Association between the expression of T-cadherin and vascular endothelial growth factor and the prognosis of patients with gastric cancer

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Abstract. T-cadherin has been identified as a tumor-suppressor gene in several types of cancer. The present study aimed to investigate the association of the expression of T-cadherin with angiogenesis, and to evaluate its prognostic value for patients with primary gastric cancer. Gastric cancer tissues and matched adjacent tissues from 166 patients receiving surgical resection were included in the present study. The expression of T-cadherin was detected using immunohistochemistry, western blotting and reverse transcription-quantitative polymerase chain reaction. The expression of vascular epidermal growth factor (VEGF) was detected using immunohistochemistry, and its association with the expression of T-cadherin was analyzed. In addition, the association between the expression of T-cadherin and clinicopathological features were analyzed. The mRNA and protein expression levels of T-cadherin were significantly lower in the gastric cancer tissue compared with the corresponding adjacent normal tissue (P<0.05). The expression of VEGF was not associated with the expression of T-cadherin in the gastric cancer tissue. The decreased protein expression of T-cadherin correlated with smoking, larger tumor size (diameter, >4 cm), lymph node metastasis and a higher tumor-lymph node-metastasis stage (P<0.05 or P<0.01). However, the expression of T-cadherin was not correlated with gender, age, alcohol intake, Helecobacter pylori infection or differentiation (P>0.05). The multivariate analysis demonstrated that the expression of T-cadherin was an independent prognostic factor for the overall survival rate of patients with gastric cancer. This data suggested that the downregulation of T-cadherin may contribute to gastric cancer progression, representing a useful biomarker for predicting the

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biological behavior and prognosis of gastric cancer. However, no significant association was observed between the expression of VEGF and T-cadherin.

Introduction

Gastric cancer remains the fourth most common type of cancer and the second most common cause of cancer-associated mortality worldwide (1). The majority of cases of gastric cancer are diagnosed at an advanced stage of development and, in patients no longer suitable for surgery or in cases of post-surgery recurrence, no effective treatment is currently available. Chemotherapy and radiotherapy often cause serious side effects as, in addition to causing cancer cell death, they affect normal tissue cells (2).

Cell-cell adhesion determines cell polarity and is involved in cell differentiation and the maintenance of tissue homeostasis. This cell-cell adhesion is disturbed during oncogenesis, resulting in changes in signaling, loss of contact inhibition and altered cell migration (3). There is increasing evidence implicating cell-cell adhesion molecules in cancer, as potent suppressors or as proto-oncogenic proteins (4). T-cadherin, also termed cadherin-13 (CDH13), is an atypical member of the cadherin superfamily, which is anchored to cell membranes via glycosylphosphatidylinositol (GPI) anchors, rather than transmembrane domains (5). Several previous studies have led to T-cadherin being considered a tumor suppressor, as it is frequently silenced in various types of cancer, including hepatocellular carcinoma (6), colon carcinoma (7), gallbladder carcinoma (8), melanoma (9), lung cancer (10) and breast cancer (11). Previous studies have demonstrated that the re-expression of T-cadherin may suppress cell proliferation, angiogenesis and invasiveness, increase sensitivity to apoptosis and decrease tumor growth (3).

Tang *et al* (12) revealed that the expression of T-cadherin was significantly reduced in tumor tissue samples compared with the adjacent normal tissue, and can be used as a biomarker for the progression and prognosis of gastric cancer. However, its tumor-suppressor mechanism in gastric cancer remains to be elucidated. Angiogenesis is required for invasive tumor growth and metastasis, and constitutes an important stage in the control of cancer progression (13,14). A previous

study demonstrated that the hyperexpression of T-cadherin in melanoma cells suppresses the growth of tumor masses and angiogenesis *in vivo* (15). However, Hebbar *et al* (16) demonstrated that T-cadherin promotes tumor angiogenesis in breast cancer. Therefore, the association between the expression of T-cadherin and angiogenesis remains inconsistent.

The present study aimed to evaluate the expression of T-cadherin in gastric cancer, and analyze the association between the expression of T-cadherin and clinicopathological features in a larger number of patients. In addition, the association between the expression of T-cadherin and angiogenesis was investigated.

Materials and methods

Specimens and samples. Specimens of primary gastric cancer and matched adjacent normal tissue were obtained from 166 patients who underwent surgery at The Second Affiliated Hospital of College of Xi'an Jiaotong University (Xi'an, China), between 2010 and 2012. The mean patient age was 62.3 years (range, 41-82). The tissue samples were frozen in liquid nitrogen immediately following resection and rinsing with phosphate-buffered saline (PBS), and were maintained at -80°C until RNA and protein extraction. Data on the gender, age, smoking, alcohol intake, Helicobacter pylori (Hp) infection, tumor size, lymph node metastasis and the tumor-lymph node-metastasis (TNM) stage of the patients with gastric carcinoma was also obtained. The International Union against Cancer TNM staging system (17) was used to classify the patients. Information on the survival rates of the 166 patients was followed up through written and telephone communication. The present study was approved by the Ethics Committee of The Second affiliated hospital of College of Xi'an Jiaotong University.

Reverse transcription-quantitative polymerase chain reaction (RT-qPCR). Total RNA was extracted from the gastric cancer and adjacent normal tissue samples using a TRIzol kit (Gibco BRL, Grand Island, NY, USA), according to the manufacturer's instructions. The concentration of the total RNA was measured by spectrophotometry (ND-1000; Thermo Fisher Scientific, Waltham, MA, USA). The RNA was subsequently reverse-transcribed into cDNA using reverse transcriptase reagents (dNTP mixture and RNase free dH₂O; Takara Bio Inc., Otsu, Shiga, Japan). RT-qPCR was performed using the SYBR GreenI fluorescent dye method (SYBR Premix Ex Taq™ II; Takara Bio Inc.) and a Rotor Gene 3000 RT-qPCR apparatus (Qiagan, Shanghai, China). The following primers from (Beijing AuGCT Biotechnology Co., Ltd., Beijing, China) were used: T-cadherin, forward 5'-GATGTTGGCAAGGTAGTCGAT-3' and reverse 5'-GCT CCCTGTGTTCTCATTGAT and β-actin, forward 5'-ATCGTG CGTGACATTAAGGAGAAG-3' and reverse 5'-AGGAAGGA AGGCTGGAAGAGTG-3'. The expression of β-actin was used as an internal control to assess the relative expression of T-cadherin. The PCR reaction was performed in a final volume of 20 µl, which contained 10 µl SYBR Premix Ex TaqTM II, $1 \mu l$ cDNA, $0.5 \mu l$ of each primer and $8 \mu l$ enzyme-free water. The PCR conditions were as follows: Predenaturing at 95°C for 3 min, followed by 40 cycles of denaturation at 95°C for 10 sec and annealing/extension at 60°C for 30 sec. The amplification specificity was assessed by melting curve analysis. The PCR products were run on a 2% agarose gel (Sigma-Aldrich, St. Louis, MO, USA) stained with ethidium bromide (Shaanxi Pioneer Biotech Co., Ltd., Xi'an, China) and were observed to be 177 base pairs in size. The 2-ΔΔCT method (18) was used to calculate the relative expression levels of the target gene produced, using a Bio-Rad Real-Time analysis system (Bio-Rad Laboratories, Hercules, CA, USA).

Western blotting. The cytosolic protein extraction from the gastric cancer and adjacent normal tissuse samples were performed, as described previously. Samples of 50 μ g protein were mixed with gel loading buffer (Beyotime Institute of Biotechnology, Shanghai, China), boiled for 5 min and loaded onto 8 or 10% polyacrylamide gels (Shaanxi Pioneer Biotech Co., Ltd.). Electrophoresis was performed and the proteins were transferred onto nitrocellulose membranes (Millipore, Billerica, MA, USA). Non-specific antibody binding was blocked by pre-incubation of the membranes with 1X Tris-buffered saline (TBS; Beijing ComWin Biotech Co., Ltd., Beijing, China), containing 5% non-fat milk (Yili Group, Inner Mongolia, China) for 2 h at room temperature. The membranes were incubated overnight at 4°C with primary antibodies against human T-cadherin (rabbit polyclonal; 1:1,000; ABT121; Millipore) or β-actin (mouse monoclonal; 1:1,000; sc-47778; Santa Cruz Biotechnology, Inc., Santa Cruz, CA, USA) in 1X TBS, containing 5% non-fat milk. Following washing with TBS for 5 min 3 times, the membranes were incubated with horseradish peroxidase-conjugated anti-rabbit immunoglobulin G (1:4,000; sc-2004; Santa Cruz Biotechnology, Inc.) for 2 h at room temperature. The bands were visualized using a SuperSignal Substrate Chemiluminescence kit (Millipore).

Immunohistochemistry. Paraffin-embedded (Sigma-Aldrich) tissue sections were de-waxed in xylene (Xi'an Chemical Reagent Factory, Xi'an, China) and rehydrated with a graded ethanol (Xi'an Chemical Reagent Factory) series. The slides used were from Wuhan Boster Biological Technology Ltd. (Wuhan, China). The tissue sections were subsequently treated with 3% hydrogen peroxidase (Zhongshan Jinqiao Bio., Beijing, China) for 15 min at room temperature, followed by incubation overnight at 4°C with diluted primary antibodies against T-cadherin (1:100) or vascular epidermal growth factor (VEGF; mouse monoclonal; 1:100; sc-7269; Santa Cruz Biotechnology, Inc.,). The tissue sections were then incubated with secondary antibody. Specific reactivity was detected using a 3,3'-Diaminobenzidine Tetrahydrochloride kit (Zhongshan Jinqiao Bio.) and counterstained with hematoxylin (Sigma-Aldrich). The immunostaining was assessed by two independent pathologists in a blinded-manner. The slide used as a negative control was incubated with PBS rather than primary antibody. The positive controls used were cardiac muscle tissues known to exhibit high protein expression levels of T-cadherin. The cells were considered positive for the protein expression of T-cadherin when the cell membrane was stained. Each slide was evaluated in five randomly selected fields with an E100 microscope (Nikon, Tokyo, Japan) at a magnification of x400, and 100-200 cells per field were counted. The immunohistochemical (IHC) scores comprise the product of

Table I. Correlation between the expression of T-cadherin and clinicopathological variables in gastric cancer.

Variable	Case (n)	Expression of T-cadherin			
		Low (n)	High (n)	χ^2	P-value
Gender					
Male	108	52	56	0.052	0.820
Female	58	29	29		
Age (years)					
<60	80	42	38	0.848	0.357
≥60	86	39	47		
Smoking					
Yes	69	43	26	8.643	0.003
No	97	38	59	0.043	
Alcohol consumption					
Yes	99	45	54	1.096	0.295
No	67	36	31	1.090	
<i>Hp</i> infection					
Positive	122	58	64	0.290	0.590
Negative	44	23	21	0.290	
Tumor size (cm)					
≤4	99	39	60	8.677	0.003
>4	67	42	25	0.077	
Differentiation					
Well-moderate	63	25	38	3.375	0.066
Poor	103	56	47	3.373	
Histopathological type					
Intestinal	99	49	50	0.048	0.826
Diffuse	67	32	35		
Lymph node metastasis					
Yes	115	63	52	5.371	0.020
No	51	18	33	3.371	
TNM stage					
I-II stage	79	28	51	10.755	0.001
III-IV stage	87	53	34	10./33	

Hp, Helicobacter pylori; TNM, tumor-lymph node-metastasis.

the scores of intensity (0, no staining; 1, weakly stained; 2, moderately stained; 3, markedly stained) and area (0, <5%; 1, 5-25%; 2, 25-50%; 3, >50%) of the staining signals. Based on the expression of T-cadherin, the gastric cancer tissues were divided into two groups: A low T-cadherin expression group (IHC scores≤3) and the high T-cadherin expression group (IHC scores>3). The same method was used for the expression of VEGF, which was also divided into low and high expression groups.

Statistical analysis. All statistical analyses were performed using SPSS 13.0 software (SPSS, Inc., Chicago, IL, USA). The mRNA and protein expression levels of T-cadherin in the gastric cancer and matched adjacent tissues were analyzed using a paired sample t-test. The association

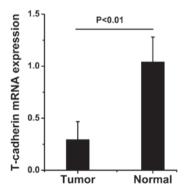


Figure 1. Reverse transcription-quantitative polymerase chain reaction analysis of the expression levels of T-cadherin in gastric cancer tissue samples. The relative mRNA expression of T-cadherin was significantly lower in the gastric cancer tissues compared with the corresponding normal tissues. The data are expressed as the mean \pm standard deviation; P<0.01.

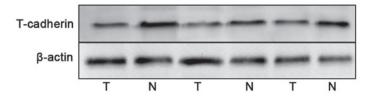


Figure 2. Protein expression levels of T-cadherin in gastric cancer, determined by western blotting. The protein expression levels of T-cadherin were markedly decreased in gastric tumor tissues compared with the corresponding adjacent normal tissues. T, gastric cancer tissue; N, matched non-tumorous gastric mucosa.

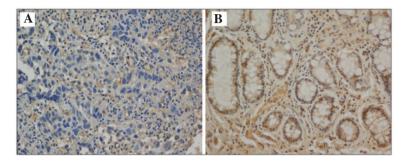


Figure 3. Protein expression of T-cadherin in (A) gastric cancer tissue and (B) surrounding normal tissue, as detected by immunohistochemistry with 3,3'-diaminobenzidine-tetrahydrochloride-dihydrate staining. Nuclei are stained blue with hematoxylin. The images demonstrated strong positive brown staining of T-cadherin expression in the cell membrane of gastric cancer cells and the adjacent normal tissue exhibited low expression levels. Magnification, x400.

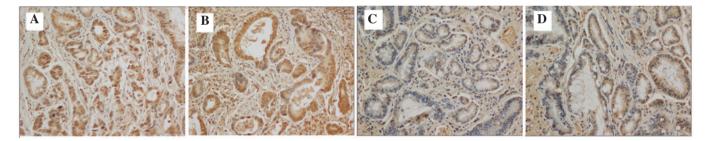


Figure 4. Correlation between the expression levels of T-cadherin and VEGF in gastric cancer tissues, detected by immunohistochemistry with 3,3'-diaminobenzidine-tetrahydrochloride-dihydrate staining. Nuclei are stained blue with hematoxylin. (A) Expression of T-cadherin in 'patient 1'. (B) Expression of VEGF in 'patient 1'. (C) Expression of T-cadherin in 'patient 2'. (D) Expression of VEGF in 'patient 2'. VEGF, vascular endothelial growth factor. The images demonstrated that the expression levels of VEGF were not associated with the expression of T cadherin. Magnification, x400.

between the protein expression levels of T-cadherin and various clinicopathological characteristics were assessed using a χ^2 test. The overall survival curves were calculated using the Kaplan-Meier method and were analyzed using the log-rank test. A Cox Proportional-Hazards regression model was produced to determine which variables demonstrated individual prognostic value in determining patient survival rates. Cox regression analysis was performed at multivariate levels. P<0.05 was considered to indicate a statistically significant difference and data are expressed as the mean \pm standard deviation.

Results

Expression levels of T-cadherin analyzed by RT-qPCR, western blotting and immunochemistry. The mRNA expression levels of T-cadherin in the tumor tissues and paired adjacent normal tissues were detected by RT-qPCR. The protein expression levels of T-cadherin were detected by western blotting and immunochemistry. The mRNA and protein expression levels of T-cadherin were significantly

reduced in the gastric cancer tissue samples compared with the adjacent normal tissue samples (Figs. 1-3).

Clinical and pathological significance of the expression of T-cadherin in gastric cancer. The present study demonstrated that T-cadherin was expressed in the cell membrane and that the expression levels varied in the gastric cancer tissue and the adjacent normal tissue samples (Fig. 3). Among the 166 gastric cancer samples, 87 samples exhibited high expression levels, whereas the remaining 81 cases exhibited low expression levels (Table I). The adjacent normal tissue samples revealed the most marked levels of T-cadherin positive staining (Fig. 3). Decreased protein expression levels of T-cadherin were found to correlate with smoking, large tumor size (diameter >4 cm), lymph node metastasis and a higher TNM stage (P<0.05 or P<0.01). However, the protein expression levels of T-cadherin did not correlate with gender, age, alcohol intake, Hp infection or differentiation (P>0.05; Table I).

Correlation between the expression levels of T-cadherin and VEGF. The present study determined the expression levels of

Table II. Correlation between the expression levels of T-cadherin and VEGF in gastric cancer.

	T-cadherin			
VEGF	Low	High	r-value	P-value
Low High	64 102	29 55	-0.084	0.283

VEGF, vascular endothelial growth factor.

Table III. Cox regression analysis of the association between clinicopathological variables, expression of T-cadherin and patient survival rates in gastric cancer.

Variable	HR (95% CI)	P-value	
Gender (male/female)	0.943 (0.535-1.663)	0.839	
Age (<60 or ≥60 years)	1.005 (0.649-1.556)	0.983	
Smoking (no/yes)	1.923 (1.064-3.475)	0.030	
Alcohol (no/yes)	0.644 (0.360-1.152)	0.138	
Hp infection (no/yes)	0.877 (0.505-1.523)	0.640	
Tumor size (≤4 cm/>4 cm)	1.761 (1.080-2.872)	0.023	
Differentiation (well or moderate/poorly)	2.094 (1.257-3.488)	0.005	
Histopathological (intestinal/diffuse)	0.840 (0.529-1.333)	0.460	
Lymph node metastasis (no/yes)	2.121 (1.176-3.825)	0.012	
TNM stages (I-II/III-IV)	2.159 (1.293-3.605)	0.003	
T-cadherin expression (low/high)	0.384 (0.230-0.640)	0.000	

Hp, Helicobacter pylori; TNM, tumor-node-metastasis; HR, hazard ratio; CI, confidence interval.

VEGF using immunochemistry in the tissue samples from 166 cases of gastric cancer. The result revealed that the expression levels of VEGF were not associated with the expression of T-cadherin (P>0.05; Fig. 4, Table II).

Correlation between the expression of T-cadherin and the overall survival rates of patients with gastric cancer. The patients with high expression levels of T-cadherin in their tissue sample had a significantly improved overall survival rate compared with patients exhibiting low expression levels of T-cadherin (P<0.01; Fig. 5). The multivariate Cox regression analysis demonstrated that smoking, smaller tumor sizes (diameter <4 cm), no lymph node metastasis, good differentiation, a lower TNM stage and higher expression levels of T-cadherin were closely associated with a higher overall survival rate and were independent risk factors for gastric cancer (P<0.05; Table III). However, the survival rate did not correlate with gender, age, alcohol intake, Hp infection or histopathological type (P>0.05).

Discussion

Previous studies have demonstrated that cancer progression is a multi-step process, in which cell-cell adhesion is important for the development of recurrent, invasive and distant metastasis (19). Multiple lines of evidence have indicated

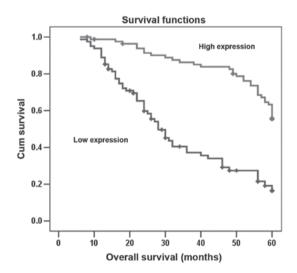


Figure 5. Kaplan-Meier survival curves demonstrating the overall survival rates of 166 patients with gastric cancer, and the association between surival rates and the expression of T-cadherin. The patients in the low T-cadherin expression group were found to have significantly lower survival rates compared with those in the high T-cadherin expression group (log-rank test, P<0.001). Cum, cumulative.

that alterations in the adhesion properties of cancer cells is vital in the development and progression of cancer. Loss of intercellular adhesion allows malignant cells to escape from their site of origin, degrade the extracellular matrix and acquire a more invasive and metastatic phenotype (20). Early evidence demonstrated certain adhesion molecules, which have been implicated in cancer as putative tumor suppressors, including E-cadherin (21). However, there certain adhesion molecules are also considered as pro-oncogenic proteins, including P-cadherin (22) and N-cadherin (23). CDH13, also termed T- or H-cadherin, is the only cadherin known to be membrane-anchored via a GPI anchor, rather than a transmembrane domain. The human CDH13 gene is often silenced in several types of cancer and it has long been considered to act against carcinogenesis (3). It has been suggested that T-cadherin may inhibit tumor progression, including proliferation, invasion and angiogenesis, through multiple pathways, including the Akt and SET7/9-p53 pathways (8,24). However, only one previous study has investigated the correlation between the expression of T-cadherin and the clinicopathological features of gastric cancer, and this included a relatively small sample size (12).

Previous studies have revealed that the loss of T-cadherin is associated with methylation, and that treatment with a demethylating agent reactivates its expression (6,7,25). The present study demonstrated that the T-cadherin protein was expressed in the cell membrane of the normal and the malignant gastric mucosa. The mRNA and protein expression levels of T-cadherin were reduced in the gastric cancer tissue samples compared with the corresponding normal tissue samples. In addition, its expression exhibited a close association with the clinicopathological features of gastric cancer. The decreased protein expression of T-cadherin was associated with a larger tumor size, lymph node metastasis and a higher TNM stage, however it was not associated with gender, age, alcohol intake differentiation or histopathological type. These results suggested that T-cadherin may be important in tumor growth, invasion and metastasis. This result was consistent with previous studies. Yan et al demonstrated an increased expression of T-cadherin and reduced cell proliferation in HepG2 cells (26), Philippova et al revealed that a loss of T-cadherin increases the metastatic potential and aggressiveness of squamous cell carcinoma (27) and Hebbard et al demonstrated that the loss of T-cadherin promotes tumor angiogenesis and metastasis in breast cancer (12). VEGF is the key mediator of angiogenesis and metastasis in cancer, and is upregulated by the expression of oncogenes (28,29). Although the loss of T-cadherin has been considered to exhibit a prometastatic effect, the underlying mechanism remains to be elucidated. The present study assessed the expression of VEGF in gastric cancer and analyzed its association with the expression T-cadherin. The result demonstrated that the expression of VEGF was not associated with the expression of T-cadherin in the gastric cancer tissue samples, which suggested that other mechanism are responsible for prometastatic effects following loss of T-cadherin loss.

Liu *et al* indicated that T-cadherin may be used as an independent prognostic biomarker for bladder transitional cell carcinoma (30), wheras Kim *et al* demonstrated that T-cadherin may be used as a prognostic marker for patients with non-small cell lung cancer (10). In the present study, the survival rates of the patients revealed that those with high expression levels of T-cadherin had a significantly higher postoperative survival

rate compared with those exhibiting low expression levels of T-cadherin. Therefore, patients with gastric cancer and reduced expression levels of T-cadherin may be a high-risk group with poor survival rates and may require more aggressive additional post-surgical systemic therapy. These results suggested that measurement of the protein expression levels of T-cadherin can assist in monitoring patient condition. According to the Cox regression analysis, a smaller tumor size, lack of lymph node metastasis, good level of differentiation, lower TNM stage and higher expression levels of T-cadherin were closely associated with increased overall survival rates, and the positive expression of T-cadherin appeared to be the most important independent prognostic predictor in gastric cancer.

Previous studies have demonstrated that smoking (31), alcohol consumption (32) and Hp infection (33) are risk factors for gastric cancer. Therefore, the present study investigated the possible associations between the expression of T-cadherin and smoking, alcohol and Hp infection. The results revealed that patients with a history of smoking exhibited low expression levels of T-cadherin. However, alcohol and Hp infection had no effect on the expression of T-cadherin. In addition, the survival rates of the patients suggested that those with a history of smoking had lower postoperative survival rates compared with the patients without a history of smoking, however, the overall survival rates were not associated with alcohol and Hp infection.

In conclusion, the expression of T-cadherin was found to decrease in gastric cancer and the expression levels were associated with smoking, tumor size, lymph node metastasis and TNM stage. The expression of T-cadherin may serve as an independent prognostic predictor and be used as a biomarker to predict the progression and prognosis of gastric cancer, however, no clear association was observed between the expression levels of VEGF and T-cadherin. In addition the results suggested that T-cadherin may exhibit a tumor suppressor function in gastric cancer and has the potential to be used as a target for therapeutic interventions for gastric cancer.

Acknowledgements

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