Gene expression of adipose tissue, endothelial cells and platelets in subjects with metabolic syndrome (Review)

PABLO PÉREZ M. 1 , RODRIGO MOORE-CARRASCO 1,2 , DANIEL GONZÁLEZ R. 3 , EDUARDO FUENTES Q. $^{1,2}\,$ and IVÁN PALOMO G. $^{1,2}\,$

Department of Clinical Biochemistry and Immunohematology, Faculty of Health Sciences, Programa de Investigación en Factores de Riesgo de Enfermedades Cardiovasculares (PIFRECV), Universidad de Talca;
Centro de Estudios en Alimentos Procesados (CEAP), Conicyt-Regional, Gore Maule R09I2001;
Department of Basic Biomedical Sciences, Faculty of Health Sciences, Programa de Investigación en Factores de Riesgo de Enfermedades Cardiovasculares (PIFRECV), Universidad de Talca, Talca, Chile

Received November 4, 2011; Accepted January 30, 2012

DOI: 10.3892/mmr.2012.785

Abstract. Metabolic syndrome is a combination of medical disorders including hypertension, dyslipidemia, hyperglycemia, insulin resistance and increased waist circumference, and is associated with a higher risk of cardiovascular disease. An increase in adipose tissue mass is associated with the augmented secretion of certain adipokines, such as interleukin-6, tumor necrosis factor-α and resistin, which cause endothelial dysfunction (an increase in vasoconstrictor molecules and in the expression of adhesion molecules as well as a decrease of vasodilator molecules, amongst other features) and hemostasis alterations that also favor a prothrombotic state (increased fibrinogen and plasminogen activator inhibitor-1 concentrations and platelet activation/aggregation). This interaction between adipose tissue, endothelial cells and platelets is associated with an increase or decrease in the expression of several transcription factors (peroxisome proliferator-activated receptors, CCAAT-enhancer-binding proteins, carbohydrate responsive element-binding proteins and sterol regulatory element-binding proteins) that play a crucial role in the regulation of distinct metabolic pathways related to the metabolic syndrome. In the present review, we present the primary changes in adipose tissue, endothelial cells and platelets in subjects with metabolic syndrome and their possible target sites at the gene expression level.

Correspondence to: Dr Iván G. Palomo, Department of Clinical Biochemistry and Immunohematology, Faculty of Health Sciences, Universidad de Talca, Mailbox 747, Talca, Chile E-mail: ipalomo@utalca.cl

Key words: metabolic syndrome, endothelial dysfunction, adipose tissue, platelets, transcription factors

Contents

- 1. Introduction
- 2. Gene expression and the metabolic syndrome
- 3. Gene expression of adipose tissue, endothelial cells and platelets during metabolic syndrome
- 4. Conclusion

1. Introduction

Metabolic syndrome (MS) is a combination of pathophysiological changes that include arterial hypertension, insulin resistance, dyslipidemia and abdominal obesity (1). Previous studies have revealed that subjects with MS present a high risk of developing type-2 diabetes mellitus (T2DM) and cardiovascular disease (CVD) (2). MS presents a prothrombotic state, a result of endothelial dysfunction and hypercoagulability produced by an imbalance between coagulation factors and the proteins that regulate fibrinolysis (3). The principal characteristic of endothelial dysfunction is a decrease in the bioavailability of nitric oxide (NO) at the vascular level, accompanied by other changes in the endothelial phenotype along with an increase in vasoconstriction and inflammation (4). As regards hypercoagulability, MS has also been found to present elevated levels of fibringen, vitamin K-dependent coagulation factors (FII, FVII, FIX and FX), factor XIII and von Willebrand factor (5). This increase in fibrinogen levels is proportional to the overexpression of GPIIb/ IIIa and the size of the platelet aggregates (6).

This series of changes presented in MS (in which adipose tissue, endothelial cells and platelets are involved) favors the secretion of several molecular mediators (adipokines, cytokines and chemokines, amongst others) capable of activating or suppressing a number of transcription factors [peroxisome proliferator-activated receptors (PPARs), sterol regulatory element-binding proteins (SREBPs), carbohydrate responsive element-binding proteins (C/EBPs)]; these transcription factors (depending on DNA-binding affinity, the recruitment of

co-activators and proteasomal degradation) regulate different MS-related metabolic pathways (7). To a certain extent, the action of agonists on these transcription factors varies the metabolic changes associated with MS (8).

This review presents the principal molecular aspects of MS and how these can become modifiable target sites through the action of the mediators produced by adipose tissue, endothelial cells and platelets.

2. Gene expression and the metabolic syndrome

The pathophysiological changes presented in MS are diverse and include endothelial dysfunction and hypercoagulability (9). Changes specifically associated with endothelial dysfunction include decreased synthesis of NO and prostacyclin, increased vascular cell adhesion molecule-1 (VCAM-1), soluble CD40 ligand (sCD40L), endoperoxide, reactive oxygen species (ROS) and endothelin-1 levels and asymmetric dimethylarginine (10). Changes in hemostasis and the fibrinolytic system, elevated plasma levels of plasminogen activator inhibitor-1 (PAI-1) (significantly associated with diastolic arterial pressure, triglycerides and waist-line circumference) and of fibrinogen (negatively associated with HDL-C) contribute to the expansion of CVD (11).

Dietary habits and physical activity are important factors of the first-line intervention for MS (12). However, there are key factors in MS regulation that depend on those transcription factors that, by responding and adapting to signals from the environment, are able to change the levels of relevant gene expression and thereby regulate energy expenditure, substrate metabolism and food consumption in MS (13,14). The genes expressed during the development of MS include various groups of genes: genes specific to adipose tissue, genes involved in the proliferation and differentiation of adipocytes and genes that code for insulin-receptor substrate proteins, amongst others (15).

To date, four families of nuclear receptors involved in MS have been described (16). These families correspond to those transcription factors that, when activated by ligands, bind to target gene regulatory regions and thus modulate the expression of MS (17). PPARs, C/EBPs, ChREBPs and SREBPs have been identified as regulators of distinct metabolic pathways during the transcription of genes expressed in MS (18). The following descriptions provide more background on each.

PPARs are transcriptions factors of a superfamily of nuclear receptors. Three isoforms exist: PPAR- α , PPAR- β (before PPAR- δ) and PPAR- γ (19). PPARs form heterodimers with the retinoid X receptor and, after ligand binding, modulate the downstream gene expression of target genes, which are translated into specific complexes that regulate proliferation, differentiation and cell survival (20). PPARs play a crucial role in the regulation of intermediary metabolism and inflammation, where their activity is not affected by phosphorylation resulting from p38-MAPK, PKA, PKC, AMPK and GSK3 (21,22).

The main components are PPAR- α and PPAR- γ , which also have a direct relation to hypertriglyceridemia and insulin resistance, respectively (23). PPAR- α participates in the initiation and development of atherosclerosis, promoting an increase in angiotensin II levels, oxidative stress and arterial pressure (24). PPAR- γ plays a fundamental role in adipogenesis (25), as

a key regulator in the differentiation and function of adipocytes and the absorption of stored fatty acids (26). Additionally, it has been recently suggested that PPAR- γ is a key regulator of inflammatory and immune response (27). Mutations and single nucleotide polymorphism in PPAR- γ are associated with metabolic disease and inflammation (28). PPAR- β participates in the regulation of several metabolic pathways: metabolism of fatty acids, cellular respiration and the programming of muscle fiber types (29). It is particularly active in the regulation of the various genes involved in the inflammatory process (30).

C/EMPs are a family of transcription factors that feature a basic region leucine zipper domain (31). They participate in the differentiation of a wide range of cell types, with a key role in the regulation of cell proliferation via interaction with cell cycle proteins (32). Furthermore, they are important regulators of hepatic metabolism, affecting the expression of genes implicated in gluconeogenesis, glycogen storage and lipid metabolism (33). At least six members of this family have been isolated and characterized (34). Of these, C/EBP- α , β and δ are tissue-specific and highly expressed in adipose tissue. Additionally, C/EBP- α and β are expressed in the liver. Based on this, C/EBP- α , β and δ may represent potential targets for obesity and the metabolic changes that contribute to the development of MS (35).

ChREBP is a transcription factor that plays a critical role in the glucose-mediated induction of genes involved in glycolysis, lipogenesis and gluconeogenesis (36). Blood glucose levels affect the activity of ChREBP in hepatocytes largely via dephosphorylation (37), which increases its translocation to the nucleus, in order to stimulate lipogenic genes (38). During fasting, PKA and AMPK phosphorylate ChREBP, which inhibits its activity. During feeding, protein phosphatase 2 activates ChREBP during its dephosphorylation (39).

SREBPs constitute the major family of transcription factors regulating the expression of genes that encode the necessary enzymes for the biosynthesis of fatty acids and cholesterol (40,41). The activation of Akt induces the synthesis and accumulation of SREBP-1, necessary for the expression of fatty acid synthase, a key regulatory enzyme in the biosynthesis of lipids (42). The activation/degradation of SREBPs and the stability of this rate-limiting enzyme during the synthesis of lipids are both regulated by the ubiquitin-proteasome system in a sterol-dependent process (43). Insulin, by means of post-translational modification, activates the transcription and proteolytic maturation of the SREBP-1c present in the membranes of endoplasmic reticulum (44).

3. Gene expression of adipose tissue, endothelial cells and platelets during metabolic syndrome

An increase in the amount of free fatty acids participates in platelet activation and leads to an increase in the production of low-density lipoprotein (LDL) and very-low-density lipoprotein (VLDL). This, in turn, is associated with an increase in ROS yielding oxidized LDL (ox-LDL) and its subsequent accumulation in damaged subendothelium (45). Lipoproteins affect the platelet function through binding to specific receptors (CD36, SR-B1 and LOX-1) (46,47).

It is precisely the erosion or rupture of these lesions that provokes the platelet/endothelium interaction, whose communication at various levels is a key in the response to vascular

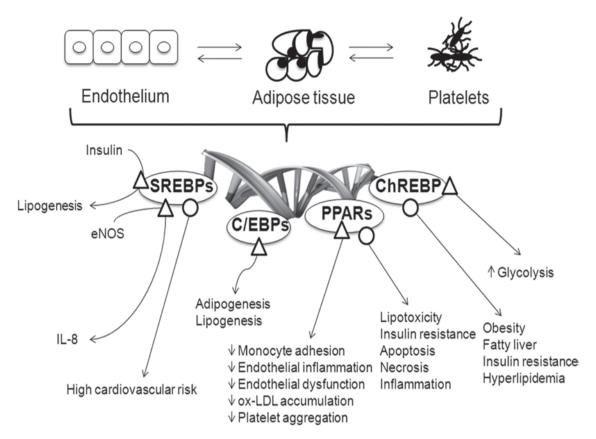


Figure 1. Molecular interactions and gene expression of adipose tissue, endothelial cells and platelets during metabolic syndrome. The triangle symbol indicates transcription factor activation and the circular symbol indicates suppression of the transcription factor. eNOS, endothelial nitric oxide synthase; IL-8, interleukin-8. SREBPs, sterol regulatory element-binding proteins; C/EBPs, CCAAT-enhancer-binding proteins; PPARs, peroxisome proliferator-activated receptors; ChREBP, carbohydrate responsive element-binding protein; ox-LDL, oxidized low-density lipoprotein.

damage (48). These interactions between adipose tissue, endothelial cells and platelets secrete a series of atherogenic and pro-inflammatory factors that have implications in the genetic expression of MS (Fig. 1) (49). The principle genetic expression present in adipose tissue, endothelial cells and platelets during MS is described below.

Adipose tissue and its gene expression in MS. In MS, adipose tissue secretes elevated quantities of adipokines, in particular tumor necrosis factor- α (TNF- α), interleukin (IL)-6 and resistin (50), which contributes to insulin resistance during diabetes and obesity by inhibiting glucose uptake, glycogen synthesis and glucose oxidation (51).

The global burden of metabolic disease necessitates the development of new therapeutic strategies, in which the available alternative is to alter the main transcription pathways that regulate the absorption of glucose, lipid manipulation or adipokine secretion (52). Changes in the gene expression of adipose tissue suggest that carbohydrate modification affects the risk of CVD and T2DM (53). The gene expression in the adipose tissue of individuals with MS seems to be affected by changes in tissue morphology or insulin sensitivity, where a diet high in saturated fatty acids produces a pro-inflammatory state via the repression of PPARs (54). The exceptions to ligands for PPAR are short-chain fatty acids and long-chain monounsaturated fatty acids (55).

The maturation of adipocytes is regulated by a series of transcription factors, mainly PPAR- γ and C/EBP, which in

conjunction regulate the expression of hundreds of proteins that participate in the metabolism and storage of lipids and, as such, the secretion of adipocytes (56). Adiponectin is a protein specific to adipose tissue with anti-inflammatory and anti-atherogenic properties (57,58). The double action of PPAR- α and PPAR- γ increases the action of adiponectin and the expression of its receptors, which results in an improvement in obesity and a reduction of the inflammatory process (59). On the contrary, the loss of function of PPAR- γ due to dominant mutations brings about a resistance to insulin and the early onset of severe hypertension (60).

A hyperglycemic state is the result of a minor expression of glucose transporter-2 (GLUT-2), which is related to a low expression of SREBP-1c (61). Glucose also stimulates ChREBP by dephosphorylation of specific amino acids (62), driving the activation of target genes that code for acetyl-CoA carboxylase, fatty acid synthase and liver-specific pyruvate kinase (63). It has also been shown that ChREBP increases the expression of enzymes that participate in the fatty acid synthesis, such as stearoyl-CoA desaturase, which facilitates the conversion of glucose into fatty acids (64).

Endothelial cells and gene expression in MS. Endothelial dysfunction in MS appears to be the consequence of an increase in oxidative stress with a decrease in the bioavailability of NO (65). In addition to catalyzing NO production (66), endothelial NO synthase (eNOS) participates in the activation and increase in the transcription of

SREBP, which finally induces the expression of IL-8 (67). In combination, vascular endothelial growth factor (VEGF) mediates the activation of SREBP through its receptor in endothelial cells (VEGFR2), which is translated into the overexpression of IL-8 and the LDL receptor, accelerating the inflammatory process via the recruitment and adhesion of leukocytes and the accumulation of LDL in the subendothelial space, respectively (68,69).

On the contrary, PPAR-δ inhibits the production of ROS induced by TNF- α , while also stimulating the expression of important antioxidant enzymes (superoxide dismutase, catalase and thioredoxins), thereby acquiring a potential role in endothelial survival and proliferation (70). Within this same family of transcription factors, PPAR-α inhibits the overexpression of VCAM-1 and the hyperactivity of endothelial cells prior to stimulation with TNF-α (71). In addition, PPAR-γ (by inhibiting the activity of NF-κB and AP-1) suppresses the expression of chemokine genes (IFN-γ) and inhibits the expression of pro-inflammatory adhesion molecules (VCAM-1, ICAM-1 and E-selectin) with a decrease in the adhesion of monocytes to active endothelial cells (72). Consistent with this, heterozygous mice with a minor expression of PPAR-y present with endothelial dysfunction and systolic hypertension (73). Additionally, transgenic mice with a recessive expression of PPAR-γ develop endothelial dysfunction in response to a high-fat diet (74).

Platelets and gene expression in MS. Platelets are small fragments of circulating cells that perform a critical role in the progress of CVD, T2DM, inflammation and metastasis of cancerous cells (75). Numerous biochemical abnormalities have been found that correlate with the platelet hyperactivity in MS (76), apparently where hyperglycemia seems to promote platelet hyperactivity and increases the overexpression of glycoproteins (77). This, in turn, brings about the production of thromboxane A2 with increased platelet sensitivity (76).

Despite their absence of nucleus, selective agonists for their nuclear receptors (principally PPARs) regulate platelet activation and aggregation (78). PPAR-β is present in platelets, by action of the synergistic effect between prostacyclin and NO, decreases the inflammatory process and delays the formation of atheromatous plaque, thus attenuating the progression of atherosclerosis (79). Another transcription factor present in platelets is PPAR-y, which, in response to two natural ligands [lysophosphatidic acid and 15-deoxy-δ 12 14-prostaglandin J2 (metabolite of prostaglandin-d)], attenuates the release of pro-inflammatory mediators, such as sCD40L and thromboxane A2 (80). Furthermore, PPAR-γ agonists inhibit the platelet aggregation induced by collagen by modulating the signal pathways of GPVI (81). On the contrary, the transitory activation of the genes that express PAI-1 via the activation of PPAR-γ in fatty liver disease could be implicated in the increased risk of CVD due to a decrease in fibrinolytic activity (82). PPAR-α performs biological functions not only in the liver, but also in the skeletal muscle, heart, endothelial cells, platelets, macrophages and lymphocytes (83). Activators of PPAR-α provide vascular protection by suppressing the production of platelet-derived growth factor as much as in megakaryocytes as platelets (83).

4. Conclusion

MS is a pathophysiological state characterized by insulin resistance, dyslipidemia and arterial hypertension, with a major predisposition for T2DM and CVD. The main molecular alterations present in MS create a pro-inflammatory state which, along with increased free fatty acids in adipose tissue, favors the oxidation of LDL and its accumulation in the subendothelium. This vascular environment produces a state of platelet hyperreactivity, favoring the formation of arterial thrombosis. Indeed, the molecular interactions of adipose tissue, endothelial cells and platelets and their participation in gene expression represent important pathophysiological aspects of MS. The expression of PPARs, C/EBPs, ChREBP and SREBPs depends on their DNA-binding affinity, proteasomal degradation and on activators or repressors developed by adipose tissue, endothelium and platelets during MS. Ultimately, these findings indicate that the transcription factors may represent valid, modifiable binding sites where their controlled regulation can be translated into the gradual improvement of MS.

References

- 1. Palomo I, Contreras A, Alarcón M, *et al*: Elevated concentration of asymmetric dimethylarginine (ADMA) in individuals with metabolic syndrome. Nitric Oxide 24: 224-228, 2011.
- 2. Mujica V, Leiva E, Icaza G, *et al*: Evaluation of metabolic syndrome in adults of Talca city, Chile. Nutr J 7: 1-6, 2008.
- 3. Palomo I, Moore-Carrasco R, Alarcon M, *et al*: Pathophysiology of the proatherothrombotic state in the metabolic syndrome. Front Biosci (Schol Ed) 2: 194-208, 2010.
- 4. Huang P: A comprehensive definition for metabolic syndrome. Dis Model Mech 2: 231-237, 2009.
- Palomo I, Moore-Carrasco R, Alarcón M, et al: Fisiopatología del estado protrombótico en el síndrome metabólico. Acta Med Colomb 34: 80-84, 2009.
- Sossa C: Estado protrombótico y síndrome metabólico. Acta Med Colomb 30: 140-143, 2005.
- Sozio M, Liangpunsakul S and Crabb D: The role of lipid metabolism in the pathogenesis of alcoholic and nonalcoholic hepatic steatosis. Semin Liver Dis 30: 378-390, 2010.
- 8. Duan S, Usher M and Mortensen R: PPARs: the vasculature, inflammation and hypertension. Curr Opin Nephrol Hypertens 18: 128-133, 2009.
- Palomo I, Alarcón M, Moore-Carrasco R and Argilés J: Hemostasis alterations in metabolic syndrome (Review). Int J Mol Med 18: 969-974, 2006.
- Palomo I, Jaramillo J, Alarcón M, et al: Increased concentrations of soluble vascular cell adhesion molecule-1 and soluble CD40L in subjects with metabolic syndrome. Mol Med Report 2: 481-485, 2009.
- Palomo I, Gutiérrez C, Alarcón M, et al: Increased concentration of plasminogen activator inhibitor-1 and fibrinogen in individuals with metabolic syndrome. Mol Med Report 2: 253-257, 2009.
- Pan Y and Pratt C: Metabolic syndrome and its association with diet and physical activity in US adolescents. J Am Diet Assoc 108: 276-286, 2008.
- Martínez JA, Moreno MJ, Marques-Lopes I and Martí A: Causes of obesity. An Sist Sanit Navar 25: 17-27, 2002 (In Spanish).
- 14. Mujica V, Urzua A, Leiva E, *et al*: Intervention with education and exercise reverses the metabolic syndrome in adults. J Am Soc Hypertens 4: 148-153, 2010.
- Schnell M, Dominguez Z and Carrera C: Aspectos genéticos, clínicos y fisiopatológicos de síndrome metabólico. An Venez Nutr 20: 92-98, 2007.
- 16. Zheng Z, Zhang C and Zhang K: Role of unfolded protein response in lipogenesis. World J Hepatol 2: 203-207, 2010.

- 17. Bocher V, Pineda-Torra I, Fruchart J and Staels B: PPARs: transcription factors controlling lipid and lipoprotein metabolism. Ann NY Acad Sci 967: 7-18, 2002.
- Moreno MJ and Martínez JA: El tejido adiposo: Adipose tissue: a storage and secretory organ. An Sist Sanit Navar 25: 29-39, 2002 (In Spanish).
- 19. Jay M and Ren J: Peroxisome proliferator-activated receptor (PPAR) in metabolic syndrome and type 2 diabetes mellitus. Curr Diabetes Rev 3: 33-39, 2007.
- Wagner K and Wagner N: Peroxisome proliferator-activated receptor beta/delta (PPARbeta/delta) acts as regulator of metabolism linked to multiple cellular functions. Pharmacol Ther 125: 423-435, 2010.
- Moore-Carrasco R, Poblete M, González O, et al: Peroxisome proliferator-activated receptors: Targets for the treatment of metabolic illnesses (Review). Mol Med Report 1: 317-324, 2008
- 22. Harmon G, Lam M and Glass C: PPARs and lipid ligands in inflammation and metabolism. Chem Rev 111: 6321-6340, 2011.
- Barish G, Narkar V and Evans R: PPAR delta: a dagger in the heart of the metabolic syndrome. J Clin Invest 116: 590-597, 2006.
- 24. Tordjman K, Semenkovich C, Coleman T, *et al*: Absence of peroxisome proliferator-activated receptor-alpha abolishes hypertension and attenuates atherosclerosis in the Tsukuba hypertensive mouse. Hypertension 50: 945-951, 2007.
- 25. Sugii S and Evans R: Epigenetic codes of PPARγ in metabolic disease. FEBS Lett 585: 2121-2128, 2011.
- Heikkinen S, Auwerx J and Argmann C: PPARγ in human and mouse physiology. Biochim Biophys Acta 1771: 999-1013, 2007.
- 27. Luconi M, Cantini G and Serio M: Peroxisome proliferator-activated receptor gamma (PPARgamma): is the genomic activity the only answer? Steroids 75: 585-594, 2010.
- 28. Barroso I, Gurnell M, Crowley V, *et al*: Dominant negative mutations in human PPARgamma associated with severe insulin resistance, diabetes mellitus and hypertension. Nature 402: 880-883, 1999.
- Varga T, Czimmerer Z and Nagy L: PPARs are a unique set of fatty acid regulated transcription factors controlling both lipid metabolism and inflammation. Biochim Biophys Acta 1812: 1007-1022, 2011.
- 30. Barish G, Atkins A, Downes M, *et al*: PPARdelta regulates multiple proinflammatory pathways to suppress atherosclerosis. Proc Natl Acad Sci USA 105: 4271-4276, 2008.
- 31. Tsukada J, Yoshida Y, Kominato Y and Auron P: The CCAAT/enhancer (C/EBP) family of basic-leucine zipper (bZIP) transcription factors is a multifaceted highly-regulated system for gene regulation. Cytokine 54: 6-19, 2011.
- 32. Nerlov C: The C/EBP family of transcription factors: a paradigm for interaction between gene expression and proliferation control. Trends Cell Biol 17: 318-324, 2007.
- 33. Olofsson L, Orho-Melander M, William-Olsson L, *et al*: CCAAT/ enhancer binding protein α (C/EBPα) in adipose tissue regulates genes in lipid and glucose metabolism and a genetic variation in C/EBPα is associated with serum levels of triglycerides. J Clin Endocrinol Metab 93: 4880-4886, 2008.
- 34. Ramji D and Foka P: CCAAT/enhancer-binding proteins: structure, function and regulation. Biochem J 365: 561-575, 2002.
- 35. Bennett C, Nsengimana J, Bostock J, *et al*: CCAAT/enhancer binding protein alpha, beta and delta gene variants: associations with obesity related phenotypes in the Leeds Family Study. Diab Vasc Dis Res 7: 195-203, 2010.
- 36. Ge Q, Nakagawa T, Wynn RM, Chook YM, Miller BC and Uyeda K: Importin-alpha protein binding to a nuclear localization signal of carbohydrate response element-binding protein (ChREBP). J Biol Chem 286: 28119-28127, 2011.
- 37. Sakiyama H, Wynn R, Lee W, *et al*: Regulation of nuclear import/export of carbohydrate response element-binding protein (ChREBP): interaction of an alpha-helix of ChREBP with the 14-3-3 proteins and regulation by phosphorylation. J Biol Chem 283: 24899-24908, 2008.
- 38. Wakil S and Abu-Elheiga L: Fatty acid metabolism: target for metabolic syndrome. J Lipid Res 50: 138-143, 2009.
- 39. Iizuka K and Horikawa Y: ChREBP: a glucose-activated transcription factor involved in the development of metabolic syndrome. Endocr J 55: 617-624, 2008.

- Tontonoz P, Kim J, Graves R and Spiegelman B: ADD1: a novel helix-loop-helix transcription factor associated with adipocyte determination and differentiation. Mol Cell Biol 13: 4753-4759, 1993
- 41. Lewis C, Griffiths B, Santos C, Pende M and Schulze A: Regulation of the SREBP transcription factors by mTORC1. Biochem Soc Trans 39: 495-499, 2011.
- Porstmann T, Griffiths B, Chung Y, et al: PKB/Akt induces transcription of enzymes involved in cholesterol and fatty acid biosynthesis via activation of SREBP. Oncogene 24: 6465-6481, 2005.
- 43. Bengoechea-Alonso M and Ericsson J: SREBP in signal transduction: cholesterol metabolism and beyond. Curr Opin Cell Biol 19: 215-222, 2007.
- 44. Ferré P and Foufelle F: SREBP-1c transcription factor and lipid homeostasis: clinical perspective. Horm Res 68: 72-82, 2007.
- 45. Li D and Mehta J: Intracellular signaling of LOX-1 in endothelial cell apoptosis. Circ Res 104: 566-568, 2009.
- Daub K, Seizer P, Stellos K, et al: Oxidized LDL-activated platelets induce vascular inflammation. Semin Thromb Hemost 36: 146-156, 2010.
- 47. Badimón L, Vilahur G and Padró T: Lipoproteins, platelets and atherothrombosis. Rev Esp Cardiol 62: 1161-1178, 2009.
- 48. Palomo I, Fuentes E, Moore-Carrasco R, *et al*: Consumption of fruits and vegetables in the prevention of endothelial damage. Rev Chil Nutr 38: 343-355, 2011 (In Spanish).
- Furukawa S, Fujita T, Shimabukuro M, et al: Increased oxidative stress in obesity and its impact on metabolic syndrome. J Clin Invest 114: 1752-1761, 2004.
- 50. Anastassios G, Nandini A and Greenberg A: Adipocytokines and insulin resistance. J Clin Endocrinol Metab 89: 447-452, 2004.
- 51. Kohler H: Insulin resistance syndrome: interaction with coagulation and fibrinolysis. Swiss Med Wkly 132: 241-252, 2002.
- 52. Rosen E, Eguchi J and Xu Z: Transcriptional targets in adipocyte biology. Expert Opin Ther Targets 13: 975-986, 2009.
- 53. Kallio P, Kolehmainen M, Laaksonen D, et al: Dietary carbohydrate modification induces alterations in gene expression in abdominal subcutaneous adipose tissue in persons with the metabolic syndrome: the FUNGENUT Study. Am J Clin Nutr 85: 1417-1427, 2007.
- 54. Van Dijk S, Feskens E, Bos M, *et al*: A saturated fatty acid-rich diet induces an obesity-linked proinflammatory gene expression profile in adipose tissue of subjects at risk of metabolic syndrome. Am J Clin Nutr 90: 1656-1664, 2009.
- 55. Uauy R, Nartínez J and Rojas C: Molecular nutrition, role of the PPAR system in lipidic metabolism and its importance in obesity and diabetes mellitus. Rev Med Chil 128: 437-446, 2000 (In Spanish).
- 56. Vernochet C, Peres S, Davis K, *et al*: C/EBPalpha and the corepressors CtBP1 and CtBP2 regulate repression of select visceral white adipose genes during induction of the brown phenotype in white adipocytes by peroxisome proliferator-activated receptor gamma agonists. Mol Cell Biol 29: 4714-4728, 2009.
- 57. Wajchenberg B, Nery M, Cunha M and Silva M: Adipose tissue at the crossroads in the development of the metabolic syndrome, inflammation and atherosclerosis. Arq Bras Endocrinol Metabol 53: 145-150, 2009.
- 58. Palomer X, Pérez A and Blanco-Vaca F: Adiponectin: a new link between obesity, insulin resistance and cardiovascular disease. Med Clin (Barc) 124: 388-395, 2005 (In Spanish).
- 59. Tsuchida A, Yamauchi T, Takekawa S, *et al*: Peroxisome proliferator-activated receptor (PPAR)alpha activation increases adiponectin receptors and reduces obesity-related inflammation in adipose tissue: comparison of activation of PPARalpha, PPARgamma, and their combination. Diabetes 54: 3358-3370, 2005.
- 60. Ketsawatsomkron P, Pelham CJ, Groh S, Keen HL, Faraci FM and Sigmund CD: Does peroxisome proliferator-activated receptorgamma (PPAR gamma) protect from hypertension directly through effects in the vasculature? J Biol Chem 285: 9311-9316, 2010.
- 61. Holvoet P: Relations between metabolic syndrome, oxidative stress and inflammation and cardiovascular disease. Verh K Acad Geneeskd Belg 70: 193-219, 2008.
- Letexier D, Peroni O, Pinteur C and Beylot M: In vivo expression of carbohydrate responsive element binding protein in lean and obese rats. Diabetes Metab 31: 558-566, 2005.

- 63. Feingold KR, Shigenaga JK, Patzek SM, Chui LG, Moser A and Grunfeld C: Endotoxin, zymosan, and cytokines decrease the expression of the transcription factor, carbohydrate response element binding protein, and its target genes. Innate Immun 17: 174-182, 2011.
- 64. Ziouzenkova O and Plutzky J: Retinoid metabolism and nuclear receptor responses: new insights into coordinated regulation of the PPAR-RXR complex. FEBS Lett 582: 32-38, 2008.
- 65. Rodríguez-Rodríguez E, Perea J, López-Sobaler A and Ortega R: Obesity, insulin resistance and increase in adipokines levels: importance of the diet and physical activity. Nutr Hosp 24: 415-421, 2009.
- 66. Wei X, Schneider J, Shenouda S, *et al*: De novo lipogenesis maintains vascular homeostasis through endothelial nitric-oxide synthase (eNOS) palmitoylation. J Biol Chem 286: 2933-2945, 2011.
- 67. Gharavi N, Baker N, Mouillesseaux K, *et al*: Role of endothelial nitric oxide synthase in the regulation of SREBP activation by oxidized phospholipids. Circ Res 98: 768-776, 2006.
- 68. Hastings N, Feaver R, Lee M, Wamhoff B and Blackman B: Human IL-8 regulates smooth muscle cell VCAM-1 expression in response to endothelial cells exposed to atheroprone flow. Arterioscler Thromb Vasc Biol 29: 725-731, 2009.
- 69. Zimman A, Mouillesseaux K, Le T, *et al*: Vascular endothelial growth factor receptor 2 plays a role in the activation of aortic endothelial cells by oxidized phospholipids. Arterioscler Thromb Vasc Biol 27: 332-338, 2007.
- 70. Fan Y, Wang Y, Tang Z, *et al*: Suppression of pro-inflammatory adhesion molecules by PPAR-delta in human vascular endothelial cells. Arterioscler Thromb Vasc Biol 28: 315-321, 2008.
- Huang T, Tran V, Roufogalis B and Li Y: Gypenoside XLIX, a naturally occurring PPAR-alpha activator, inhibits cytokineinduced vascular cell adhesion molecule-1 expression and activity in human endothelial cells. Eur J Pharmacol 565: 158-165, 2007.
- 72. Duan S, Usher M and Mortensen R: Peroxisome proliferator-activated receptor-gamma-mediated effects in the vasculature. Circ Res 102: 283-294, 2008.

- 73. Kleinhenz J, Kleinhenz D, You S, et al: Disruption of endothelial peroxisome proliferator-activated receptor-γ reduces vascular nitric oxide production. Am J Physiol Heart Circ Physiol 297: 1647-1654, 2009.
- Sugawara A, Uruno A, Kudo M, Matsuda K, Yang CW and Ito S: PPARγ agonist beyond glucose lowering effect. Korean J Intern Med 26: 19-24, 2011.
- 75. Palomo I, Toro C and Alarcón M: The role of platelets in the pathophysiology of atherosclerosis (Review). Mol Med Report 1: 170-184, 2008
- 76. Colwell J and Nesto R: The platelet in diabetes: focus on prevention of ischemic events. Diabetes Care 26: 2181-2188, 2003.
- 77. Schneider D: Factors contributing to increased platelet reactivity in people with diabetes. Diabetes Care 32: 525-527, 2009.
- 78. Bishop-Bailey D: The platelet as a model system for the acute actions of nuclear receptors. Steroids 75: 570-575, 2010.
- 79. Falcetti E, Flavell DM, Staels B, Tinker A, Haworth SG and Clapp LH: IP receptor-dependent activation of PPARgamma by stable prostacyclin analogues. Biochem Biophys Res Commun 360: 821-827, 2007.
- 80. Ray D, Spinelli S, O'Brien J, Blumberg N and Phipps R: Platelets as a novel target for PPARgamma ligands: implications for inflammation, diabetes, and cardiovascular disease. BioDrugs 20: 231-241 2006
- 81. Moraes L, Spyridon M, Kaiser W, *et al*: Non-genomic effects of PPARgamma ligands: inhibition of GPVI-stimulated platelet activation. J Thromb Haemost 8: 577-587, 2010.
- 82. Oishi K, Tomita T, Itoh N and Ohkura N: PPARγ activation induces acute PAI-1 gene expression in the liver but not in adipose tissues of diabetic model mice. Thromb Res 128: 81-85, 2011.
- 83. Hashizume S, Akaike M, Azuma H, *et al*: Activation of proliferator-activated receptor α in megakaryocytes reduces platelet-derived growth factor-BB in platelets. J Atheroscler Thromb 18: 138-147, 2011.