

DNA methylation and leukemia susceptibility in China: Evidence from an updated meta-analysis

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Abstract. Mounting evidence supports a role for DNA methylation in the pathogenesis of leukemia; however, there no overview of these results in the Chinese population. The present study performed a comprehensive meta-analysis to establish candidate genes with an altered methylation status in Chinese leukemia patients. Eligible studies were identified through searching the National Center of Biotechnology Information PubMed and Wanfang databases. Studies were pooled and overall odds ratios with corresponding confidence intervals were calculated. A total of 4,325 leukemia patients and 2,010 controls from 94 studies on 53 genes were included in this meta-analysis, and 47 genes were found to be aberrantly methylated in leukemia patients. A further subgroup meta-analysis by leukemia subtype demonstrated that hypermethylation of 5 genes, namely cyclin-dependent kinase (*CDKN2A*), DNA-binding protein inhibitor-4, *CDKN2B*, glioma pathogenesis-related protein 1 and *p73*, contributed to the risk of various subtypes of leukemia. In addition, a strong association between *CDKN2A* and leukemia was identified in Chinese ($P<0.00001$) but not in European patients. The aberrantly methylated genes identified in the present meta-analysis may help elucidate the mechanisms underlying the development of leukemia in Chinese patients.

Introduction

Leukemia is a complex hematological malignancy, characterised by clonal proliferation of malignant hematopoietic stem cells in the blood and bone marrow (1), with a total of 350,000 new cases and 25,700 deaths annually (2). Genetic as well as environmental factors have been suggested to be associated with leukemia, including trisomy 21, gender, cytotoxicity of anticancer drugs, exposure to benzene and ionising radiation (3-6). Leukemia is a heterogeneous disease that comprises acute lymphocytic leukemia (ALL), chronic lymphocytic leukemia (CLL), acute myelogenous leukemia (AML) and chronic myelogenous leukemia (CML). ALL accounts for 81% of childhood leukemia cases, while CLL and AML frequently occur in adults (7).

Racial and ethnic disparities have been identified in the expression of leukemia-related genes, the clinical outcome and the mortality rate of leukemia (8-12). These disparities are likely due to a combination of genetic, environmental and socio-economic factors (13), which may affect epigenetic changes. Epigenetics, such as DNA methylation, have been shown to play an important role in cancer susceptibility (14,15). Therefore, DNA methylation studies may help elucidate these racial and ethnic disparities in leukemia patients.

Aberrant DNA methylation of genes has been shown to be associated with a large number of human malignancies (16,17). Although a recent meta-analysis by our group identified significant associations between a number of aberrantly methylated genes and leukemia (18), the majority of the studies published in the Chinese language are overlooked. Thus, the aim of the present study was to focus on the association of aberrant DNA methylation and leukemia susceptibility in the Chinese population and to investigate ethnic differences in DNA methylation using subgroup meta-analyses.

Materials and methods

Selection of studies. A systematic literature search was performed through the National Center for Biotechnology Information (NCBI) PubMed and Wanfang literature databases, updated until July 10, 2014. The search was performed using the keywords 'leukemia' and 'methylation'. Potentially relevant articles were identified by their titles and abstracts, followed by selection of eligible studies based on full-text analysis.

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Case-control studies on gene methylation in Chinese leukemia patients containing sufficient information on methylation to calculate the odds ratios (ORs) and 95% confidence intervals (CIs) were considered to be eligible. A flow chart of the study selection process is shown in Fig. 1.

Data extraction. The following characteristics were extracted from each eligible study: First author's name, year of publication, disease category and methylation status of cases and controls. All the studies included were reviewed by three authors (D.J., Y.S. and C.X.). For genes with methylation data in other populations, the corresponding data were retrieved and subjected to meta-analyses for comparison with the Chinese population.

Statistical analysis. Review Manager 5.0 software (The Nordic Cochrane Centre, The Cochrane Collaboration, Copenhagen, Denmark) was used for the meta-analysis. The ORs and 95% CIs were calculated to evaluate the association between gene methylation and leukemia. Heterogeneity of the included studies was assessed using I^2 statistics (19). When there was significant heterogeneity ($I^2 > 50\%$), the random-effects model was used to calculate the overall OR and 95% CI; otherwise, the fixed-effects model was applied (20).

Results

Eligible studies. As shown in Fig. 1, 1,477 potentially relevant studies were identified for initial review from the NCBI PubMed and Wanfang literature databases. A total of 1,380 studies were excluded (1,036 irrelevant studies, 227 non-case-control studies and 117 studies with insufficient data). Finally, a total of 94 studies (61 studies on ALL, 76 on AML, 11 on CLL, 31 on CML and 3 other studies on leukemia), were included in the present meta-analysis (31,38,44-135). A total of 53 genes were identified among 4,325 leukemia patients and 2,010 control subjects, of which 19 were reported by only 1 study, 15 by 2 studies and 19 by ≥ 3 studies (Tables I and II). For 47 of these 53 genes, aberrant methylation was proved to be significantly associated with leukemia.

Meta-analysis of the association between cyclin-dependent kinase (CDKN)2A methylation and leukemia. As shown in Fig. 2, 566 cases and 361 controls were included in the meta-analysis of *CDKN2A* methylation. The results indicated that hypermethylation of *CDKN2A* was a risk factor for leukemia ($P < 0.00001$; OR=19.99; 95% CI: 11.37-35.17). Subgroup analysis by type of leukemia revealed that hypermethylation of *CDKN2A* was associated with an increased risk of AML ($P < 0.00001$; OR=17.86; 95% CI: 7.79-40.93), ALL ($P < 0.00001$; OR=24.01; 95% CI: 10.23-56.33) and CLL ($P = 0.04$; OR=15.95; 95% CI: 1.16-218.94), but not of CML ($P = 0.08$). However, there was no significant difference in the association results among different leukemia types ($P = 0.87$).

Meta-analysis of the association between *CDKN2B* methylation and leukemia. The meta-analysis of the association between *CDKN2B* methylation and leukemia included 24 studies comprising 463 cases and 302 controls (Fig. 3).

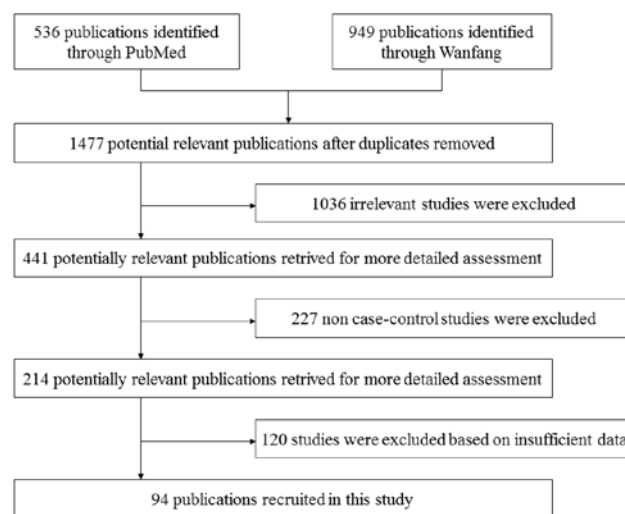


Figure 1. Flow diagram of the study selection process.

The results revealed that hypermethylation of the *CDKN2B* gene was associated with the risk of leukemia ($P < 0.00001$; OR=42.45; 95% CI=22.98-78.42). These 24 studies included 12 studies on ALL, 10 studies on AML and 2 studies on CML. A subtype meta-analysis revealed that *CDKN2B* promoter methylation was a risk factor for AML ($P < 0.00001$; OR=54.11; 95% CI: 21.07-138.93), ALL ($P < 0.00001$; OR=35.76; 95% CI: 14.92-85.69) and CML ($P = 0.004$; OR=27.06; 95% CI: 2.88-254.55). There was no significant difference in the association results among different leukemia types ($P = 0.76$).

Meta-analysis of the association between DNA-binding protein inhibitor-4 (*ID4*) methylation and leukemia. A total of 10 studies were included in the *ID4* methylation analysis (Fig. 4). The meta-analysis revealed that *ID4* methylation was a risk factor for leukemia ($P < 0.00001$; OR=70.08; 95% CI: 24.12-203.64). Hypermethylation of the *ID4* gene was associated with an increased risk of AML ($P < 0.00001$; OR=116.32; 95% CI: 25.40-532.59), ALL ($P < 0.00001$; OR=104.68; 95% CI: 17.27-634.39), CML ($P = 0.002$; OR=20.17; 95% CI: 3.05-133.21) and CLL ($P = 0.002$; OR=693.00; 95% CI: 11.87-40460.96). There was no significant difference in the association results among different leukemia types ($P = 0.33$).

Meta-analysis of the association between glioma pathogenesis-related protein 1 (*GliPR1*) methylation and leukemia. As shown in Fig. 5, the meta-analysis of the association between *GliPR1* methylation and leukemia included 9 studies. The results revealed that *GliPR1* methylation was a risk factor for leukemia ($P < 0.00001$; OR=6.45; 95% CI: 2.88-14.45). Hypermethylation of the *GliPR1* gene was associated with an increased risk of AML ($P < 0.00001$; OR=30.33; 95% CI: 15.83-58.11), ALL ($P < 0.0001$; OR=3.39; 95% CI: 1.88-6.13) and CML ($P = 0.006$; OR=2.49; 95% CI: 1.30-4.77). Moreover, there was a significant difference in the association of *GliPR1* hypermethylation with the risk of leukemia among the different subtypes ($P < 0.00001$).

Table I. Eligible case-control leukemia studies in the Chinese population.

| Gene | No. of studies | Overall OR (95% CI) |
|----------------|----------------|------------------------|
| <i>CDKN2A</i> | 26 | 19.99 (11.37-35.17) |
| <i>CDKN2B</i> | 24 | 42.45 (22.98-78.42) |
| <i>ID4</i> | 10 | 70.08 (24.12-203.64) |
| <i>GliPR1</i> | 9 | 6.45 (2.88-14.45) |
| <i>p73</i> | 7 | 17.07 (6.20-47.02) |
| <i>CT</i> | 6 | 46.85 (12.15-180.65) |
| <i>DAPK</i> | 6 | 17.19 (5.43-54.41) |
| <i>PRB</i> | 6 | 32.10 (9.19-112.16) |
| <i>SFRP5</i> | 5 | 11.45 (3.19-41.12) |
| <i>IGSF4</i> | 4 | 14.41 (3.21-64.72) |
| <i>PRA</i> | 4 | 38.11 (8.14-178.29) |
| <i>RASSF1A</i> | 4 | 22.62 (5.11-100.17) |
| <i>SFRP2</i> | 4 | 30.28 (7.18-127.74) |
| <i>LRP15</i> | 3 | 41.45 (6.63-259.10) |
| <i>RIZ1</i> | 3 | 9.86 (1.84-52.78) |
| <i>SFRP4</i> | 3 | 14.31 (2.77-73.90) |
| <i>WT1</i> | 3 | 0.24 (0.10-0.54) |
| <i>ZO-1</i> | 3 | 99.65 (18.14-547.54) |
| <i>RARA</i> | 3 | 3.46 (0.65-18.39) |
| <i>AR</i> | 2 | 98.28 (5.22-1849.18) |
| <i>CDH13</i> | 2 | 10.18 (1.71-60.56) |
| <i>DDIT3</i> | 2 | 41.35 (5.41-316.24) |
| <i>DKK-1</i> | 2 | 24.28 (3.15-187.26) |
| <i>EDNRB</i> | 2 | 32.57 (3.92-270.80) |
| <i>FANCF</i> | 2 | 7.09 (3.71-13.53) |
| <i>GRAF</i> | 2 | 64.36 (8.67-477.53) |
| <i>HAGE</i> | 2 | 12.23 (1.66-90.39) |
| <i>hPER3</i> | 2 | 85.56 (10.95-668.76) |
| <i>miR-34b</i> | 2 | 74.99 (9.65-582.64) |
| <i>RAGE-1</i> | 2 | 38.88 (5.25-287.87) |
| <i>RUNX3</i> | 2 | 11.91 (1.45-97.86) |
| <i>SFRP1</i> | 2 | 23.65 (3.05-183.57) |
| <i>SHP1</i> | 2 | 11.05 (1.41-86.28) |
| <i>WIF1</i> | 2 | 14.15 (1.78-112.81) |
| <i>AKAP12</i> | 1 | 34.44 (1.85-640.43) |
| <i>CDH1</i> | 1 | 33.00 (1.78-610.61) |
| <i>CEBPZ</i> | 1 | 35.84 (2.12-604.80) |
| <i>DRD4</i> | 1 | 27.26 (1.44-516.59) |
| <i>E-cad</i> | 1 | 33.00 (1.78-610.61) |
| <i>JUNB</i> | 1 | 55.00 (1.86-1622.60) |
| <i>MT3</i> | 1 | 5.76 (1.17-28.24) |
| <i>PLCD1</i> | 1 | 39.38 (2.21-702.41) |
| <i>PRAME</i> | 1 | 42.49 (2.45-737.45) |
| <i>PRDX2</i> | 1 | 36.82 (2.14-633.67) |
| <i>RIL</i> | 1 | 197.19 (11.04-3523.69) |
| <i>SOCS-1</i> | 1 | 34.26 (1.79-654.46) |
| <i>WNT5A</i> | 1 | 121.51 (7.08-2085.83) |
| <i>WWOX</i> | 1 | 41.70 (2.43-715.18) |
| <i>DLC-1</i> | 1 | 17.52 (0.90-342.83) |
| <i>p53</i> | 1 | 19.46 (0.92-411.20) |
| <i>PDLIM4</i> | 1 | 14.23 (0.81-249.59) |

Table I. Continued.

| Gene | No. of studies | Overall OR (95% CI) |
|--------------|----------------|---------------------|
| <i>PTEN</i> | 1 | 16.03 (0.83-308.79) |
| <i>SALL4</i> | 1 | 10.67 (0.59-192.94) |

OR, odds ratio; CI, confidence interval.

Meta-analysis of the association between p73 methylation and leukemia. The meta-analysis of p73 methylation included 7 case-control studies (Fig. 6). The results revealed that hypermethylation of p73 was associated with an increased risk of leukemia ($P<0.00001$; OR=17.07; 95% CI: 6.20-47.02). In addition, the results showed that p73 methylation was a risk factor for AML ($P=0.002$; OR=20.83; 95% CI: 3.01-143.95) and ALL ($P<0.00001$; OR=15.92; 95% CI: 4.87-52.07), while there was no significant difference between the two subtypes.

Subgroup meta-analysis of gene methylation and leukemia by ethnicity. Based on our previous study (18), a further subgroup meta-analysis by ethnicity was performed for *CDKN2A* and *CDKN2B* methylation. Hypermethylation of *CDKN2A* and *CDKN2B* was associated with an increased risk of leukemia in Chinese populations ($P<0.00001$), while only *CDKN2B* was associated with leukemia in Europeans ($P=0.007$) (Figs. 7 and 8). Of note, there was a significant difference between European and Chinese populations regarding the association of *CDKN2A* and *CDKN2B* methylation with leukemia ($P<0.00001$ and $P=0.02$, respectively).

Discussion

In the present study, eligible studies were retrieved from the NCBI PubMed and Wanfang literature databases and a systematic meta-analysis was performed to investigate the association between the methylation status of 53 genes and leukemia, with the aim of providing evidence regarding the role of gene methylation in the pathogenesis of leukemia, particularly in different leukemia subgroups and ethnic groups.

Aberrant gene promoter methylation, occurring in almost every tumor type, is one of several mechanisms of gene inactivation (21). Promoter hypermethylation of tumor suppressor genes often contributes to loss of function and cancer development (22,23). One potential mechanism for hypermethylation-induced silencing is changing the structure of specific binding sites for certain transcriptional regulators (24). Epigenetic silencing of genes by promoter hypermethylation is associated with the loss of tumor suppression, increasing tumor severity and reducing patient survival (25). The present meta-analysis revealed significant changes in the methylation status of the *CDKN2A*, *CDKN2B*, *ID4*, *GliPR1*, *p73* and Wilms' tumor 1 (*WT1*) genes in the major types of leukemia (21,23,26-28).

Numerous studies revealed that *CDKN2A* and *CDKN2B* methylation is frequent during malignant transformation (29-31). As tumor suppressors, *CDKN2A* and *CDKN2B* generate 3 transcript variants (p16^{INK4A}, p14^{ARF} and p15^{INK2B})

Table II. List of methylated genes and associated case-control studies.

| Authors | Year | Gene | Disease | Cases (n) | | Controls (n) | | Refs. |
|--------------------|------|-----------------------|---------|-----------|-------|--------------|-------|-------|
| | | | | Meth | Total | Meth | Total | |
| Hsiao <i>et al</i> | 2008 | <i>CDKN2A</i> | ALL | 11 | 13 | 0 | 8 | (31) |
| Hsiao <i>et al</i> | 2008 | <i>CDKN2A</i> | CLL | 1 | 1 | 0 | 8 | (31) |
| Hsiao <i>et al</i> | 2008 | <i>CDKN2A</i> | AML | 5 | 6 | 0 | 8 | (31) |
| Hsiao <i>et al</i> | 2008 | <i>CDKN2A</i> | CML | 1 | 3 | 0 | 8 | (31) |
| Zheng <i>et al</i> | 2004 | <i>CDKN2A</i> | ALL | 12 | 20 | 0 | 20 | (32) |
| Xiao <i>et al</i> | 2010 | <i>CDKN2A</i> | AML | 7 | 21 | 0 | 16 | (33) |
| Xiao <i>et al</i> | 2010 | <i>CDKN2A</i> | ALL | 7 | 17 | 0 | 16 | (33) |
| Xiao <i>et al</i> | 2010 | <i>CDKN2A</i> | CML | 1 | 7 | 0 | 16 | (33) |
| Xiao <i>et al</i> | 2010 | <i>CDKN2A</i> | CLL | 1 | 6 | 0 | 16 | (33) |
| Yang <i>et al</i> | 2003 | <i>CDKN2A</i> | ALL | 5 | 28 | 0 | 20 | (34) |
| Yang <i>et al</i> | 2003 | <i>CDKN2A</i> | AML | 9 | 43 | 0 | 20 | (34) |
| Song <i>et al</i> | 2004 | <i>CDKN2A</i> | ALL | 5 | 28 | 0 | 20 | (35) |
| Tan <i>et al</i> | 2001 | <i>CDKN2A</i> | AML | 14 | 20 | 0 | 20 | (36) |
| Zhu <i>et al</i> | 2005 | <i>CDKN2A</i> | ALL | 8 | 19 | 0 | 10 | (37) |
| Zhang <i>et al</i> | 2000 | <i>CDKN2A</i> | ALL | 20 | 40 | 0 | 15 | (38) |
| Fan <i>et al</i> | 2007 | <i>CDKN2A</i> | AML | 24 | 58 | 0 | 16 | (39) |
| Fan <i>et al</i> | 2007 | <i>CDKN2A</i> | ALL | 8 | 24 | 0 | 16 | (39) |
| Jiang <i>et al</i> | 2002 | <i>CDKN2A</i> | ALL | 19 | 31 | 0 | 20 | (40) |
| Jiang <i>et al</i> | 2002 | <i>CDKN2A</i> | AML | 14 | 18 | 0 | 20 | (40) |
| Meng <i>et al</i> | 2005 | <i>CDKN2A</i> | AML | 3 | 26 | 0 | 10 | (41) |
| Meng <i>et al</i> | 2005 | <i>CDKN2A</i> | ALL | 2 | 14 | 0 | 10 | (41) |
| Wang <i>et al</i> | 2002 | <i>CDKN2A</i> | ALL | 11 | 15 | 0 | 12 | (42) |
| Yin <i>et al</i> | 2002 | <i>CDKN2A</i> | ALL | 6 | 15 | 0 | 12 | (43) |
| Chen <i>et al</i> | 2003 | <i>CDKN2A (HapII)</i> | AML | 11 | 31 | 0 | 8 | (44) |
| Chen <i>et al</i> | 2003 | <i>CDKN2A (NruI)</i> | AML | 22 | 31 | 2 | 8 | (44) |
| Chen <i>et al</i> | 2003 | <i>CDKN2A (SacII)</i> | AML | 19 | 31 | 1 | 8 | (44) |
| Lin <i>et al</i> | 2012 | <i>CDKN2B</i> | ALL | 17 | 25 | 0 | 10 | (45) |
| Zheng <i>et al</i> | 2004 | <i>CDKN2B</i> | ALL | 18 | 26 | 0 | 20 | (32) |
| Chen <i>et al</i> | 2003 | <i>CDKN2B</i> | AML | 16 | 31 | 0 | 8 | (44) |
| Tan <i>et al</i> | 2001 | <i>CDKN2B</i> | AML | 16 | 20 | 0 | 20 | (36) |
| Zhu <i>et al</i> | 2001 | <i>CDKN2B</i> | ALL | 12 | 21 | 0 | 10 | (46) |
| Zhu <i>et al</i> | 2005 | <i>CDKN2B</i> | ALL | 7 | 19 | 0 | 10 | (37) |
| Shen <i>et al</i> | 2002 | <i>CDKN2B</i> | ALL | 6 | 10 | 0 | 10 | (47) |
| Shen <i>et al</i> | 2002 | <i>CDKN2B</i> | AML | 10 | 25 | 0 | 10 | (47) |
| Fan <i>et al</i> | 2001 | <i>CDKN2B</i> | CML | 5 | 7 | 0 | 20 | (48) |
| Tong <i>et al</i> | 2004 | <i>CDKN2B</i> | AML | 5 | 10 | 0 | 10 | (49) |
| Tong <i>et al</i> | 2004 | <i>CDKN2B</i> | ALL | 4 | 10 | 0 | 10 | (49) |
| Tong <i>et al</i> | 2004 | <i>CDKN2B</i> | CML | 5 | 14 | 0 | 10 | (49) |
| Guo <i>et al</i> | 2000 | <i>CDKN2B</i> | AML | 26 | 31 | 0 | 30 | (50) |
| Qiao <i>et al</i> | 2005 | <i>CDKN2B</i> | AML | 34 | 42 | 0 | 14 | (51) |
| Qiao <i>et al</i> | 2005 | <i>CDKN2B</i> | ALL | 9 | 14 | 0 | 14 | (51) |
| Chen <i>et al</i> | 2000 | <i>CDKN2B</i> | ALL | 5 | 10 | 0 | 10 | (52) |
| Yin <i>et al</i> | 2003 | <i>CDKN2B</i> | ALL | 6 | 15 | 0 | 12 | (53) |
| Yin <i>et al</i> | 2003 | <i>CDKN2B</i> | AML | 17 | 22 | 0 | 12 | (53) |
| Meng <i>et al</i> | 2005 | <i>CDKN2B</i> | AML | 24 | 26 | 0 | 10 | (41) |
| Meng <i>et al</i> | 2005 | <i>CDKN2B</i> | ALL | 10 | 14 | 0 | 10 | (41) |
| Wu <i>et al</i> | 2013 | <i>CDKN2B</i> | ALL | 14 | 14 | 0 | 14 | (54) |
| Wu <i>et al</i> | 2013 | <i>CDKN2B</i> | AML | 6 | 14 | 0 | 14 | (54) |
| Wang <i>et al</i> | 2002 | <i>CDKN2B</i> | ALL | 7 | 10 | 0 | 7 | (55) |
| Wang <i>et al</i> | 2002 | <i>CDKN2B</i> | AML | 22 | 33 | 0 | 7 | (55) |

Table II. Continued.

| Authors | Year | Gene | Disease | Cases (n) | | Controls (n) | | Refs. |
|--------------------|------|--------|---------|-----------|-------|--------------|-------|-------|
| | | | | Meth | Total | Meth | Total | |
| Zhao <i>et al</i> | 2008 | ID4 | AML | 15 | 32 | 0 | 18 | (56) |
| Wang <i>et al</i> | 2010 | ID4 | CML | 6 | 48 | 0 | 10 | (57) |
| Liu <i>et al</i> | 2011 | ID4 | AML | 39 | 46 | 0 | 10 | (58) |
| Jie <i>et al</i> | 2012 | ID4 | AML | 21 | 23 | 1 | 20 | (59) |
| Jie <i>et al</i> | 2012 | ID4 | ALL | 9 | 13 | 1 | 20 | (59) |
| Jie <i>et al</i> | 2012 | ID4 | CML | 9 | 11 | 1 | 20 | (59) |
| Zhao <i>et al</i> | 2005 | ID4 | AML | 21 | 25 | 0 | 49 | (60) |
| Zhao <i>et al</i> | 2005 | ID4 | CML | 2 | 4 | 0 | 49 | (60) |
| Zhao <i>et al</i> | 2005 | ID4 | ALL | 12 | 14 | 0 | 49 | (60) |
| Zhao <i>et al</i> | 2005 | ID4 | CLL | 3 | 3 | 0 | 49 | (60) |
| Xiao <i>et al</i> | 2011 | GLIPR1 | AML | 58 | 70 | 14 | 93 | (61) |
| Xiao <i>et al</i> | 2011 | GLIPR1 | CML | 11 | 40 | 14 | 93 | (61) |
| Xiao <i>et al</i> | 2011 | GLIPR1 | ALL | 22 | 57 | 14 | 93 | (61) |
| Liang <i>et al</i> | 2009 | GLIPR1 | AML | 44 | 54 | 5 | 35 | (62) |
| Liang <i>et al</i> | 2009 | GLIPR1 | CML | 11 | 40 | 5 | 35 | (62) |
| Liang <i>et al</i> | 2009 | GLIPR1 | ALL | 18 | 48 | 5 | 35 | (62) |
| Jie <i>et al</i> | 2012 | GLIPR1 | AML | 22 | 23 | 4 | 20 | (59) |
| Jie <i>et al</i> | 2012 | GLIPR1 | ALL | 5 | 13 | 4 | 20 | (59) |
| Jie <i>et al</i> | 2012 | GLIPR1 | CML | 6 | 11 | 4 | 20 | (59) |
| Zhang <i>et al</i> | 2010 | p73 | AML | 1 | 30 | 1 | 123 | (63) |
| Zhang <i>et al</i> | 2010 | p73 | ALL | 10 | 112 | 1 | 123 | (63) |
| Zhang <i>et al</i> | 2012 | p73 | AML | 21 | 58 | 0 | 31 | (64) |
| Wu <i>et al</i> | 2008 | p73 | ALL | 10 | 30 | 0 | 16 | (65) |
| Liu <i>et al</i> | 2005 | p73 | ALL | 10 | 26 | 0 | 18 | (66) |
| Xu <i>et al</i> | 2005 | p73 | ALL | 12 | 42 | 0 | 10 | (67) |
| Yu <i>et al</i> | 2014 | p73 | ALL | 10 | 32 | 0 | 30 | (68) |
| Xie <i>et al</i> | 2003 | CT | AML | 25 | 31 | 0 | 14 | (69) |
| Xie <i>et al</i> | 2003 | CT | CML | 13 | 45 | 0 | 14 | (69) |
| Tang <i>et al</i> | 2001 | CT | CLL | 1 | 3 | 0 | 30 | (70) |
| Tang <i>et al</i> | 2001 | CT | CML | 8 | 10 | 0 | 30 | (70) |
| Tang <i>et al</i> | 2001 | CT | ALL | 12 | 14 | 0 | 30 | (70) |
| Wang <i>et al</i> | 1998 | CT | CML | 13 | 31 | 0 | 10 | (71) |
| Qian <i>et al</i> | 2010 | DAPK | AML | 82 | 112 | 0 | 15 | (72) |
| Niu <i>et al</i> | 2014 | DAPK | AML | 33 | 102 | 0 | 7 | (73) |
| Niu <i>et al</i> | 2014 | DAPK | ALL | 8 | 17 | 0 | 7 | (73) |
| Zhao <i>et al</i> | 2009 | DAPK | AML | 3 | 60 | 0 | 17 | (74) |
| Zhao <i>et al</i> | 2009 | DAPK | ALL | 16 | 55 | 0 | 17 | (74) |
| Qian J | 2008 | DAPK | CML | 25 | 49 | 0 | 13 | (75) |
| Lin W | 2010 | PRB | CLL | 18 | 27 | 0 | 15 | (76) |
| Wu B | 2008 | PRB | CLL | 5 | 9 | 0 | 5 | (77) |
| Zhang <i>et al</i> | 2003 | PRB | ALL | 6 | 11 | 0 | 10 | (78) |
| Zhang <i>et al</i> | 2003 | PRB | CLL | 6 | 8 | 0 | 10 | (78) |
| Zhang <i>et al</i> | 2003 | PRB | AML | 9 | 15 | 0 | 10 | (78) |
| Zhang <i>et al</i> | 2003 | PRB | CML | 6 | 10 | 0 | 10 | (78) |
| Shi <i>et al</i> | 2011 | SFRP5 | AML | 10 | 99 | 1 | 70 | (79) |
| Wang <i>et al</i> | 2012 | SFRP5 | CML | 3 | 3 | 0 | 6 | (80) |
| Wang <i>et al</i> | 2012 | SFRP5 | AML | 4 | 7 | 0 | 6 | (80) |
| Xu <i>et al</i> | 2010 | SFRP5 | AML | 6 | 59 | 0 | 20 | (81) |
| Xu <i>et al</i> | 2010 | SFRP5 | ALL | 9 | 28 | 0 | 20 | (81) |
| Li <i>et al</i> | 2004 | IGSF4 | AML | 16 | 29 | 0 | 8 | (82) |

Table II. Continued.

| Authors | Year | Gene | Disease | Cases (n) | | Controls (n) | | Refs. |
|--------------------|------|----------------|----------|-----------|-------|--------------|-------|-------|
| | | | | Meth | Total | Meth | Total | |
| Li <i>et al</i> | 2004 | <i>IGSF4</i> | ALL | 12 | 21 | 0 | 8 | (82) |
| Li <i>et al</i> | 2004 | <i>IGSF4</i> | CML | 6 | 18 | 0 | 8 | (82) |
| Li <i>et al</i> | 2004 | <i>IGSF4</i> | CLL | 2 | 7 | 0 | 8 | (82) |
| Zhang <i>et al</i> | 2003 | <i>PRA</i> | ALL | 7 | 11 | 0 | 10 | (78) |
| Zhang <i>et al</i> | 2003 | <i>PRA</i> | CLL | 5 | 8 | 0 | 10 | (78) |
| Zhang <i>et al</i> | 2003 | <i>PRA</i> | AML | 10 | 15 | 0 | 10 | (78) |
| Zhang <i>et al</i> | 2003 | <i>PRA</i> | CML | 7 | 10 | 0 | 10 | (78) |
| Chen <i>et al</i> | 2012 | <i>RASSF1A</i> | AML | 2 | 24 | 0 | 60 | (83) |
| Chen <i>et al</i> | 2012 | <i>RASSF1A</i> | CML | 1 | 23 | 0 | 60 | (83) |
| Chen <i>et al</i> | 2012 | <i>RASSF1A</i> | ALL | 5 | 19 | 0 | 60 | (83) |
| Chen <i>et al</i> | 2012 | <i>RASSF1A</i> | CLL | 4 | 20 | 0 | 60 | (83) |
| Song <i>et al</i> | 2011 | <i>SFRP2</i> | CML | 25 | 38 | 0 | 13 | (84) |
| Shi <i>et al</i> | 2011 | <i>SFRP2</i> | AML | 27 | 99 | 0 | 70 | (79) |
| Xu <i>et al</i> | 2010 | <i>SFRP2</i> | AML | 14 | 59 | 0 | 20 | (81) |
| Xu <i>et al</i> | 2010 | <i>SFRP2</i> | ALL | 8 | 28 | 0 | 20 | (81) |
| Dou <i>et al</i> | 2004 | <i>LRP15</i> | AML | 37 | 53 | 0 | 9 | (85) |
| Dou <i>et al</i> | 2004 | <i>LRP15</i> | ALL | 15 | 20 | 0 | 9 | (85) |
| Dou <i>et al</i> | 2004 | <i>LRP15</i> | CLL | 1 | 2 | 0 | 9 | (85) |
| Yao <i>et al</i> | 2010 | <i>RIZ1</i> | AML | 11 | 37 | 0 | 15 | (86) |
| Cai <i>et al</i> | 2012 | <i>RIZ1</i> | ALL | 15 | 64 | 0 | 9 | (87) |
| Cai <i>et al</i> | 2012 | <i>RIZ1</i> | AML | 12 | 32 | 0 | 9 | (87) |
| Shi <i>et al</i> | 2011 | <i>SFRP4</i> | AML | 17 | 99 | 0 | 70 | (88) |
| Xu <i>et al</i> | 2010 | <i>SFRP4</i> | AML | 4 | 59 | 0 | 20 | (81) |
| Xu <i>et al</i> | 2010 | <i>SFRP4</i> | ALL | 7 | 28 | 0 | 20 | (81) |
| Jie <i>et al</i> | 2012 | <i>WT1</i> | AML | 8 | 23 | 15 | 20 | (59) |
| Jie <i>et al</i> | 2012 | <i>WT1</i> | ALL | 4 | 13 | 15 | 20 | (59) |
| Jie <i>et al</i> | 2012 | <i>WT1</i> | CML | 7 | 11 | 15 | 20 | (59) |
| Dou <i>et al</i> | 2009 | <i>ZO-1</i> | Leukemia | 7 | 10 | 0 | 10 | (89) |
| Wang <i>et al</i> | 2008 | <i>ZO-1</i> | AML | 32 | 52 | 0 | 40 | (90) |
| Wang <i>et al</i> | 2008 | <i>ZO-1</i> | ALL | 17 | 29 | 0 | 40 | (90) |
| Chim <i>et al</i> | 2005 | <i>RARA</i> | APL | 25 | 63 | 0 | 8 | (91) |
| Chim <i>et al</i> | 2005 | <i>RARA</i> | AML | 1 | 50 | 0 | 8 | (91) |
| Chim <i>et al</i> | 2005 | <i>RARA</i> | ALL | 1 | 25 | 0 | 8 | (91) |
| Wang <i>et al</i> | 2007 | <i>AR</i> | ALL | 4 | 4 | 0 | 3 | (92) |
| Wang <i>et al</i> | 2007 | <i>AR</i> | AML | 11 | 11 | 0 | 3 | (92) |
| Wang <i>et al</i> | 2009 | <i>CDH13</i> | CML | 4 | 8 | 0 | 5 | (93) |
| Liu <i>et al</i> | 2013 | <i>CDH13</i> | AML | 23 | 44 | 1 | 10 | (94) |
| Wang <i>et al</i> | 2009 | <i>DDIT3</i> | AML | 62 | 133 | 0 | 16 | (95) |
| Wang <i>et al</i> | 2009 | <i>DDIT3</i> | CML | 39 | 59 | 0 | 16 | (95) |
| Zhu <i>et al</i> | 2012 | <i>DKK-1</i> | ALL | 14 | 34 | 0 | 20 | (96) |
| Zhu <i>et al</i> | 2012 | <i>DKK-1</i> | AML | 10 | 31 | 0 | 20 | (96) |
| Yuan <i>et al</i> | 2010 | <i>EDNRB</i> | AML | 15 | 22 | 0 | 8 | (97) |
| Yuan <i>et al</i> | 2010 | <i>EDNRB</i> | ALL | 11 | 17 | 0 | 8 | (97) |
| Yu <i>et al</i> | 2008 | <i>FANCF</i> | AML | 41 | 58 | 7 | 20 | (98) |
| Deng <i>et al</i> | 2009 | <i>FANCF</i> | AML | 85 | 111 | 11 | 42 | (99) |
| Qian <i>et al</i> | 2010 | <i>GRAF</i> | AML | 87 | 132 | 0 | 20 | (100) |
| Qian <i>et al</i> | 2010 | <i>GRAF</i> | CML | 34 | 61 | 0 | 20 | (100) |
| Chen <i>et al</i> | 2012 | <i>HAGE</i> | AML | 32 | 214 | 0 | 24 | (101) |
| Chen <i>et al</i> | 2012 | <i>HAGE</i> | CML | 22 | 87 | 0 | 24 | (101) |
| Li <i>et al</i> | 2011 | <i>hPER3</i> | CML | 12 | 29 | 0 | 40 | (102) |

Table II. Continued.

| Authors | Year | Gene | Disease | Cases (n) | | Controls (n) | | Refs. |
|---------------------|------|----------------|----------|-----------|-------|--------------|-------|-------|
| | | | | Meth | Total | Meth | Total | |
| Wang <i>et al</i> | 2011 | <i>hPER3</i> | AML | 116 | 206 | 0 | 40 | (103) |
| Wang <i>et al</i> | 2013 | <i>miR-34B</i> | ALL | 24 | 31 | 0 | 23 | (104) |
| Wang <i>et al</i> | 2013 | <i>miR-34B</i> | AML | 8 | 19 | 0 | 23 | (104) |
| Chai <i>et al</i> | 2013 | <i>RAGE-1</i> | AML | 52 | 121 | 0 | 25 | (105) |
| Chai <i>et al</i> | 2013 | <i>RAGE-1</i> | CML | 33 | 76 | 0 | 25 | (105) |
| Lin <i>et al</i> | 2008 | <i>RUNX3</i> | AML | 7 | 23 | 0 | 10 | (106) |
| Lin <i>et al</i> | 2008 | <i>RUNX3</i> | ALL | 7 | 17 | 0 | 10 | (106) |
| Xu <i>et al</i> | 2010 | <i>SFRP1</i> | AML | 20 | 59 | 0 | 20 | (81) |
| Xu <i>et al</i> | 2010 | <i>SFRP1</i> | ALL | 11 | 28 | 0 | 20 | (81) |
| Chim <i>et al</i> | 2004 | <i>SHP1</i> | AML | 26 | 50 | 0 | 8 | (107) |
| Chim <i>et al</i> | 2004 | <i>SHP1</i> | ALL | 6 | 25 | 0 | 8 | (107) |
| Wang <i>et al</i> | 2011 | <i>WIF1</i> | AML | 11 | 34 | 0 | 15 | (108) |
| Wang <i>et al</i> | 2011 | <i>WIF1</i> | ALL | 6 | 21 | 0 | 15 | (108) |
| Liu <i>et al</i> | 2008 | <i>AKAP12</i> | ALL | 20 | 32 | 0 | 10 | (109) |
| Gao <i>et al</i> | 2006 | <i>CDH1</i> | AML | 38 | 55 | 0 | 7 | (110) |
| Yao <i>et al</i> | 2011 | <i>CEBPZ</i> | AML | 62 | 133 | 0 | 20 | (111) |
| Guan <i>et al</i> | 2008 | <i>DLC-1</i> | ALL | 21 | 34 | 0 | 5 | (112) |
| Yu <i>et al</i> | 2000 | <i>DRD4</i> | AML | 16 | 27 | 0 | 9 | (113) |
| Gao <i>et al</i> | 2006 | <i>E-cad</i> | AML | 38 | 55 | 0 | 7 | (110) |
| Wang <i>et al</i> | 2009 | <i>JUNB</i> | CML | 7 | 8 | 0 | 5 | (93) |
| Tao <i>et al</i> | 2014 | <i>MT3</i> | AML | 16 | 41 | 2 | 20 | (114) |
| Zheng <i>et al</i> | 2007 | <i>p53</i> | ALL | 5 | 11 | 0 | 11 | (115) |
| Li <i>et al</i> | 2013 | <i>PDLIM4</i> | CML | 13 | 59 | 0 | 24 | (116) |
| Song <i>et al</i> | 2012 | <i>PLCD1</i> | CML | 23 | 41 | 0 | 15 | (117) |
| Yao <i>et al</i> | 2013 | <i>PRAME</i> | CML | 28 | 55 | 0 | 20 | (118) |
| Yan <i>et al</i> | 2012 | <i>PRDX2</i> | AML | 17 | 55 | 0 | 40 | (119) |
| Yang <i>et al</i> | 2007 | <i>PTEN</i> | ALL | 5 | 22 | 0 | 25 | (120) |
| Du <i>et al</i> | 2013 | <i>RIL</i> | AML | 50 | 60 | 0 | 20 | (121) |
| Jiao <i>et al</i> | 2013 | <i>SALL4</i> | AML | 9 | 45 | 0 | 20 | (122) |
| Zhuang <i>et al</i> | 2011 | <i>SOCS-1</i> | AML | 15 | 24 | 0 | 10 | (123) |
| Deng <i>et al</i> | 2011 | <i>WNT5A</i> | Leukemia | 47 | 68 | 0 | 27 | (124) |
| Zhang <i>et al</i> | 2012 | <i>WWOX</i> | AML | 23 | 58 | 0 | 31 | (64) |

ALL, acute lymphocytic leukemia; CLL, chronic lymphocytic leukemia; AML, acute myelogenous leukemia; CML, chronic myelogenous leukemia; Meth, methylated.

according to differences in the first exons and control the progression of cells from the G1 to the S phase (29,125). The present meta-analysis demonstrated that hypermethylation of *CDKN2A* and *CDKN2B* are risk factors for leukemia. According to the subgroup meta-analysis, hypermethylation of *CDKN2A* was significantly associated with AML, ALL and CLL, but not with CML, while *CDKN2B* hypermethylation was significantly associated with AML, ALL and CML. The lack of association of *CDKN2A* with CML may be attributed to the limited sample included in the meta-analyses (CML power, 6.4%; and CLL power, 6.3%).

The ID4 protein is a member of the dominant-negative basic helix-loop-helix transcription factor family that lacks DNA-binding activity (126) and has a tumor suppressor

function. The promoter of *ID4* was reported to be consistently methylated to various degrees in CLL and a univariate analysis demonstrated that increased promoter methylation of *ID4* was correlated with shortened patient survival (127). Previous studies also reported that *ID4* gene promoter hypermethylation was highly correlated with acute leukemia and may reflect the malignant degree of AML (128,129). The results of the present meta-analysis demonstrated that methylation of the *ID4* gene was associated with an increased risk of leukemia, particularly CML.

The GliPR1 protein, encoded by the *GliPR1* gene, has been identified as an epigenetically regulated tumor suppressor in prostate cancer and AML. *GliPR1* may serve as a marker for monitoring disease activity in AML patients during

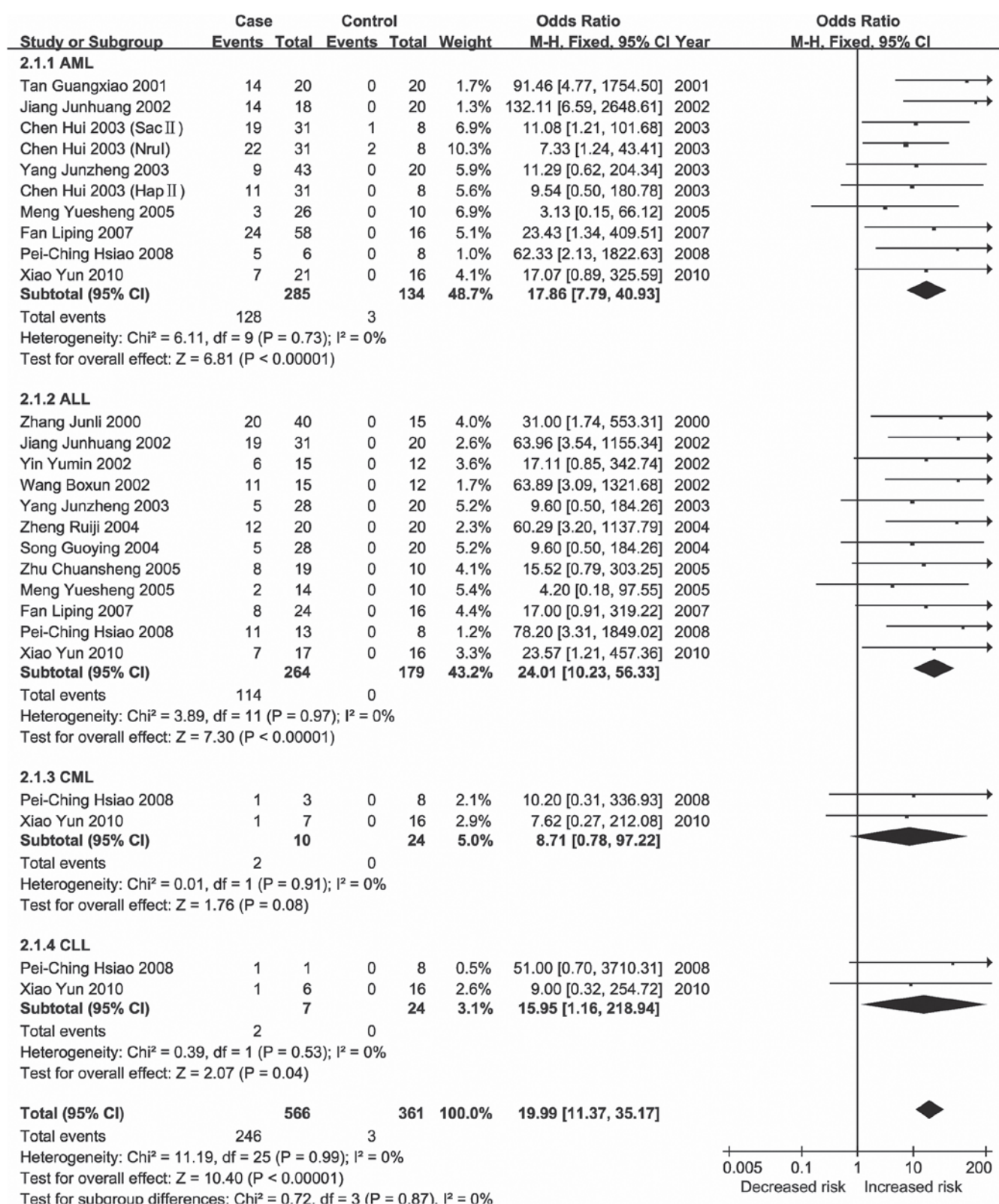


Figure 2. Meta-analyses of aberrantly methylated cyclin-dependent kinase 2A gene in leukemia. ALL, acute lymphocytic leukemia; CLL, chronic lymphocytic leukemia; AML, acute myelogenous leukemia; CML, chronic myelogenous leukemia; M-H, Mantel-Haenszel model; CI, confidence interval; df, degree of freedom.

therapy (61,130). Moreover, *GliPR1* expression was found to be significantly increased in bone marrow samples of AML patients, while being markedly reduced in ALL, unchanged in myelodysplastic syndrome and marginally decreased in CLL and CML (131). The present meta-analysis identified

hypermethylation of the *GliPR1* promoter as a risk factor for leukemia in the Chinese population.

p73, a homologue of the *p53* tumor suppressor family, is involved in neurogenesis, sensory pathways, immunity, inflammation and tumorigenesis (132). Furthermore, *p73*

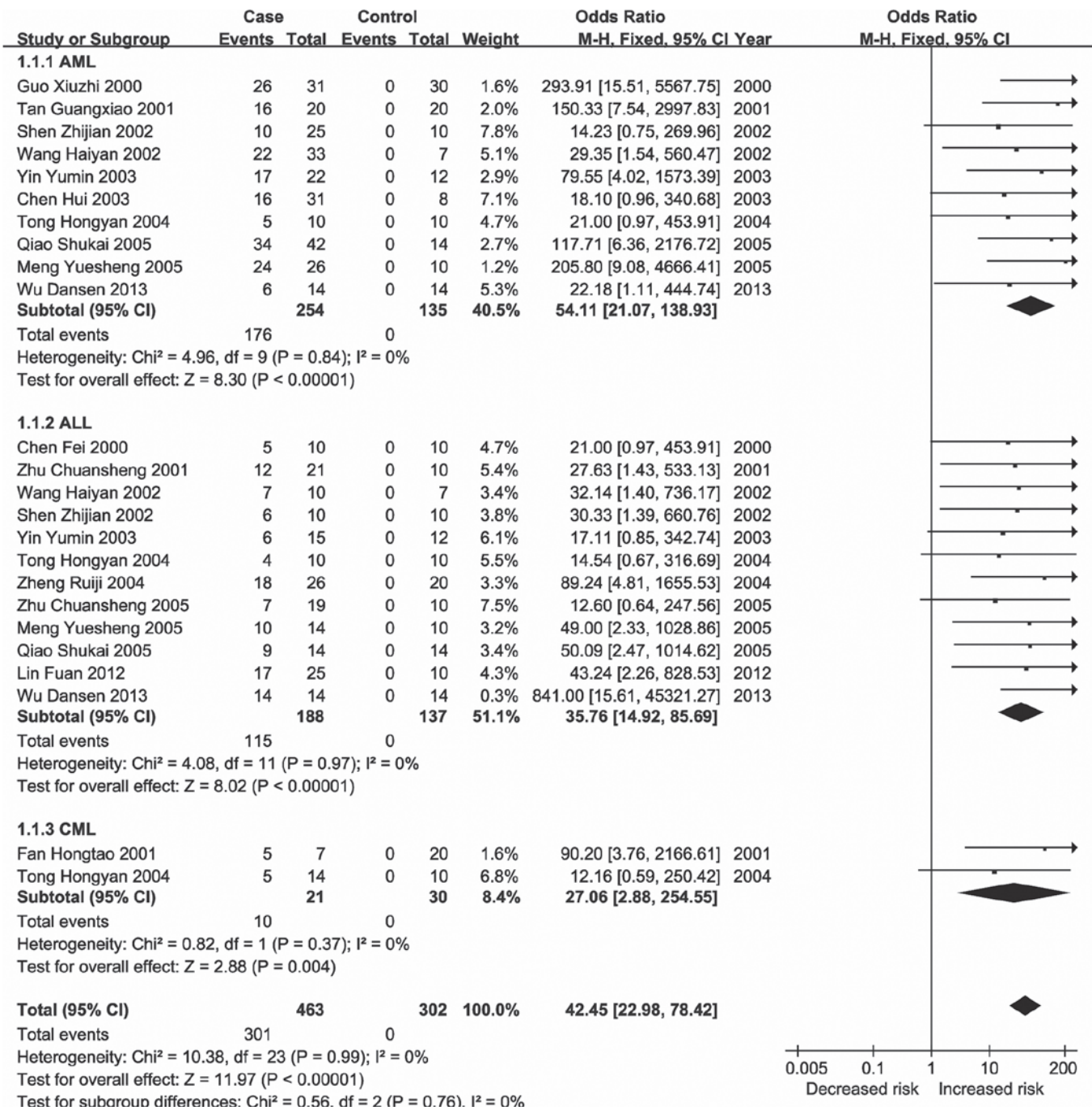


Figure 3. Meta-analyses of aberrantly methylated cyclin-dependent kinase 2B gene in leukemia. ALL, acute lymphocytic leukemia; AML, acute myelogenous leukemia; CML, chronic myelogenous leukemia; M-H, Mantel-Haenszel model; CI, confidence interval; df, degree of freedom.

hypermethylation resulting in its deactivation is frequently observed in malignant lymphoproliferative disorders, particularly ALL (21). In line with these results, the present meta-analysis also identified *p73* hypermethylation as a risk factor for leukemia in the Chinese population.

The *WT1* gene encodes a zinc finger transcription factor that is an RNA-binding protein with important roles in the development of several organs and tissues. *WT1* has been reported to have tumor suppressor as well as oncogenic activity; however, the reasons and mechanisms underlying these opposing functions remain to be fully elucidated (133). The present study demonstrated that *WT1* hypermethylation played a protective role against the progression of leukemia.

Previous studies have reported that the risk of hematological malignancies varies significantly among different ethnic groups (9,13,134,135). The present meta-analysis indicated that there was no association between *CDKN2A* methylation and the risk of leukemia ($P=0.16$) in Europeans, while a significant association was observed in Chinese populations ($P<0.00001$). A significant difference in the association of *CDKN2A* methylation with leukemia was observed between European and Chinese populations ($P<0.00001$). This result may provide molecular evidence to guide future individualization of chemotherapy for leukemia, although further research is required to elucidate the precise nature of the ethnic differences in leukemia.

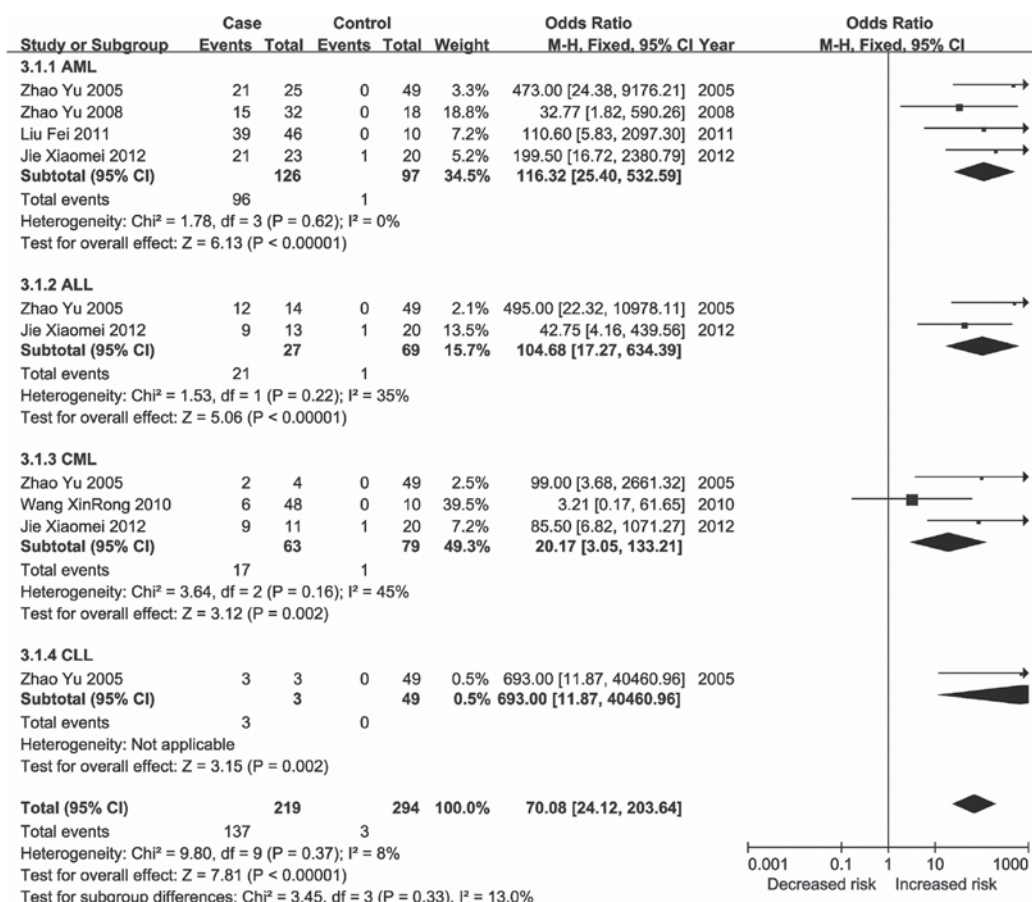


Figure 4. Meta-analyses of aberrantly methylated DNA-binding protein inhibitor-4 gene in leukemia. ALL, acute lymphocytic leukemia; CLL, chronic lymphocytic leukemia; AML, acute myelogenous leukemia; CML, chronic myelogenous leukemia; M-H, Mantel-Haenszel model; CI, confidence interval; df, degree of freedom.

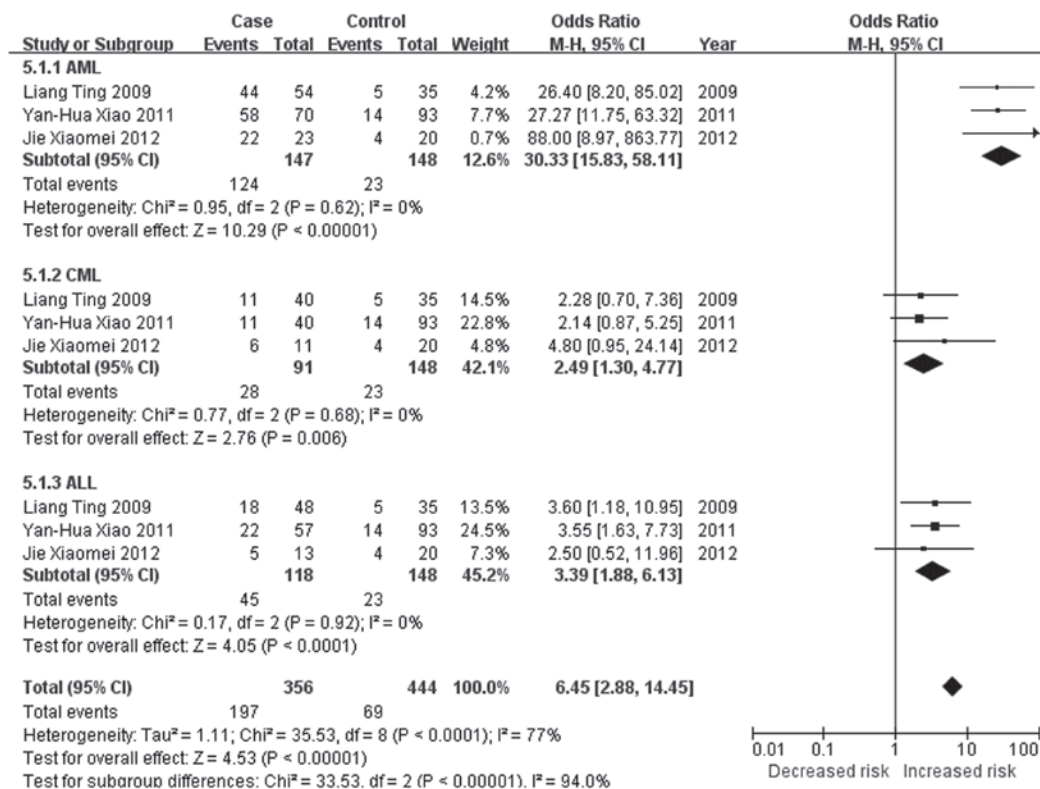


Figure 5. Meta-analyses of aberrantly methylated glioma pathogenesis-related protein 1 gene in leukemia. ALL, acute lymphocytic leukemia; AML, acute myelogenous leukemia; CML, chronic myelogenous leukemia; M-H, Mantel-Haenszel model; CI, confidence interval; df, degree of freedom.

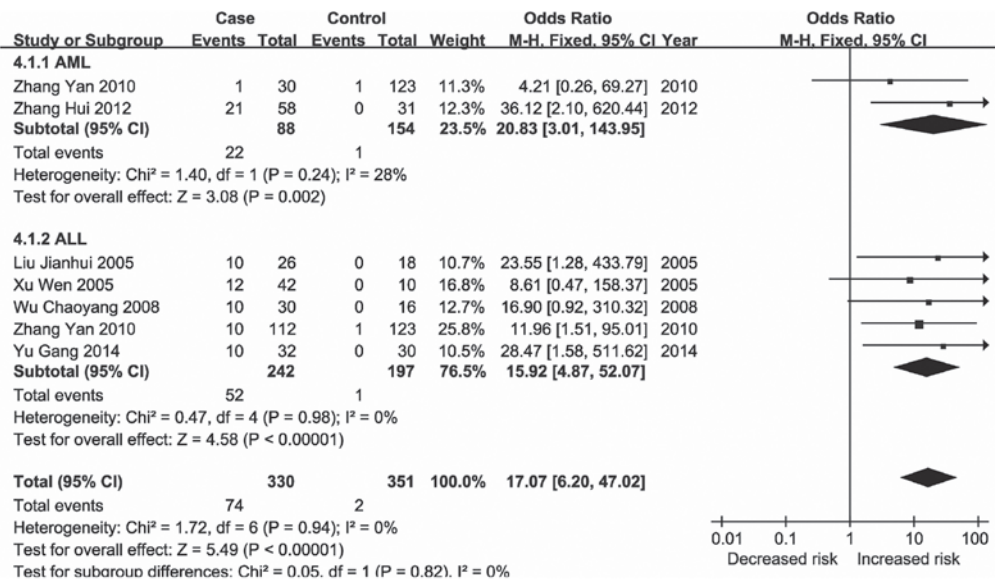


Figure 6. Meta-analyses of aberrantly methylated *p73* in leukemia. ALL, acute lymphocytic leukemia; AML, acute myelogenous leukemia; M-H, Mantel-Haenszel model; CI, confidence interval; df, degree of freedom.

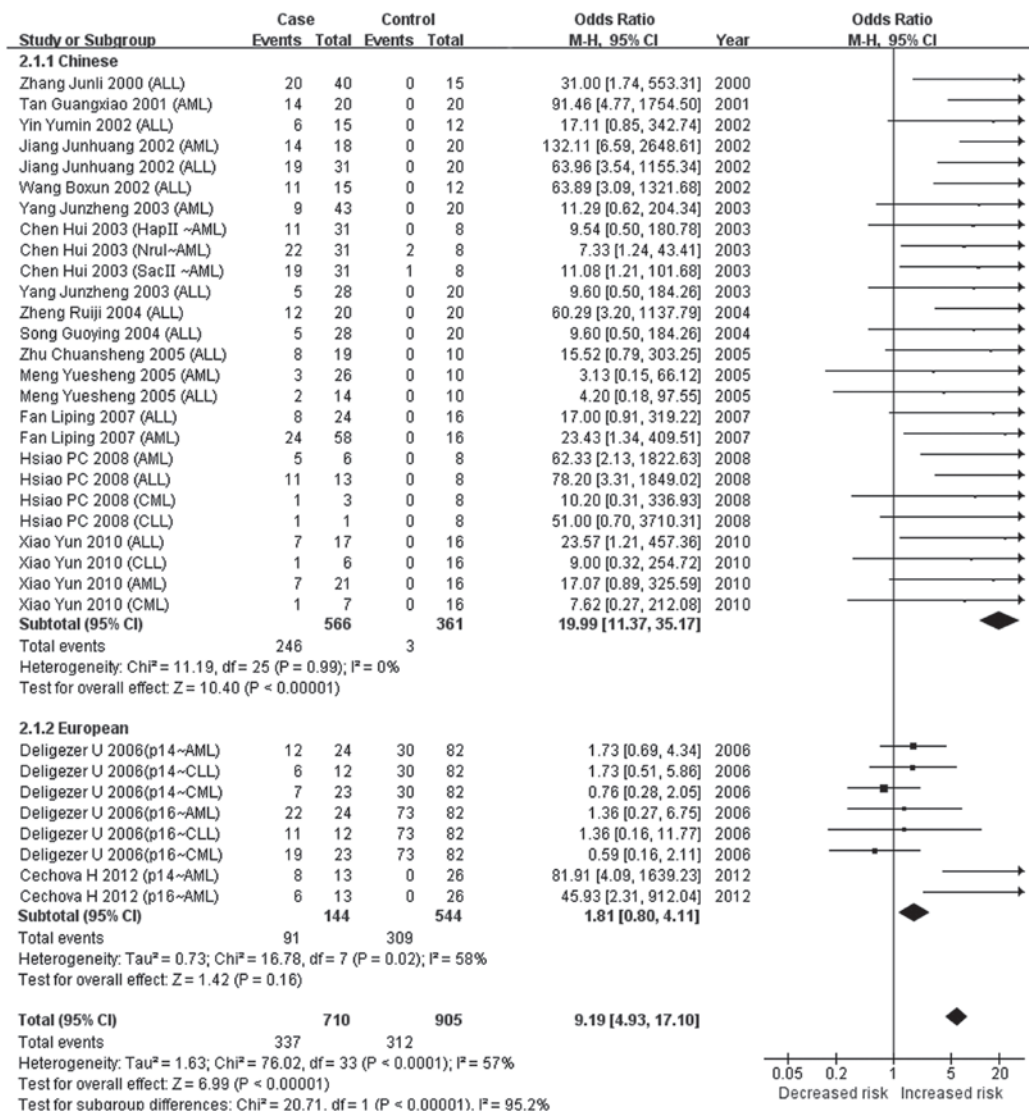


Figure 7. Meta-analyses of aberrantly methylated *CDKN2A* in Asian and European populations. ALL, acute lymphocytic leukemia; CLL, chronic lymphocytic leukemia; AML, acute myelogenous leukemia; CML, chronic myelogenous leukemia; M-H, Mantel-Haenszel model; CI, confidence interval; df, degree of freedom.

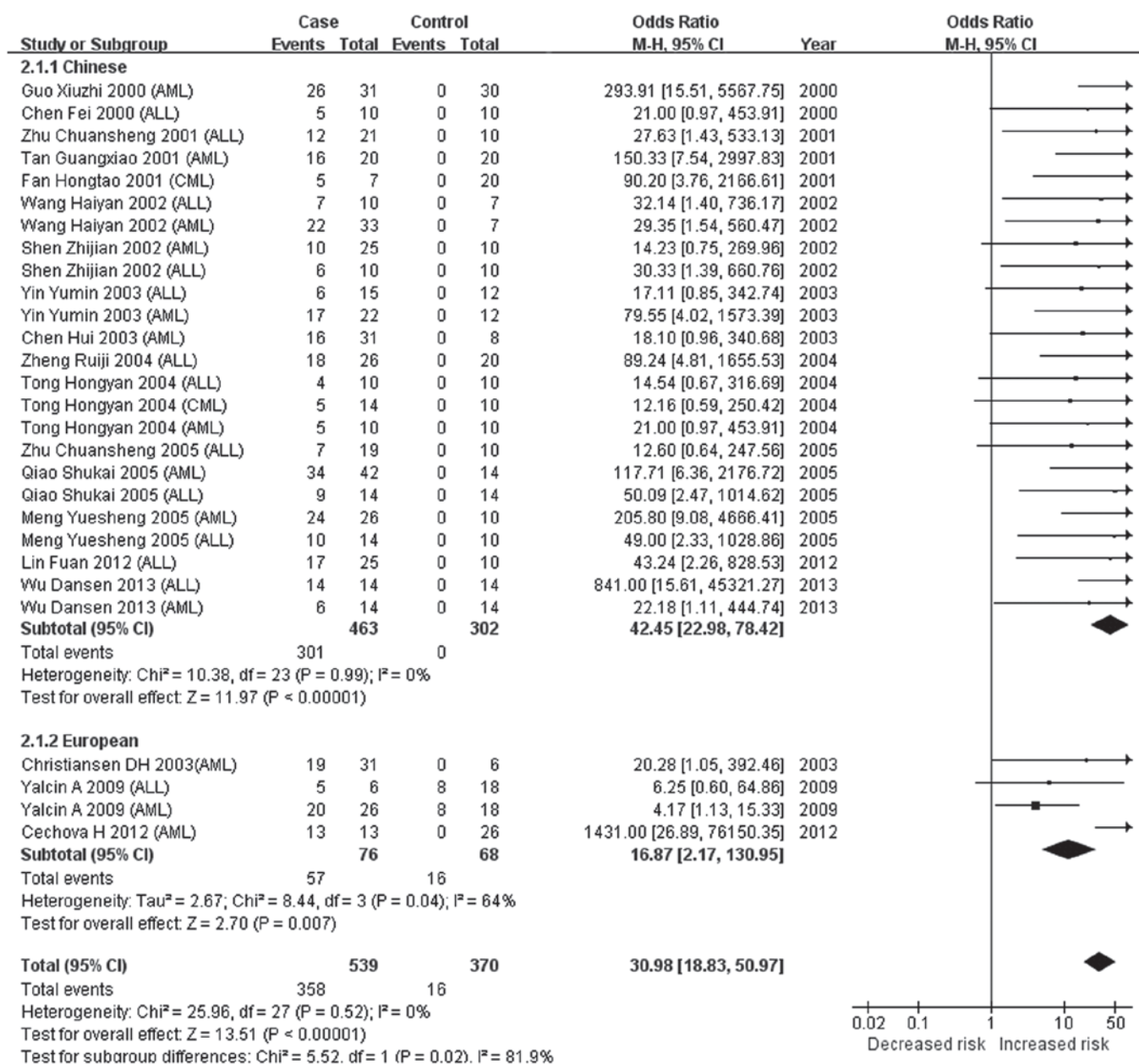


Figure 8. Meta-analyses of aberrantly methylated cyclin-dependent kinase 2B gene in Chinese and European populations. ALL, acute lymphocytic leukemia; AML, acute myelogenous leukemia; CML, chronic myelogenous leukemia; M-H, Mantel-Haenszel model; CI, confidence interval; df, degree of freedom.

Of note, the present meta-analysis had certain limitations. First, the numbers of the studies regarding each gene and leukemia subtype were uneven. For certain leukemia subtypes, only a few studies on certain genes were available. The lack of association of the methylation status of certain genes with several leukemia subtypes may have been due to a lack of statistical power of the respective studies, so that the negative results must be interpreted with caution. Furthermore, a language bias was present, as only studies written in Chinese and English were included.

In conclusion, the present meta-analysis revealed that aberrant DNA methylation of the promoters of 47 genes was associated with leukemia. Further subgroup meta-analysis revealed 5 hypermethylated genes (*CDKN2A*, *CDKN2B*, *ID4*, *GliPR1* and *p73*) in various leukemia subtypes. In addition,

a difference in the association of *CDKN2A* and *CDKN2B* hypermethylation with leukemia was identified between Chinese and European populations. The results of the present study may enhance the current understanding of the association of DNA methylation with leukemia in the Chinese population.

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References

- Davis AS, Viera AJ and Mead MD: Leukemia: An overview for primary care. *Am Fam Physician* 89: 731-738, 2014.
- Zhong S, Chen Z, Yu X, Chen W, Lv M, Ma T and Zhao J: Tea consumption and leukemia risk: A meta-analysis. *Tumour Biol* 35: 5205-5212, 2014.
- Greim H, Kaden DA, Larson RA, *et al*: The bone marrow niche, stem cells, and leukemia: Impact of drugs, chemicals, and the environment. *Ann NY Acad Sci* 1310: 7-31, 2014.
- Laurier D, Grosche B, Auvinen A, *et al*: Childhood leukaemia risks: From unexplained findings near nuclear installations to recommendations for future research. *J Radiol Prot* 34: R53-R68, 2014.
- Malinge S, Chlon T, Doré LC, *et al*: Development of acute megakaryoblastic leukemia in Down syndrome is associated with sequential epigenetic changes. *Blood* 122: e33-e43, 2013.
- Wakeford R: The risk of childhood leukaemia following exposure to ionising radiation - a review. *J Radiol Prot* 33: 1-25, 2013.
- Woo JS, Alberti MO and Tirado CA: Childhood B-acute lymphoblastic leukemia: A genetic update. *Exp Hematol Oncol* 3: 16, 2014.
- Bhatia S: Disparities in cancer outcomes: Lessons learned from children with cancer. *Pediatr Blood Cancer* 56: 994-1002, 2011.
- Carranza C, Granados L, Morales O, *et al*: Frequency of the ETV6-RUNX1, BCR-ABL1, TCF3-PBX1, and MLL-AFF1 fusion genes in Guatemalan pediatric acute lymphoblastic leukemia patients and their ethnic associations. *Cancer Genet* 206: 227-232, 2013.
- Goggins WB and Lo FF: Racial and ethnic disparities in survival of US children with acute lymphoblastic leukemia: Evidence from the SEER database 1988-2008. *Cancer Causes Control* 23: 737-743, 2012.
- Kennedy AE, Kamdar KY, Lupo PJ, *et al*: Genetic markers in a multi ethnic sample for childhood acute lymphoblastic leukemia risk. *Leuk Lymphoma* 56: 169-174, 2014.
- Savage SA: Genomic clues to ethnic differences in ALL. *Blood* 123: 2440-2442, 2014.
- Lim JY, Bhatia S, Robison LL and Yang JJ: Genomics of racial and ethnic disparities in childhood acute lymphoblastic leukemia. *Cancer* 120: 955-962, 2014.
- Adkins RM, Krushkal J, Tylavsky FA and Thomas F: Racial differences in gene-specific DNA methylation levels are present at birth. *Birth Defects Res A Clin Mol Teratol* 91: 728-736, 2011.
- Xia YY, Ding YB, Liu XQ, *et al*: Racial/ethnic disparities in human DNA methylation. *Biochim Biophys Acta* 1846: 258-262, 2014.
- Baylin SB and Jones PA: A decade of exploring the cancer epigenome - biological and translational implications. *Nat Rev Cancer* 11: 726-734, 2011.
- Plass C, Pfister SM, Lindroth AM, *et al*: Mutations in regulators of the epigenome and their connections to global chromatin patterns in cancer. *Nat Rev Genet* 14: 765-780, 2013.
- Jiang D, Hong Q, Shen Y, *et al*: The diagnostic value of DNA methylation in leukemia: A systematic review and meta-analysis. *PLoS One* 9: e96822, 2014.
- Higgins JP, Thompson SG, Deeks JJ and Altman DG: Measuring inconsistency in meta-analyses. *BMJ* 327: 557-560, 2003.
- Bax L, Ikeda N, Fukui N, Yaju Y, Tsuruta H and Moons KG: More than numbers: The power of graphs in meta-analysis. *Am J Epidemiol* 169: 249-255, 2009.
- Pluta A, Nyman U, Joseph B, Robak T, Zhivotovsky B and Smolewski P: The role of p73 in hematological malignancies. *Leukemia* 20: 757-766, 2006.
- Cahill N and Rosenquist R: Uncovering the DNA methylome in chronic lymphocytic leukemia. *Epigenetics* 8: 138-148, 2013.
- Esteller M: CpG island hypermethylation and tumor suppressor genes: A booming present, a brighter future. *Oncogene* 21: 5427-5440, 2002.
- Zhao Y, Guo J, Zhang X, *et al*: Downregulation of p21 in myelodysplastic syndrome is associated with p73 promoter hypermethylation and indicates poor prognosis. *Am J Clin Pathol* 140: 819-827, 2013.
- Volodko N, Gordon M, Salla M, Ghazaleh HA and Baksh S: RASSF tumor suppressor gene family: Biological functions and regulation. *FEBS Lett* 588: 2671-2684, 2014.
- Benezra R, Davis RL, Lockshon D, Turner DL and Weintraub H: The protein Id: A negative regulator of helix-loop-helix DNA binding proteins. *Cell* 61: 49-59, 1990.
- Bodoor K, Haddad Y, Alkhateeb A, *et al*: DNA hypermethylation of cell cycle (p15 and p16) and apoptotic (p14, p53, DAPK and TMS1) genes in peripheral blood of leukemia patients. *Asian Pac J Cancer Prev* 15: 75-84, 2014.
- Chmelařová M, Křepinská E, Spaček J, Laco J, Nekvindová J and Palička V: Methylation analysis of tumour suppressor genes in ovarian cancer using MS-MLPA. *Folia Biol (Praha)* 58: 246-250, 2012.
- Cechova H, Lassuthova P, Novakova L, *et al*: Monitoring of methylation changes in 9p21 region in patients with myelodysplastic syndromes and acute myeloid leukemia. *Neoplasma* 59: 168-174, 2012.
- Deligezer U, Erten N, Akisik EE and Dalay N: Methylation of the INK4A/ARF locus in blood mononuclear cells. *Ann Hematol* 85: 102-107, 2006.
- Hsiao PC, Liu MC, Chen LM, *et al*: Promoter methylation of p16 and EDNRB gene in leukemia patients in Taiwan. *Chin J Physiol* 51: 27-31, 2008.
- Zheng R, Shen S, Shen J and Ma X: Studies on p16, p15, p18 and p19 methylation of INK4 series cancer inhibitive gene in leukemia. *J Fujian Med Univ* 38: 257-260, 2004.
- Xiao Y, Liu Y, Jiang L and Ran L: Study on the deletion point mutation and promoter methylation analysis of p16 in leukemia. *Guizhou Med J* 34: 784-787, 2010.
- Yang J, Li X, Sun L, Song G, Chen H and Liu Y: Expression of p16 gene in adult acute leukemia. *J Zhengzhou Univ (Med Sci Ed.)* 38: 33-36, 2003.
- Song G, Yang J, Sun H, Liu C, Chen H and Li X: Deletion and methylation of p16 gene in an adult acute leukemia. *Henan J Oncol* 17: 229-231, 2004.
- Tan G and Guo X: Methods of CpG islands methylation detection. *J Jinan Univ* 22: 24-26, 2001.
- Zhu C, Xu W, Liu D, Wang Y and Bi K: Study of the methylation of p15 and p16 gene in acute leukemia. *Shandong Med J* 45: 4-5, 2005.
- Zhang J, Wu J and Tan Y: The relationship between P16 gene inactivation and expression of mRNA in ALL. *J Wenzhou Med Coll* 30: 5-7, 2000.
- Fan L, Shen J, Ye B, *et al*: Detection of p16 gene methylation status in adult patients with acute leukemia by using n-MSP. *J Exp Hematol* 15: 258-261, 2007.
- Jiang J, Shen J, Yan J, *et al*: The value of p15 gene methylation in minimal residual disease of childhood acute leukemia by MSP. *China Child Blood* 7: 1-3, 2002.
- Meng Y, Yu H, Guo C, *et al*: Quantitative analysis of p15 and p16 gene methylation and mRNA expression levels in acute leukemia patients. *Chin J Hematol* 26: 434-435, 2005.
- Wang B: The clinical significance of p16 gene methylation in leukemia and other hematologic malignancies. Master's degree dissertation, China Medical University, 2002.
- Yin Y: The clinical significance of p15 gene methylation in leukemia and other hematologic malignancies. Master's degree dissertation, China Medical University, 2002.
- Chen H, Sun H, Liu C, Liu Y, Song G and Li X: Methylation of p16/p15 exon 2 in adult acute myeloid leukemia. *J Zhengzhou Univ (Med Sci Ed.)* 38: 722-724, 2003.
- Lin F, Ye B, Shen J, *et al*: Detection of p15 methylation and deletion status in acute lymphoblastic leukemia using hn-MSP. *J Leuk Lymphoma* 21: 208-212, 2012.
- Zhu C, Bi K and Liu X: Homozygous deletion and methylation of P15 gene in acute leukemia. *Shandong Med J Shandong Yiyao* 41: 10-11, 2001.
- Shen Z, Zhang J, Yu K and Jiang S: Evaluation of methylation-specific polymerase chain reaction and its potential clinical value in detecting minimal residual disease of adult acute leukemia with P15. *J Wenzhou Med Coll* 32: 230-232, 2002.
- Fan H, Guo X, Tan G, Wu Q, Zhou T and Guo Q: Study on methylation of p15 INK4B in myelodysplastic syndrome. *Chin J Pathophysiol* 17: 349-352, 2001.
- Tong H and Lin M: Methylation of p15INK4B gene and its mechanism in patients with myelodysplastic syndrome and leukemia. *Chin J Pathophysiol* 20: 2257-2260, 2004.
- Guo X, Fan H, Wu Q, *et al*: Study on methylation of p15INK4B gene in acute myeloid leukemia and chronic myeloid leukemia. *J Exp Hematol* 8: 257-260, 2000.
- Qiao S, Xu S, Guo X and Wang Y: Clinical significance of the expression of DNA methyltransferase genes (DNMT) in acute leukemia patients. *J Exp Hematol* 13: 260-265, 2005.
- Chen F and Zeng X: Detection of p15 gene hypermethylation in acute leukemia by MSP. *Chin J Hematol* 21: 250, 2000.

53. Yin Y, Li Y and Liu D: The clinical significance of p15 gene methylation in leukemia and other hematologic malignancies. *Chin J Hematol* 24: 383-384, 2003.
54. Wu D: p15INK4b gene DNA methylation and histone modification mechanism of leukemia cell mediated by p15-piRNAs. PhD dissertation, Fujian Medical University, 2013.
55. Wang H: The study of p15 gene methylation in malignancies. Master's degree dissertation, Qingdao University, 2002.
56. Zhao Y, Wang QS, Li HH, *et al*: [Significance of id4 promoter methylation in monitoring AML patients with completely remission]. *J Exp Hematol* 16: 476-478, 2008.
57. Wang X, Kang H, Cen J, Li Y, Wang L and Yu L: Methylation Status of id4 gene promoter in patients with chronic myeloid leukemia. *J Exp Hematol* 18: 1402-1404, 2010.
58. Liu F, Xu R, Cui X, Zhang X and Wang Y: ID4 promoter methylation in acute myeloid leukemia. *J Exp Hematol* 19: 582-584, 2011.
59. Jie X: WT1, ID4, GLIPR1 aberrant gene promoter methylation and mRNA expression in leukemia patients. Master's degree dissertation, Guangdong Medical College, 2012.
60. Zhao Y: The significance of Id4 gene methylation in hematological malignancies. PhD dissertation, PLA Postgraduate Medical School, 2005.
61. Xiao YH, Li XH, Tan T, *et al*: Identification of GLIPR1 tumor suppressor as methylation-silenced gene in acute myeloid leukemia by microarray analysis. *J Cancer Res Clin Oncol* 137: 1831-1840, 2011.
62. Liang T, Tan T, Xiao Y, *et al*: Methylation and expression of glioma pathogenesis-related protein 1 gene in acute myeloid leukemia. *J Cent South Univ (Med Sci Ed.)* 34: 388-394, 2009.
63. Zhang Y, Gao Y, Wang X, *et al*: The relationship between p73 gene methylation and childhood acute leukemia. In: *Proceedings of the Ninth National Conference on Environmental and Occupational Medicine Graduate*, Shanghai, China: 254-256, 2010.
64. Zhang H, Mo W and Yang Z: The expression of WWOX and p73 in acute myeloid leukemia. *Guangdong Med J* 33: 1567-1570, 2012.
65. Wu C, Jin J, Ji Y, *et al*: Abnormal expression and significance of p73 gene in adult acute lymphoblastic leukemia. *Jiangsu Med J* 34: 339-341, 2008.
66. Liu J, Jiang Y, Xu W and Liang X: Alterations of p53 and p73 genes in lympho-plasmacytic diseases. *Pract Clin Med* 6: 9-13, 2005.
67. Xu W: Study on methylation of the p73 gene in acute lymphoblastic leukemia and its clinical significances. *Chin J Pract Int Med* 25: 905-906, 2005.
68. Yu G: Abnormal expression of p73 gene in adult acute lymphoblastic leukemia. *Med Innov Chin* 2: 121-122, 2014.
69. Xie X, Chen Z, Zhang X and Huang M: Detecting methylation of the calcitonin gene in monitoring treatment and disease evolution for myelogenous leukemia. *Chin J Cancer* 22: 616-619, 2003.
70. Tang Y, Deng C, Du Q and Li G: Study on hypermethylation of the calcitonin gene in malignant hematological disorders. *J West Chin Univ Med Sci* 32: 86-88, 2001.
71. Wang S, Hong W and Peng A: The study of CT gene methylation in chronic myeloid leukemia. *Chin J Hematol* 19: 32, 1998.
72. Qian J, Yao D, Lin J, *et al*: Alteration of methylation status of death-associated protein kinase (dapk) gene promoter in patients with acute myeloid leukemia. *J Exp Hematol* 18: 1390-1394, 2010.
73. Niu Y, Wang P, Wang Y, Wang Y, Cai D and Li Y: Expression of death-associated protein kinase gene and methylation status of promoter region in acute leukemia. *J Exp Hematol* 22: 30-34, 2014.
74. Zhao W: DAPK gene promoter methylation, expression and biological function in acute leukemia. PhD dissertation, Southern Medical University, 2009.
75. Qian J, Wang YL, Lin J, Yao DM, Xu WR and Wu CY: Aberrant methylation of the death-associated protein kinase 1 (DAPK1) CpG island in chronic myeloid leukemia. *Eur J Haematol* 82: 119-123, 2009.
76. Lin W, Ye Z and Tan J: Gene methylation abnormalities in chronic lymphocytic leukemia patients by PCR. *J Qiqihar Med Coll* 31: 2376-2377, 2010.
77. Wu B, Chen E and Zhu B: Detecting the aberrant methylation of progesterone receptor B gene in human chronic lymphocytic leukemia by MSP. *Chin J Hemorheol* 18: 140-141, 2008.
78. Zhang X: The experimental study of PR promoter CpG island methylation in leukemia. Master's degree dissertation, Third Military Medical University, 2003.
79. Shi X, Chang N, Fan Y, *et al*: Detection and clinical significance of SFRP2 promoter methylation status in patients with acute myeloid leukemia. *Chin J Clin, Electron Ed.* 5: 1962-1966, 2011.
80. Wang H, Zhang G, Yao K, *et al*: The gene methylation and protein expression of SFRP5 in leukemic cells. *J China Med Univ* 41: 2012.
81. Xu C, Shen J, Shen S, Fu H, Zhu Y and Chen L: The significance of methylation status of secreted frizzled-related protein gene promoter in acute leukemia. *Chin J Intern Med* 49: 769-771, 2010.
82. Li M: A preliminary study of inactivation of IGSF4 gene methylation in leukemia cell. PhD dissertation, PLA Postgraduate Medical School, 2004.
83. Chen W, Huang C, Zhan X, *et al*: RASSF1A gene methylation and its clinical significance in leukemia patients. *Lab Med* 27: 554-556, 2012.
84. Song J, Li Y, Yang Z, Yang L and Chen J: Hypermethylation in SFRP2 promoter in human leukemia K562 cells and bone marrow specimen with chronic myeloid leukemia. *Acad J Third Med Coll PLA* 33: 2583-2586, 2011.
85. Dou L: The study of LRP15 gene methylation in blood diseases. Master's degree dissertation, PLA Postgraduate Medical School, 2004.
86. Yao C, Fang J, Gao L, Ding J, Shu G and Yu W: Study on the relationship between the RIZ1 gene expression and methylation status of the gene promoter in acute myeloid leukemia. *J Southeast Univ (Med Sci Ed.)* 29: 119-122, 2010.
87. Cai F: RIZ1 gene expression and promoter methylation status in acute childhood leukemia. PhD dissertation, Huazhong University Science and Technology, 2012.
88. Shi X, Fan Y, Zhou C, *et al*: Study of abnormal methylation in the promoter region of SFRP4 gene in acute myeloid leukemia. *Chin J Clin, Electron Ed.* 5: 990-995, 2011.
89. Dou LP, Liu JH, Wang C, *et al*: [Study on the involvement of ZO-1 gene in leukemogenesis]. *Chin J Hematol* 30: 473-476, 2009.
90. Wang C, Wang G, Tan Y, Li W, Liu C and Yu L: The methylation pattern and clinical significance of Zonula occludens-1 gene promoter in acute leukemia. *Chin J Intern Med* 47: 111-113, 2008.
91. Chim CS, Wong SY, Pang A, *et al*: Aberrant promoter methylation of the retinoic acid receptor alpha gene in acute promyelocytic leukemia. *Leukemia* 19: 2241-2246, 2005.
92. Wang G, Liu L, Chen J, *et al*: Promoter methylation profile of androgen receptor gene in leukemic cells. *J Third Mil Med Univ* 29: 932-934, 2007.
93. Wang X, Li J, Fu B, Guo L, Zhang J and Huang S: Methylation Status of JunB and CDH13 gene promoter in CD34 CD38⁺ chronic myelogenous leukemia cells. *J Exp Hematol* 17: 1405-1408, 2009.
94. Liu J: CDH13 gene promoter methylation and expression in acute myeloid leukemia. Master's degree dissertation, Guangzhou Medical University, 2013.
95. Wang Y: The study of DDIT3 gene promoter methylation in hematological tumors. Master's degree dissertation, Jiangsu University, 2009.
96. Zhu X, Zhu C, Wan Y, *et al*: Analysis of methylation of the Dickkopf1 (DKK-1) gene in acute leukemia. *J Shandong Univ (Health Sci)* 50: 84-87, 2012.
97. Yuan X: The study of EDNRB gene promoter methylation in acute leukemia. Master's degree dissertation, Zhengzhou University, 2010.
98. Yu Y: The study of FANCF methylation in patients with acute myeloid leukemia and myelodysplastic syndrome. Master's degree dissertation, China Medical University, 2008.
99. Deng N: The study of FANCF/FANCD2 expression in patients with acute myeloid leukemia and myelodysplastic syndrome. Master's degree dissertation, China Medical University, 2009.
100. Qian Z: The significance of GRAF gene promoter methylation in hematopoietic tumors. Master's degree dissertation, Jiangsu University, 2010.
101. Chen Q: HAGE gene promoter hypomethylation in myeloid tumors. Master's degree dissertation, Jiangsu University, 2012.
102. Li Y, Wang Y, Zhou J, Zhou S, Fang G and Liu X: Analysis and clinical significance of hPer3 promoter methylation status for CML monitoring. *Chin J Pathophysiol* 27: 2111-2115, 2011.
103. Wang Y, Zhou J, Zhou S, *et al*: Promoter methylation status of hPer3 gene in AML patients and the *in vitro* effect of decitabine on the status. *Chin J Hematol* 32: 317-321, 2011.
104. Wang N: MiR-34b expression, methylation regulation and clinical significance in acute childhood leukemia. Master's degree dissertation, Suzhou University, 2013.
105. Chai H: RAGE-1 gene promoter hypomethylation in myeloid tumors. Master's degree dissertation, Shanxi Medical University, 2013.

106. Lin D, Fan R and Liu X: Significance of DNA methylation status of runx3 gene promoter region in acute leukemia. *J Exp Hematol* 16: 263-266, 2008.
107. Chim CS, Wong AS and Kwong YL: Epigenetic dysregulation of the Jak/STAT pathway by frequent aberrant methylation of SHP1 but not SOCS1 in acute leukaemias. *Ann Hematol* 83: 527-532, 2004.
108. Wang Y, Zhu CS, Bi KH, Xu WW, Dong L and Hou M: [Study of WIF-1 promoter methylation with expressions of β -catenin in acute leukemia]. *Chin Med J* 91: 2858-2860, 2011.
109. Liu W, Xu C, Guan M and Zhang Y: Aberrant promoter hypermethylation of AKAP12 gene in acute lymphoblastic leukemia in children. *Lab Med* 23: 344-348, 2008.
110. Gao F, Li Y, Liu W, *et al*: Studies on gene expression and the 5' CpG islands methylation status of E-cadherin in acute myeloid leukemia. *Chin J Hematol* 27: 25-27, 2006.
111. Yao DM, Qian J, Lin J, *et al*: Aberrant methylation of CCAAT/enhancer binding protein zeta promoter in acute myeloid leukemia. *Leuk Res* 35: 957-960, 2011.
112. Guan M, Xu C, Liu W and Chen B: Identification of aberrant methylation of DLC-1 by pyrosequencing in acute lymphoblastic leukemia in children. *Chin J Lab Med* 31: 389-393, 2008.
113. Yu L, Wang Q, Gu Q, Shi Z and Jin H: Study on aberrant DNA methylation of dopamine D4 receptor gene in acute myeloid leukemia. *Acad J Chin PLA Postgrad Med Sch* 21: 62-64, 2000.
114. Tao YF, Xu LX, Lu J, *et al*: Metallothionein III (MT3) is a putative tumor suppressor gene that is frequently inactivated in pediatric acute myeloid leukemia by promoter hypermethylation. *J Transl Med* 12: 182, 2014.
115. Zheng Y: Abnormal methylation of the promoter region of the p53 gene in acute leukemia. Master's degree dissertation, Sun Yat-sen University, 2007.
116. Li Y, Qian J, Chen Q, *et al*: Expression pattern and promoter hypermethylation of PDLIM4 gene in patients with chronic myeloid leukemia. *J Jiangsu Univ (Med Ed)* 23: 17-21, 2013.
117. Song JJ, Liu Q, Li Y, *et al*: Epigenetic inactivation of PLCD1 in chronic myeloid leukemia. *Int J Mol Med* 30: 179-184, 2012.
118. Yao D, Qian J, Lin J, Li Y, Chen Q and Chen X: Alteration of methylation status of PRAME gene promoter in patients with chronic myeloid leukemia. *J Pract Med* 29: 3495-3498, 2013.
119. Yan W, Hu W and Yang W: Expression and methylation study of PRDX2 gene in acute myeloid leukemia. *J China Med Univ* 41: 1047-1049, 2012.
120. Yang J, Du W, He Y, Liu J, Zheng J and Huang S: Research on PTEN expression and its promoter methylation in B-ALL patients. *J Intern Intens Med* 13: 141-143, 2007.
121. Du C, Mao P and Wang Y: The clinical significance of the RIL gene methylation levels in acute myeloid leukemia. *China J Mod Med* 23: 46-51, 2013.
122. Jiao X, Chen Q, Lin J, *et al*: SALL4 genes methylation changes and clinical significance in acute myeloid leukemia patients. *J Jiangsu Univ (Med Ed)* 23: 532-535, 2013.
123. Zhuang Y, Cheng Y, Wang L, Dou H, Zhu Q and Hu J: The methylation of SOCS-1 gene in adult acute myeloid leukemia. *Prog Mod Biomed* 18: 3417-3420, 2011.
124. Deng G, Li ZQ, Zhao C, *et al*: WNT5A expression is regulated by the status of its promoter methylation in leukaemia and can inhibit leukemic cell malignant proliferation. *Oncol Rep* 25: 367-376, 2011.
125. Iacobucci I, Ferrari A, Lonetti A, *et al*: CDKN2A/B alterations impair prognosis in adult BCR-ABL1-positive acute lymphoblastic leukemia patients. *Clin Cancer Res* 17: 7413-7423, 2011.
126. Hu HB and Hu Q: ID4 methylation patterns in childhood T line and B line lymphocytic leukemia. *Chin J Contemp Pediatr* 12: 940-942, 2010 (In Chinese).
127. Chen SS, Claus R, Lucas DM, *et al*: Silencing of the inhibitor of DNA binding protein 4 (ID4) contributes to the pathogenesis of mouse and human CLL. *Blood* 117: 862-871, 2011.
128. Liu Y, Kang HY, Wang LL, Lu XC, Zhu HL and Yu L: Establishment of methylation-specific quantitative PCR system for ID4 gene in acute leukemia cells and its specificity and sensitivity. *J Exp Hematol* 22: 269-274, 2014 (In Chinese).
129. Liu F and Xu RR: Study on the correlation between Chinese medical syndrome types and ID4 gene promoter methylation in human acute myeloid leukemia. *Chin J Integr Tradit West Med* 32: 471-473, 2012 (In Chinese).
130. Awasthi A, Woolley AG, Lecomte FJ, *et al*: Variable expression of GLIPR1 correlates with invasive potential in melanoma cells. *Front Oncol* 3: 225, 2013.
131. Capalbo G, Mueller-Kuller T, Koschmieder S, *et al*: Endoplasmic reticulum protein GliPR1 regulates G protein signaling and the cell cycle and is overexpressed in AML. *Oncol Rep* 30: 2254-2262, 2013.
132. He Z, Liu H, Agostini M, *et al*: p73 regulates autophagy and hepatocellular lipid metabolism through a transcriptional activation of the ATG5 gene. *Cell Death Differ* 20: 1415-1424, 2013.
133. Toska E and Roberts SG: Mechanisms of transcriptional regulation by WT1 (Wilms' tumour 1). *Biochem J* 461: 15-32, 2014.
134. Does GM, Devesa SS, Curtis RE, Linet MS and Morton LM: Acute leukemia incidence and patient survival among children and adults in the United States, 2001-2007. *Blood* 119: 34-43, 2012.
135. Shirley MH, Sayeed S, Barnes I, Finlayson A and Ali R: Incidence of hematological malignancies by ethnic group in England, 2001-7. *Br J Haematol* 163: 465-477, 2013.