Clinical significance of zinc-finger E-box binding homeobox 1 mRNA levels in peritoneal washing for gastric cancer

NORIMITSU YABUSAKI, SUGURU YAMADA, TOSHIFUMI MURAI, MITSURO KANDA, DAISUKE KOBAYASHI, CHIE TANAKA, TSUTOMU FUJII, GORO NAKAYAMA, HIROYUKI SUGIMOTO, MASAHIKO KOIKE, SHUJI NOMOTO, MICHITAKA FUJIWARA and YASUHIRO KODERA

Department of Gastroenterological Surgery (Surgery II), Nagoya University Graduate School of Medicine, Nagoya, Aichi 466-8550, Japan

Received September 22, 2014; Accepted November 10, 2014

DOI: 10.3892/mco.2014.462

Abstract. Zinc-finger E-box binding homeobox 1 (ZEB1) is an important regulator of epithelial-to-mesenchymal transition and is associated with various types of metastasis. Gastric cancer patients often develop peritoneal carcinomatosis, of which the detection of free cancer cells in the peritoneal washes is an important predictor. We analyzed the correlation of ZEB1 mRNA levels in the peritoneal washing (pZEB1) with clinicopathological variables and survival in 107 gastric cancer patients who underwent surgery and peritoneal washing cytology. Reverse transcription-polymerase chain reaction was performed to quantify pZEB1. The patients were classified into the pZEB1^{High} (n=27) and the pZEB1^{Low} (n=80) groups based on their pZEB1 expression. pZEB1 was statistically correlated with pathological T stage (P=0.03) and vascular involvement (P=0.03). At 5 years, the disease-specific survival was 36.4% for the pZEB1 $^{\rm High}$ group and 64.7% for the pZEB1 $^{\rm Low}$ group (P=0.02), whereas the disease-free survival rate was 46.9% for the pZEB1High group and 83.0% for the pZEB1Low group (P=0.03). When subclassified into 4 categories based on washing cytology and pZEB1, survival was significantly lower in the pZEB1High compared to the pZEB1Low group (cytology-negative group, P=0.01; cytology-positive group, P=0.13). Therefore, pZEB1 may add valuable information to conventional peritoneal washing cytology as a prognostic determinant in gastric cancer.

Correspondence to: Dr Suguru Yamada, Department of Gastroenterological Surgery (Surgery II), Nagoya University Graduate School of Medicine, 65 Tsurumai-cho, Showa-ku, Nagoya, Aichi 466-8550, Japan

E-mail: suguru@med.nagoya-u.ac.jp

Key words: zinc-finger E-box binding homeobox 1, peritoneal washing, epithelial-to-mesenchymal transition, gastric cancer

Introduction

Although the survival of patients with gastric cancer has improved due to the recent advances in treatment, the prognosis of locally advanced or metastatic cancer remains poor (1-3). A proportion of the patients develop recurrences even after curative resection, possibly reflecting the presence of residual cancer cells and micrometastases that had not been detected by the currently available diagnostic technology (4,5). Therefore, the accurate evaluation of microscopic residual disease may lead to more appropriate therapeutic strategies and improvement in survival.

Epithelial-to-mesenchymal transition (EMT) is a critical process during which the adhesion and migration properties of cancer cells change dramatically (6,7). During EMT, the cells lose epithelial polarity and acquire a spindle-shaped, highly motile fibroblastoid phenotype. Various transcription factors are known to trigger EMT (8-10), including zinc-finger E-box binding homeobox 1 (ZEB1), a central EMT mediator (11,12). ZEB1 reportedly affects cancer progression by regulating EMT in gastric, breast, prostate, ovarian and colorectal cancers (13-20).

In gastric cancer, carcinoembryonic antigen (CEA) mRNA levels in peritoneal washing have been reported to be potential predictors of peritoneal recurrence (21,22). Kodera et al reported that the combination of CEA and cytokeratin-20 in peritoneal washes may more accurately predict prognosis (23). ZEB1 expression has also been recently reported as a novel biomarker in cancer tissue that may independently predict overall survival (13,14,24). We recently reported on a significant correlation between ZEB1 expression and diffuse phenotype in gastric cancer (24). Okugawa et al reported that ZEB1 was an independent predictor of peritoneal dissemination in gastric cancer patients and was expressed in disseminated cancer cells in the peritoneum in the same pattern as that seen in the primary lesions (13). Therefore, we hypothesized that the ZEB1 mRNA levels in peritoneal washing (pZEB1) in conjunction with peritoneal washing cytology may predict intraperitoneal recurrence and prognosis.

This study investigated the association of pZEB1 with clinicopathological parameters and prognosis and the potential of pZEB1 as a predictive marker. To the best of our knowledge,

this is the first report on the clinical implication of pZEB1 in gastric cancer.

Materials and methods

Patients. We enrolled 107 consecutive gastric cancer patients who underwent surgical procedures that included collection of peritoneal washing samples at the left subphrenic area at the beginning of surgery, between January, 2005 and August, 2010 at the Department of Gastroenterological Surgery, Nagoya University Hospital, Nagoya, Aichi, Japan. All the patients had histologically confirmed gastric cancer. Of the 107 patients, 4 had received chemotherapy prior to surgery, 2 of whom achieved a complete response. All the patients had been staged according to the Union for International Cancer Control staging criteria for gastric cancer (7th edition, 2009) as follows: 2 patients had stage 0; 12 had stage IA; 11 had stage IB; 7 had stage IIA; 12 had stage IIB; 8 had stage IIIA; 10 had stage IIIB; 10 had stage IIIB; 10 had stage IIIC; and 35 had stage IV disease. Overall, 72 patients underwent curative resection, 35 patients underwent non-curative resection, of whom 2 patients did not receive gastrectomy due to disseminated cancer. All the patients underwent gastrectomy with D2 lymphadenectomy when potentially curative R0 resection was planned. The median follow-up period was 41.9 months (range, 1-106 months). This study was approved by the Ethics Committee of our hospital and signed informed consent was obtained from all the participating patients.

Peritoneal washes. At the beginning of each surgery, 100-200 ml saline was introduced into the left subphrenic area and aspirated soon after gentle stirring. Half of each fluid sample was sent for routine cytopathology with conventional Papanicolaou and Giemsa staining, whereas the other half was used to measure ZEB1 mRNA levels. The sample was centrifuged at 540 x g for 5 min to collect intact cells, rinsed with phosphate-buffered saline, dissolved in ISOGEN-LS RNA extraction buffer (Nippon Gene, Tokyo, Japan) and stored immediately in liquid nitrogen at -80°C until analysis.

Reverse transcription-quantitative polymerase chain reaction (RT-qPCR). Total RNA was isolated from each of the frozen samples with the RNeasy mini kit (Qiagen, Hilden, Germany) according to manufacturer's instructions. cDNA was synthesized using the QuantiTect Reverse Transcription kit (Qiagen, Hilden, Germany) and amplified by PCR primers as follows: ZEB1: 5'-TGCACTGAGTGTGGAAAAGC-3' (forward) and 5'-TGGTGATGCTGAAAGAGACG-3' (reverse), which amplify a 237-bp product. RNA expression was determined using the real-time quantitative PCR method. To quantify and demonstrate the integrity of the isolated RNA, glyceraldehyde-3-phophate dehydrogenase was also analyzed with RT-qPCR using the primer set 5'-AACGGCTCCGGCATGTGCAA-3' (forward) and 5'-GGCTCCTGTGCAGAGAAAGC-3' (reverse). All the PCR reactions were performed as follows: 1 cycle at 50°C for 2 min, 1 cycle at 95°C for 10 min, followed by 40 cycles at 95°C for 15 sec and at 60°C for 60 sec. Real-time detection of the emission intensity of SYBR-Green was performed with an ABI prism 7000 Sequence Detector (Perkin-Elmer

Table I. Patient characteristics.

Characteristics	Patient no.
Age, years (mean ± SD)	63±13.5
Gender	
Male	83
Female	24
Operative method	
TGX	45
DGX	57
PGX	3
Gastrojejunostomy	1
Exploratory laparotomy	1
UICC stage	
0	2
IA	12
IB	11
IIA	7
IIB	12
IIIA	8
IIIB	10
IIIC	10
IV	35

SD, standard deviation; DGX, distal gastrectomy; PGX, proximal gastrectomy; TGX, total gastrectomy; UICC, Union for International Cancer Control.

Applied Biosystems, Foster City, California, USA). qPCR was performed at least 3 times, including a negative no-template control.

Statistical analysis. Correlations between pZEB1 expression and clinicopathological variables were analyzed by the χ^2 and Fisher's exact tests. Disease-specific survival (DSS) and disease-free survival (DFS) were calculated using the Kaplan-Meier method and differences in survival curves were analyzed using the log-rank test. The Cox proportional hazards model was used for multivariate analysis, after relevant prognostic variables had been defined by univariate analysis. Data were analyzed using JMP v10 software (JMP, SAS Institute, Cary, North Carolina, USA). P<0.05 was considered to indicate statistically significant differences.

Results

Patient demographics. The 107 subjects in this study included 83 men and 24 women, with a median age of 63 years (range, 20-84 years) (Table I). Of the 107 patients, 45 underwent total gastrectomy, 57 distal gastrectomy, 3 proximal gastrectomy, 1 gastrojejunostomy and 1 exploratory laparotomy.

Correlation between pZEB1 and clinicopathological factors. pZEB1 was technically detectable in all 107 patients by qPCR.

Table II. Correlation between clinicopathological variables and pZEB1 expression in patients with gastric cancer.

Variables	pZEB1 ^{Low} (n=80)	$pZEB1^{\rm High}\\(n=27)$	P-value
Gender			0.30
Male	64	19	
Female	16	8	
Age, years			0.61
≥65	46	14	
<65	34	13	
Tumor size, cm			0.78
≥5	39	13	
<5	41	12	
Histological type			0.38
Diffuse	52	20	
Intestinal	28	7	
Pathological T stage			0.03^{a}
pT1/2	30	4	
pT3/4	50	23	
Vascular involvement			0.03^{a}
Present	37	18	0.00
Absent	42	7	
Lymphatic vessel involvement			0.20
Present	64	23	3 .2 3
Absent	15	2	
Lymph node metastasis			0.45
Present	52	19	0.15
Absent	28	7	
Liver metastasis			0.16
Present	7	5	0.10
Absent	73	22	
Peritoneal dissemination	, 0		0.22
Present	10	6	0.22
Absent	70	21	
Peritoneal washing cytology	, 0	21	0.46
Present	18	8	0.40
Absent	62	19	
TNM stage	32	17	0.16
I/II	36	8	0.10
III/IV	44	19	
111/1 V	***	19	

^aStatistically significant. pZEB1, zinc-finger E-box binding homeobox 1 mRNA levels in peritoneal washing.

The values ranged from 3.0×10^{-6} to $7.0 \times 10^{-3} \ \mu g/\mu l$ (median, $1.2 \times 10^{-} \ \mu g/\mu l$). The pZEB1 cut-off point was set at the top quartile, which was $3.5 \times 10^{-4} \ \mu g/\mu l$. Accordingly, patients with low pZEB1 expression (< $3.5 \times 10^{-4} \ \mu g/\mu l$) were assigned to the pZEB1^{Low} group (n=80), whereas those with high expression ($\ge 3.5 \times 10^{-4} \ \mu g/\mu l$) were assigned to the pZEB1^{High} group (n=27).

The analysis of pZEB1 expression and various clinicopathological factors (Table II) revealed that pZEB1 was correlated with pathological T stage (P=0.03) and vascular involvement (P=0.03), but not with gender, age, tumor size, histological type, lymphatic vessel involvement, lymph node metastasis, liver metastasis, peritoneal dissemination, peritoneal washing cytology, or TNM stage.

Patient survival by pZEB1 expression. The survival curves of patients with gastric cancer by pZEB1 expression are presented in Fig. 1. DSS was significantly lower in patients with pZEB1^{High} expression compared to those with pZEB1^{Low} expression. The

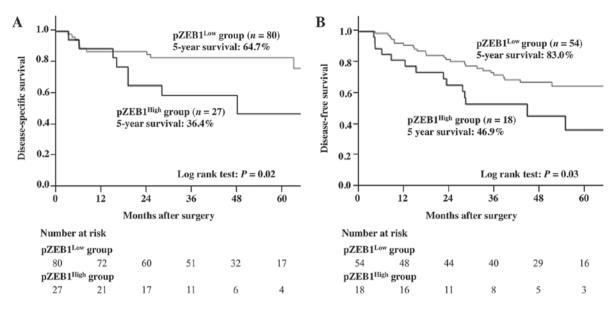


Figure 1. Survival curves for gastric cancer patients by pZEB1 expression status. (A) Disease-specific survival; (B) disease-free survival. The patients with pZEB1^{High} expression exhibited a significantly poorer prognosis compared to those with pZEB1^{Low} expression (A) P=0.02, (B) P=0.03, log-rank test. pZEB1, zinc-finger E-box binding homeobox 1 mRNA levels in peritoneal washing.

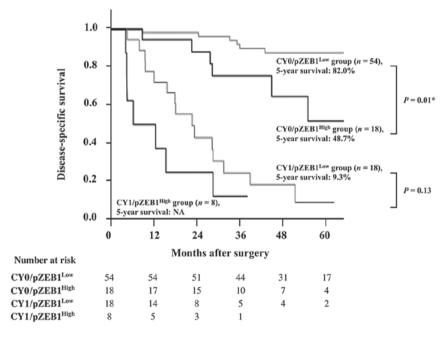


Figure 2. Survival curves for gastric cancer patients based on washing cytology and pZEB1 expression. The patients were subclassified into 4 types based on washing cytology and pZEB1 expression. Disease-specific survival was significantly lower in patients with pZEB1^{High} expression compared to those with pZEB1^{Low} expression. (CY0 group, P=0.01; CY1 group, P=0.13). pZEB1, zinc-finger E-box binding homeobox 1 mRNA levels in peritoneal washing; CY0, negative cytology; CY1, positive cytology; NA, not applicable.

5-year DSS was 36.4% in the pZEB1^{High} group and 64.7% in the pZEB1^{Low} group (P=0.02), whereas the 5-year DFS was 46.9%, in the pZEB1^{High} group and 83.0% in the pZEB1^{Low} group (P=0.03).

The patients were next subclassified into 4 groups according to negative or positive peritoneal washing cytology (CY0 and CY1, respectively) as follows: CY0/pZEB1^{Low}, CY0/pZEB1^{High}, CY1/pZEB1^{Low} and CY1/pZEB1^{High}. In the CY0 group, DSS was significantly lower in the pZEB1^{High} group compared to that in the pZEB1^{Low} group. The 5-year survival rate was 48.7% in the CY0/pZEB1^{High} group and 82.0% in the CY0/pZEB1^{Low}

group (P=0.01). In the CY1 group, DSS was also lower among patients with pZEB1 $^{\rm High}$ expression compared to those with pZEB1 $^{\rm Low}$ expression. The 5-year survival rate was 0% in the CY1/pZEB1 $^{\rm High}$ group and 9.3% in the CY1/pZEB1 $^{\rm Low}$ group (P=0.13) (Fig. 2).

pZEB1 as a predictor of recurrence after surgery. Among the 18 patients who developed recurrences after surgery, 10 patients had pZEB1^{Low} expression and 8 had pZEB1^{High} expression. The recurrence rate in the pZEB1^{High} group (8/27) was significantly higher compared to that in the pZEB1^{Low} group (10/80; P=0.03,

Table III. Correlation of pZEB1 expression status with recurrence of gastric cancer and recurrence site.

A, Correlation of pZEB1 expression with recurrence

Recurrence	pZEB1 ^{Low} (n=54)	pZEB1 ^{High} (n=18)	P-value
Yes	10	8	0.03ª
No	44	10	

B, Correlation of pZEB1 expression with recurrence site

Recurrence site	No.	$pZEB1^{Low/High}$	
Lymph nodes	6	4/2	
Peritoneum	6	2/4	
Liver	5	3/2	
Lung	1	1/0	

aStatistically significant. pZEB1, zinc-finger E-box binding homeobox 1 mRNA levels in peritoneal washing.

Table IV. Characteristics of patients with pZEB1^{High} expression excluding those with stage IV disease.

Patients	Age (yrs)	Gender	DFS	Recurrence site	T stage	Metastasis ^a	Histology
1	62	F	48	Peritoneum	T4a	N3a	Diffuse
2	60	F	28	Peritoneum	T4a	N1	Diffuse
3	55	M	3.2	Peritoneum	T4a	N0	Diffuse
4	55	M	19	Peritoneum	T2	N0	Diffuse
5	63	M	15	Liver	Т3	N3b	Intestinal
6	61	M	6	Liver	Т3	N2	Intestinal
7	71	M	16	Lymph node	Т3	N2	Diffuse
8	75	F	19	Lymph node	T4a	N3a	Diffuse
9	56	M	70	None	Т3	N1	Diffuse
10	70	M	9.5	None	Т3	N0	Intestinal
11	71	F	69	None	T2	N0	Diffuse
12	67	M	27	None	T1a	N0	Intestinal
13	52	M	31	None	Т3	N0	Diffuse
14	72	M	45	None	T4a	N1	Diffuse
15	74	M	35	None	T1b	N0	Intestinal
16	65	M	58	None	T4a	N1	Intestinal
17	35	F	50	None	T4a	N2	Diffuse
18	59	M	43	None	T2	N0	Diffuse

^aMetastatic lymph nodes. pZEB1, zinc-finger E-box binding homeobox 1 mRNA levels in peritoneal washing; DFS, disease-free survival (in months).

Table IIIA). Of these 18 patients 6 developed lymph node metastases, 6 peritoneal metastases, 5 liver metastases and 1 lung metastasis. Of the 6 patients with recurrent peritoneal metastases, 4 were in the pZEB1^{High} group (Table IIIB).

The characteristics of the 18 patients with pZEB1^{High} and CYO, excluding those with stage IV disease, are summarized in Table IV. Among these, 8 patients ultimately developed recurrent metastases (4 in the peritoneum, 2 in the liver and 2 in the lymph nodes).

Prognostic factors of gastric cancer patients by univariate and multivariate analysis. The univariate analysis using the Cox proportional hazards model identified 9 prognostic factors, namely tumor size, T stage, histological type, lymph node metastasis, lymphatic vessel involvement, vascular involvement, peritoneal metastasis, liver metastasis and pZEB1 expression (Table V). However, in the multivariate analysis of these parameters, pZEB1 was not identified as an independent predictor of DSS.

Table V. Univariate and multivariate analysis of clinicopathological factors for disease-specific survival.

Variables		Univariate analy	rsis	Multivariate analysis		
	HR	95% CI	P-value	HR	95% CI	P-value
Gender (female)	1.3	0.6-2.5	0.52			
Age (≥65 years)	1.0	0.6-2.0	0.89			
Tumor size (≥5 cm)	2.3	1.2-4.6	0.01a	1.1	0.5-2.5	0.76
Pathological T stage (pT3/4)	8.4	3.0-34.9	<0.001a	4.4	1.1-24.8	0.04^{a}
Histological type (diffuse)	2.3	1.1-5.4	0.02^{a}	1.3	0.5-3.5	0.57
Lymph node metastasis	4.2	1.8-12.4	<0.001a	2.2	0.7-10.1	0.22
Lymphatic vessel involvement	4.7	1.4-28.9	0.008^{a}	0.4	0.05-3.8	0.40
Vascular involvement	3.9	1.9-8.7	<0.001a	2.0	0.8-5.3	0.13
Peritoneal metastasis	10.6	5.2-21.2	<0.001a	4.1	1.8-9.4	0.001a
Liver metastasis	5.2	2.2-11.1	<0.001a	2.9	0.9-7.9	0.06
$pZEB1^{\rm High}$	2.1	1.1-4.0	0.03^{a}	1.0	0.4-2.1	0.98

^aStatistically significant. HR, hazard ratio; CI, confidence interval; pZEB1, zinc-finger E-box binding homeobox 1 mRNA levels in peritoneal washing.

Discussion

EMT is a process through which epithelial cells attain fibroblastic characteristics, which enable them to invade neighboring tissues (25,26). ETM is regulated by several transcription factors, including Snail, Slug, Twist, CarB-box-binding factor, mesenchyme forkhead 1, Krüppel-like factor and ZEB1 (26-29).

ZEB1 is reportedly a key player in cancer progression (17,30-32). In particular, high expression of ZEB1 in endometrial and colorectal cancers and hepatocellular carcinoma has been associated with poor prognosis (15,33,34). In gastric cancer, ZEB1 expression in cancer tissues has been identified as an independent prognostic factor (13,14). We have also reported a correlation between high ZEB1 expression and diffuse pathological cancer type (24). However, the diffuse type is a known risk factor for peritoneal recurrence in gastric cancer, which supports the findings of Okugawa *et al* (13), who reported that high ZEB1 expression is an independent factor for peritoneal carcinomatosis.

Comparisons of the expression of EMT markers in the primary tumor and corresponding lymph node metastases have been performed for several cancer types (35,36,37). These studies demonstrated that the expression of EMT markers in mature metastatic lymph nodes was lower compared to that in the primary lesions; therefore it was hypothesized that mesenchymal-to-epithelial transition (MET), the reverse phenomenon of EMT, may occur at secondary metastatic sites before the metastasized cells develop into clinically significant metastatic lesions. However, Okugawa et al (13) observed through immunostaining that ZEB1 expression in the peritoneal metastatic sites exhibited the same pattern as that observed in the primary lesions. The role of EMT and MET in the development of peritoneal metastasis may be different from that of nodal metastasis and it may be of value to investigate the EMT status of intraperitoneal cancer cells that likely develop into visible peritoneal deposits. To the best of our knowledge, there are no available studies investigating pZEB1 in gastric cancer patients.

The major finding in this study was that pZEB1 expression was significantly associated with DSS and DFS in patients with gastric cancer. Furthermore, pZEB1 may be a more sensitive diagnostic tool for poor prognosis compared to conventional peritoneal washing cytology, as the RT-qPCR more sensitively detects intraperitoneal free cancer cells and also because positive pZEB1 reflects the capability of the primary tumor to disseminate ZEB1-positive mesenchymally transformed cells into the peritoneal cavity as well as through the hematogeneous and lymphatic metastatic pathways. Although ZEB1 expression in the primary lesion is already known as an independent prognostic factor (13,14,24), pZEB1 expression may also represent a novel marker of a poorer prognosis.

However, our results failed to demonstrate statistical correlations between pZEB1 and peritoneal dissemination and peritoneal recurrence. As stated above, although local ZEB1 production by cancer cells in the peritoneal cavity is the most important factor in pZEB1 expression, the primary pZEB1-high tumor may disseminate metastatic and ZEB1-producing carcinoma cells to any other sites in the body, leading to various other types of metastasis and consequent cancer-related death. Thus, pZEB1 may be correlated with poor prognosis, but not necessarily with peritoneal dissemination. There is also a possibility that a proportion of the patients did actually harbor peritoneal recurrence, but its manifestation was preceded by other types of metastasis that were clinically more relevant. Further investigation is required to elucidate the mechanisms underlying pZEB1 expression in a large population with a long-term follow-up.

In conclusion, pZEB1 may be a predictive marker for poor prognosis or tumor aggressiveness in gastric cancer, similar to ZEB1 expression in primary lesions. pZEB1 may add valuable information to conventional peritoneal washing cytology and, thus, help with the selection of candidates for more aggressive chemotherapies.

References

- Macdonald JS, Smalley SR, Benedetti J, et al: Chemoradiotherapy after surgery compared with surgery alone for adenocarcinoma of the stomach or gastroesophageal junction. N Engl Med 345: 725-730, 2001.
- Cunningham D, Allum WH, Stenning SP, et al: Perioperative chemotherapy versus surgery alone for resectable gastroesophageal cancer. N Engl Med 355: 11-20, 2006.
- 3. Sakuramoto S, Sasako M, Yamaguchi T, *et al*: Adjuvant chemotherapy for gastric cancer with S-1, an oral fluoropyrimidine. N Engl Med 357: 1810-1820, 2007.
- Allum W, Garofalo A, Degiuli M and Schuhmacher C: The first European Union Network of Excellence for Gastric Cancer conference, Rome, Italy, April 2008. Gastric Cancer 12: 56-65, 2009.
- 5. Yonemura Y, Elnemr A, Endou Y, *et al*: Multidisciplinary therapy for treatment of patients with peritoneal carcinomatosis from gastric cancer. World J Gastrointest Oncol 2: 85-97, 2010.
- 6. Thiery JP: Epithelial-mesenchymal transitions in tumour progression. Nat Rev Cancer 2: 442-454, 2002.
- 7. Gotzmann J, Mikula M, Eger A, et al: Molecular aspects of epithelial cell plasticity: implications for local tumor invasion and metastasis. Mutat Res 566: 9-20, 2004.
- Cavallaro U and Christofori G: Cell adhesion and signalling by cadherins and Ig-CAMs in cancer. Nature Rev Cancer 4: 118-132, 2004.
- 9. Tomita K, van Bokhoven A, van Leenders GJ, *et al*: Cadherin switching in human prostate cancer progression. Cancer Res 60: 3650-3654, 2000.
- Rieger-Christ KM, Cain JW, Braasch JW, et al: Expression of classic cadherins type I in urothelial neoplastic progression. Hum Pathol 32: 18-23, 2001.
- Christiansen JJ and Rajasekaran AK: Reassessing epithelial to mesenchymal transition as a prerequisite for carcinoma invasion and metastasis. Cancer Res 66: 8319-8326, 2006.
- Klymkowsky MW and Savagner P: Epithelial-mesenchymal transition: a cancer researcher's conceptual friend and foe. Am J Pathol 174: 1588-1593, 2009.
- 13. Okugawa Y, Toiyama Y, Tanaka K, et al: Clinical significance of zinc finger E-box binding homeobox 1 (ZEB1) in human gastric cancer. J Surg Oncol 106: 280-285, 2012.
- Jia B, Liu H, Kong Q, et al: Overexpression of ZEB1 associated with metastasis and invasion in patients with gastric carcinoma. Mol Cell Biochem 366: 223-229, 2012.
- 15. Zhang GJ, Zhou T, Tian HP, *et al*: High expression of ZEB1 correlates with liver metastasis and poor prognosis in colorectal cancer. Oncol Lett 5: 564-568, 2013.
- Spaderna S, Schmalhofer O, Hlubek F, et al: A transient, EMT-linked loss of basement membranes indicates metastasis and poor survival in colorectal cancer. Gastroenterology 131: 830-840, 2006.
- 17. Eger A, Aigner K, Sonderegger S, *et al*: DeltaEF1 is a transcriptional repressor of E-cadherin and regulates epithelial plasticity in breast cancer cells. Oncogene 24: 2375-2385, 2005.
- Ohira T, Gemmill RM, Ferguson K, et al: WNT7a induces E-cadherin in lung cancer cells. Proc Natl Acad Sci USA 100: 10429-10434, 2003.
- Chua HL, Bhat-Nakshatri P, Clare SE, et al: NF-kappaB represses E-cadherin expression and enhances epithelial to mesenchymal transition of mammary epithelial cells: potential involvement of ZEB-1 and ZEB-2. Oncogene 26: 711-724, 2007.
- Graham TR, Zhau HE, Odero-Marah VA, et al: Insulin-like growth factor-I-dependent up-regulation of ZEB1 drives epithelial-to-mesenchymal transition in human prostate cancer cells. Cancer Res 68: 2479-2488, 2008.

- 21. Abe N, Watanabe T, Toda H, *et al*: Prognostic significance of carcinoembryonic antigen levels in peritoneal washes in patients with gastric cancer. Am J Surg 181: 356-361, 2001.
- 22. Ito S, Nakanishi H, Kodera Y, et al: Prospective validation of quantitative CEA mRNA detection in peritoneal washes in gastric carcinoma patients. Br J Cancer 93: 986-992, 2005.
- 23. Kodera Y, Nakanishi H, Ito S, et al: Prognostic significance of intraperitoneal cancer cells in gastric carcinoma: detection of cytokeratin 20 mRNA in peritoneal washes, in addition to detection of carcinoembryonic antigen. Gastric Cancer 8: 142-148, 2005.
- Murai T, Yamada S, Fuchs BC, et al: Epithelial-to-mesenchymal transition predicts prognosis in clinical gastric cancer. J Surg Oncol 109: 684-689, 2014.
- Schmalhofer O, Brabletz S and Brabletz T: E-cadherin, beta-catenin and ZEB1 in malignant progression of cancer. Cancer Metastasis Rev 28: 151-166, 2009.
- 26. De Wever O, Pauwels P, De Craene B, et al: Molecular and pathological signatures of epithelial-mesenchymal transitions at the cancer invasion front. Histochem Cell Biol 130: 481-494, 2008.
- Waldmann J, Feldmann G, Slater EP, et al: Expression of the zinc-finger transcription factor Snail in adrenocortical carcinoma is associated with decreased survival. Br J Cancer 99: 1900-1907, 2008
- 28. Iwatsuki M, Mimori K, Yokobori T, *et al*: Epithelial-mesenchymal transition in cancer development and its clinical significance. Cancer Sci 101: 293-299, 2010.
- 29. Rosivatz E, Becker I, Specht K, *et al*: Differential expression of the epithelial-mesenchymal transition regulators Snail, SIP1 and Twist in gastric cancer. Am J Pathol 161: 1881-1891, 2002.
- 30. Spaderna S, Schmalhofer O, Wahlbuhl M, *et al*: The transcriptional repressor ZEB1 promotes metastasis and loss of cell polarity in cancer. Cancer Res 68: 537-544, 2008.
- 31. Drake JM, Strohbehn G, Bair TB, *et al*: ZEB1 enhances transendothelial migration and represses the epithelial phenotype of prostate cancer cells. Mol Biol Cell 20: 2207-2217, 2009.
- 32. Takeyama Y, Sato M, Horio M, *et al*: Knockdown of ZEB1, a master epithelial-to-mesenchymal transition (EMT) gene, suppresses anchorage-independent cell growth of lung cancer cells. Cancer Lett 296: 216-224, 2010.
- 33. Singh M, Spoelstra NS, Jean A, *et al*: ZEB1 expression in type I vs type II endometrial cancers: a marker of aggressive disease. Mod Pathol 21: 912-923, 2008.
- 34. Zhou YM, Cao L, Li B, *et al*: Clinicopathological significance of ZEB1 protein in patients with hepatocellular carcinoma. Ann Surg Oncol 19: 1700-1706, 2012.
- 35. Kurahara H, Takao S, Maemura K, *et al*: Epithelial-mesenchymal transition and mesenchymal-epithelial transition via regulation of ZEB-1 and ZEB-2 expression in pancreatic cancer. J Surg Oncol 105: 655-661, 2012.
- 36. Toll A, Masferrer E, Hernández-Ruiz ME, et al: Epithelial to mesenchymal transition markers are associated with an increased metastatic risk in primary cutaneous squamous cell carcinomas but are attenuated in lymph node metastases. J Dermatol Sci 72: 93-102, 2013.
- 37. Aokage K, Ishii G, Ohtaki Y, et al: Dynamic molecular changes associated with epithelial-mesenchymal transition and subsequent mesenchymal-epithelial transition in the early phase of metastatic tumor formation. Int J Cancer 128: 1585-1595, 2011.