luna JL: Resistance to chemotherapy via Stat3-dependent overexpression of Bcl-2 in metastatic breast cancer cells. *Oncogene* 21: 7611-7618, 2002.
21. Masuda M, Suzui M, Yasumatu R, Nakashima T, Kuratomi Y, Azuma K, Tomita K, Komiyama S and Weinstein IB: Constitutive activation of signal transducers and activators of transcription 3 correlates with cyclin D1 overexpression and may provide a novel prognostic marker in head and neck squamous cell carcinoma. *Cancer Res* 62: 3351-3355, 2002.
22. Huang M, Page C, Reynolds RK and Lin J: Constitutive activation of stat3 oncogene product in human ovarian carcinoma cells. *Gynecol Oncol* 79: 67-73, 2000.
23. Bartoli M, Gu X, Tsai NT, Venema RC, Brooks SE, Marrero MB and Caldwell RB: Vascular endothelial growth factor activates STAT proteins in aortic endothelial cells. *J Biol Chem* 275: 33189-33192, 2000.
24. Campbell CL, Jiang Z, Savarese DM and Savarese TM: Increased expression of the interleukin-11 receptor and evidence of Stat3 activation in prostate carcinoma. *Am J Pathol* 158: 25-32, 2001.
25. Nishi M, Omori Y and Miwa K (eds): Japanese Classification of Gastric Carcinoma, Japanese Research Society for Gastric Cancer. English edition. Kanehara & Co., Ltd., Tokyo, 1995.
26. Seefeld PH and Barga JA: The spread of cancer of the rectum: invasion of the lymphatic, veins and nerves. *Ann Surg* 118: 76-89, 1943.
27. Talbot IC, Ritchie S, Leighton M, Hughes AO, Bussey HJ and Morson BC: Invasion of veins by carcinoma of rectum: methods of detection, histological features and significance. *Histopathology* 5: 141-163, 1981.
28. Darnell JE Jr, Kerr IM and Stark GR: Jak-STAT pathways and transcriptional activation in response to IFNs and other extracellular signaling proteins. *Science* 264: 1415-1421, 1994.
29. Burke WM, Jin X, Lin HJ, Huang M, Liu R, Reynolds RK and Lin J: Inhibition of constitutively active Stat3 suppresses growth of human ovarian and breast cancer cells. *Oncogene* 20: 7925-7934, 2001.
30. Rahaman SO, Harbor PC, Chernova O, Barnett GH, Vogelbaum MA and Haque SJ: Inhibition of constitutively active Stat3 suppresses proliferation and induces apoptosis in glioblastoma multiforme cells. *Oncogene* 21: 8404-8413, 2002.
31. Kanda N, Seno H, Konda Y, Marusawa H, Kanai M, Nakajima T, Kawashima T, Nanakin A, Sawabu T, Uenoyama Y, Sekikawa A, Kawada M, Suzuki K, Kayahara T, Fukui H, Sawada M and Chiba T: STAT3 is constitutively activated and supports cell survival in association with survivin expression in gastric cancer cells. *Oncogene* 23: 4921-4929, 2004.
32. Ma XT, Wang S, Ye YJ, Du RY, Cui ZR and Somsouk M: Constitutive activation of Stat3 signaling pathway in human colorectal carcinoma. *World J Gastroenterol* 10: 1569-1573, 2004.
33. Giri D, Ozen M and Ittmann M: Interleukin-6 is an autocrine growth factor in human prostate cancer. *Am J Pathol* 159: 2159-2165, 2001.
34. Bisping G, Leo R, Wenning D, Dankbar B, Padro T, Kropff M, Scheffold C, Kroger M, Mesters RM, Berdel WE and Kienast J: Paracrine interactions of basic fibroblast growth factor and interleukin-6 in multiple myeloma. *Blood* 101: 2775-2783, 2003.
35. Brown LF, Berse B, Jackman RW, Tognazzi K, Manseau EJ, Senger DR and Dvorak HF: Expression of vascular permeability factor (vascular endothelial growth factor) and its receptors in adenocarcinomas of the gastrointestinal tract. *Cancer Res* 53: 4727-4735, 1993.
36. Sier CF, Kuben FJ, Ganesh S, Heerding MM, Griffionen G, Hanemaaijer R, van Krieken JH, Lamers CB and Verspaget HW: Tissue levels of matrix metalloproteinases MMP-2 and MMP-9 are related to the overall survival of patients with gastric carcinoma. *Br J Cancer* 74: 413-417, 1996.
37. Kusaba T, Nakayama T, Yamazumi K, Yakata Y, Yoshizaki A, Inoue K, Nagayasu T and Sekine I: Activation of STAT3 is a marker of poor prognosis in human colorectal cancer. *Oncol Rep* 15: 1445-1451, 2006.
38. Rivat C, De Wever O, Bruyneel E, Mareel M, Gespach C and Attoub S: Disruption of STAT3 signaling leads to tumor cell invasion through alterations of homotypic cell-cell adhesion complexes. *Oncogene* 23: 3317-3327, 2004.
39. Berclaz G, Altermatt HJ, Rohrbach V, Siragusa A, Dreher E and Smith PD: EGFR dependent expression of Stat3 (but not STAT1) in breast cancer. *Int J Oncol* 19: 1155-1160, 2001.
40. Xia Z, Sait SN, Baer MR, Barcos M, Donohue KA, Lawrence D, Ford LA, Block AM, Baumann H and Wetzler M: Truncated STAT proteins are prevalent at relapse of acute myeloid leukemia. *Leuk Res* 25: 473-482, 2001.
41. Werner M, Becker KF, Keller G and Hofler H: Gastric adenocarcinoma: pathomorphology and molecular pathology. *J Cancer Res Clin Oncol* 127: 207-216, 2001.