

# Differentiation of osteoblast-like cells and ectopic bone formation induced by bone marrow stem cells transfected with chitosan nanoparticles containing plasmid-BMP2 sequences

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Received August 11, 2015; Accepted November 1, 2016

DOI: 10.3892/mmr.2017.6128

**Abstract.** The present study investigated the efficiency of the use of chitosan nanoparticles containing plasmid-bone morphogenetic protein 2 (pBMP2) sequences (CNPBs) to induce the differentiation of bone marrow stem cells (BMSCs) into osteoblast-like cells that may be able to promote ectopic bone formation. pBMP2s were constructed, and chitosan nanoparticles were incubated with 50, 100 or 200  $\mu\text{g/ml}$  pBMP2. BMSCs were collected from the tibiae and femurs of 6-week old rats, cultured and treated with the CNPBs or 200  $\mu\text{g/ml}$  pBMP2 as a positive control. Transfection efficiency was confirmed using the green fluorescent protein assay. Histological staining methods, including alkaline phosphatase, Wright's and von Kasso staining, were used to identify features of osteoblast-like cells differentiated from BMSCs. Expression levels of the markers of osteoblasts, such as alkaline phosphatase, osteoprotegerin, osteocalcin and osteopontin, were determined to verify the differentiation of BMSCs into osteoblast-like cells. Ectopic bone formation was observed following the integration of polyglycolic acid (PGA) scaffolds with CNPBs and BMSCs, which were implanted into the dorsal muscles of Sprague-Dawley rats. Exposure to CNPBs led to the transfection of BMSCs with BMP2. The transfected BMSCs possessed the characteristic phenotypes of osteoblasts. The expression levels of alkaline phosphatase, osteoprotegerin, osteocalcin and osteopontin were significantly higher in the transfected cells compared with the control group, particularly

the CBP200 group. PGA scaffolds integrated with BMSCs and CNPBs induced ectopic bone formation, as changes in the morphology of cells were observed using histological staining. Therefore, CNPBs may be a promising method of promoting the formation of novel bone tissue.

## Introduction

Bone tissue engineering provides a method for repair of bone defects (1,2). The rapid development of bone tissue engineering in combination with neogenetic osteoblast-like cells, bone tissues induced by seed cells, genetic carrier delivery systems, bioscaffolds and growth factors may have the potential to replace autogenous bone transplantation in segmental bone repair (3,4). Previous studies have used techniques on the nanometer scale for tissue engineering (5,6).

Bone marrow stem cells (BMSCs) may be induced to differentiate into osteoblast-like cells during osteogenesis. However, there is a limited number of BMSCs in the bone marrow, and BMSCs alone are unable to induce new bone formation (7). Therefore, it is difficult to use only BMSCs to promote the repair of bone defects. In order to induce BMSCs to differentiate into osteoblast-like cells, bone growth factors are required (8).

Bone morphogenetic protein 2 (BMP2) is a bone growth factor that may be used to induce osteogenesis (9). However, it is difficult to sustain the slow release of high concentrations of exogenous bone growth factors in the region of the bone defect. In addition, high doses of exogenous bone growth factors may lead to adverse side-effects in the patient. Therefore, it is crucial to identify a method of releasing BMP2 slowly, and at high concentrations, within the regions of defective bone by using genetic carriers (10).

Non-viral carriers are safer alternatives to viral carriers (11). Chitosan is a biocompatible and bioresorbable polymer of N-acetylglucosamine and glucosamine, and it is extensively used as a carrier for bone growth factors (12). Favorable characteristics of chitosan for this purpose include its easy availability, low cytotoxicity, low immunogenicity, excellent biocompatibility and improved biodegradability. A previous study confirmed that chitosan may deliver exogenous BMP2 into various seed cells, such as BMSCs (13). In addition, chitosan nanoparticles are advantageous for delivering bone growth factors or gene sequences into seed cells in order to

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*Abbreviations:* BMP2, bone morphogenetic protein 2; CNPBs, chitosan nanoparticles containing plasmid-bone morphogenetic protein 2 sequences; BMSC, bone marrow stem cells; pBMP2s, plasmid-BMP2s; PGA, polyglycolic acid; GFP, green fluorescent protein

*Key words:* chitosan, BMP2, osteoblast, bone marrow, bone tissue engineering, bone repair

induce the formation of bone tissues with high efficiency (14) due to their excellent properties of infiltration and absorption. In the course of delivery, DNA combined with chitosan nanoparticles was effectively protected, and consequently, genes and proteins remained functional for longer periods of time.

In our previous study, chitosan nanoparticles containing plasmid-BMP2 (pBMP2) sequences (CNPBs) were constructed through using re-coacervation and gene recombination techniques (15). Our previous study determined that the average diameter of chitosan nanoparticles was  $90\pm 20$  nm. CNPBs with higher enveloping ratios were able to effectively protect BMP2 genes. Following incubation of BMSCs with CNPBs for 12 days, cells had maintained their normal morphology and function (15).

In the present study, CNPBs were constructed with different concentrations of pBMP2, specifically containing 50  $\mu\text{g/ml}$  (CPB50), 100  $\mu\text{g/ml}$  (CPB100) or 200  $\mu\text{g/ml}$  (CPB200) pBMP2. Following treatment with CNPB, groups were separately phagocytized by BMSCs. The transfection efficiency of the CNPBs was confirmed, and features of osteoblast-like cells derived from BMSCs were observed through histological staining, including alkaline phosphatase, Wright's and von Kasso staining. Expression levels of osteoblast-associated molecules, such as alkaline phosphatase (ALP), osteoprotegerin (OPG), osteocalcin (OC) and osteopontin (OPN), were detected and analyzed in the differentiated osteoblast-like cells. Ectopic bone formation was observed following the integration of polyglycolic acid (PGA) scaffolds with CNPBs and BMSCs, which were implanted into the dorsal muscles of Sprague-Dawley rats.

## Materials and methods

**Ethics statement.** All animals used in this study were provided by the experimental animal center of Zhejiang University, (Zhejiang, China). The animal use and care protocol was approved by the Institutional Animal Use and Care Committee of Zhejiang University. Experimenters were approved through the Health Department of Zhejiang Province for experiments with animals (certificate no. x0901616).

**Culture of BMSCs and construction of CNPBs.** A total of 10 female Sprague Dawley rats were anesthetized with pentobarbital sodium (50 mg/kg, Sigma-Aldrich Merck Millipore, Darmstadt, Germany) and sacrificed by cervical dislocation (6-weeks old; weight, 180-220 g; sanitary degree). The animals were kept in air-circulated housing a 14 h light/10 h dark cycle at  $22\pm 2^\circ\text{C}$ . Bones were dissected from the rats using a surgical knife. Bone marrow cavities of tibiae and femoral bones were rinsed using phosphate buffered saline (PBS). Cells were centrifuged at  $300 \times g$  for 5 min at  $37^\circ\text{C}$  and cultured in Dulbecco's modified Eagle's medium (DMEM; GE Healthcare Life Sciences, Logan, UT, USA) containing 10% fetal bovine serum (FBS; Gibco; Thermo Fisher Scientific, Inc., Waltham, MA, USA). Next, BMSCs were subjected to digestion with 0.25% trypsin (Gibco; Thermo Fisher Scientific, Inc.), cells were cultured until passage 3 and used in the subsequent experiments. In our previous study (15), CNPBs were constructed using re-coacervation and gene recombination

techniques. *Escherichia coli* bacteria were transfected with plasmids (cat. no. 6085-1; Addgene, Cambridge, MA, USA) that included an insertion of BMP2 cDNA, and extracted plasmid-DNA was subjected to restriction enzyme analysis. The successful insertion of BMP2 cDNA fragments was confirmed through DNA sequencing, performed by the laboratory in Zhejiang University (Hangzhou, China). Purified pBMP2 was dissolved into  $\text{Na}_2\text{SO}_4$  solution. The concentrations of pBMP2 used were 50, 100 or 200  $\mu\text{g/ml}$ . Subsequently, chitosan was mixed with the different concentrations of pBMP2 to form CNPBs (CPB50, CPB100 and CPB200, respectively). Chitosan was purchased from Sigma-Aldrich; Merck Millipore (50 g; cat. no. 448877) (15).

**Transfection efficiency of CNPBs.** Experimental groups were established as follows: i) Blank control (untreated BMSCs); ii) positive control (200  $\mu\text{g/ml}$  pBMP2); iii) CPB50 (chitosan + 50  $\mu\text{g/ml}$  pBMP2); iv) CPB100 (chitosan + 100  $\mu\text{g/ml}$  pBMP2); and v) CPB200 (chitosan + 200  $\mu\text{g/ml}$  pBMP2). BMSCs were seeded into 6-well culture plates ( $5.0 \times 10^5$  cells/well) for 24 h. Following the removal of DMEM, BMSCs were rinsed using 1 ml DMEM. Next, pBMP2 and the different CNPB concentrations were added separately into the experimental wells for a 6-h incubation with Lipofectamine 2000 (Invitrogen; Thermo Fisher Scientific, Inc.). Green fluorescent protein (Guduo Corporation, Shanghai, China) was used as reporter. Following the removal of the transfection solution, BMSCs were continuously cultured with DMEM containing 1 ml 10% FBS for 24 h. BMSCs transfected with CNPBs were photographed using an inverted fluorescence microscope (XD30-RFL; Sunny Instruments Co., Ltd., Ningbo, China). The number of, and total area occupied by, the cells in the micrographs were analyzed using ImageJ version 1.42 (National Institutes of Health, Bethesda, MD, USA).

**Cellular staining.** Alkaline phosphatase staining, Wright's staining, and von Kossa staining were used for identification of osteoblast-like cells differentiated from the transfected BMSCs. Experimental groups included: i) Blank control; ii) CPB50; iii) CPB100; and iv) CPB200. Initially, BMSCs were seeded into 6-well culture plates ( $1.0 \times 10^5$  cells/well) and maintained overnight. Next, DMEM was removed, cells were rinsed using 1 ml DMEM, the various concentrations of CNPB were added for the aforementioned specific treatment groups, and the cells were cultured for 24 h at  $37^\circ\text{C}$ .

For alkaline phosphatase staining, the transfected BMSCs were fixed for 10 min in 4% paraformaldehyde and then rinsed using distilled water for 5-10 min. Cells were immersed in alkaline phosphatase solution for 25 min at  $37^\circ\text{C}$ . Finally, cells were rinsed using distilled water for 5-10 min.

For Wright's staining, the transfected BMSCs were rinsed with PBS and fixed in methanol for 3-5 min. Cells were treated with Wright's staining solution for 2 min. Subsequently, transfected BMSCs were treated with Wright's phosphate buffer solution for 4-10 min and rinsed using distilled water.

Von Kossa staining was performed after transfected BMSCs were fixed in 4% paraformaldehyde for 15 min. The cells were then rinsed with distilled water. Then, cells were treated in 1% silver nitrate solution and incubated

Table I. Primers of ALP, OPG and OC.

Primers	Sequences (5'-3')	Length (bp)
Actin	F: CTAAGGCCAACCGTGAAA; R: TGGAAGGTGGACAGTGAG	724
ALP	F: GGTGGACGCAAAAATTTTCAT; R: ATGCCTTGATCGGTTTGTTC	379
OPG	F: CGACTGGAGAGCGAAGAC; R: CTAAGCAATGTTGGCGTA	364
OC	F: GAGGACCCTCTCTCTGCTCA; R: AGCTGTGCCGTCATACTTT	405
OPN	F: CATCAGAGCCACCACTTTCA R: TCAGGGCCCAAAACACTATC	273

ALP, alkaline phosphatase; OPG, osteoprotegerin; OC, osteocalcin; OPN, osteopontin; F, forward, R, reverse.

Table II. Quantity and total area of fluorescence particles in every group following transfection of bone marrow stem cells with chitosan nanoparticles containing plasmid-bone morphogenetic protein 2 sequences.

Treatment Groups	Count	Total area ( $\mu\text{m}^2$ )
Positive control	133.3 $\pm$ 45.0	9,189.3 $\pm$ 5,632.1
CPB50	401.3 $\pm$ 132.8	33,494.0 $\pm$ 19,076.8
CPB100	1,997.3 $\pm$ 490.7	250,702.6 $\pm$ 20,510.9
CPB200	1,162.0 $\pm$ 105.1	219,652.0 $\pm$ 65,961.3

Statistical analysis showed that there was a significant difference in particle quantity among the groups ( $P=0.000$ ). There was a significant difference in total areas among groups ( $P=0.000$ ). CPB50/100/200, chitosan nanoparticles containing plasmid-bone morphogenetic protein 2 at concentrations of 50, 100 and 200  $\mu\text{g}/\text{ml}$ .

under an ultraviolet lamp for 15 min. Finally, the cells were stained with 5% natrium hyposulfurosum solution (Shengrui Transmission Corporation Ltd., Xian, China) for 2 min.

**Reverse transcription-polymerase chain reaction (RT-PCR).** Experimental groups included: i) Blank control; ii) CPB50; iii) CPB100; and iv) CPB200. BMSCs were seeded into 6-well culture plates ( $1.0 \times 10^5$  cells/well) for 24 h culture, pBMP2 was added, and each of the CNPB treatment groups were set up in separate experimental wells for a 48 h transfection. RT-PCR was used to determine the mRNA expression levels of ALP, OPG and OC in osteoblast-like cells. mRNA was extracted using TRIzol reagent (Invitrogen; Thermo Fisher Scientific, Inc.). The primers used for RT-PCR of ALP, OPG and OC are presented in Table I. A PCR kit with DNA-free sensitive *Taq* DNA polymerase (Amresco, LLC, Solon, OH, USA) was used. The RT conditions were: 37°C for 1 h and 95°C for 5 min in order to deactivate M-MLV reverse transcriptase. The thermocycling conditions were as follows: 95°C for 15 min, 30 cycles at 95°C for 15 sec, 52°C 30 sec, 72°C for 30 sec and 72°C for 2 min.

**Western blotting.** Protein expression levels of OPN in cells was detected using western blot analysis. Cells were rinsed twice using PBS. Cell lysis buffer (100  $\mu\text{l}$ ) was added into each culture well (cat. no. P0013; Beyotime Institute of Biotechnology, Beijing, China). Cells were centrifuged for 5 min at 300  $\times g$  at 37°C. Polyvinylidene fluoride membranes (Merck Millipore) were rinsed using PBS with Tween-20 (PBST). The membranes were incubated with the rabbit anti-OPN primary antibody (cat. no. 000019-R; 1:500; CellChip Biotechnology Co., Ltd., Beijing, China) for 1 h at 37°C. PBST was used to wash the membranes 4 times for 10 min each. The membranes were then incubated for 1 h with a secondary biotin-labeled rabbit anti-goat IgG antibody (cat. no. E030330; 1:5,000; EarthOx Life Sciences, Millbrae, CA, USA). PBST was used to wash the membranes 4 times for 10 min each. The protein was then visualized using a Developer and fixer kit (cat. no. P0019, Beyotime Institute of Biotechnology), and the development time was 1-2 min.

**Ectopic bone formation.** Polyglycolic acid scaffold materials (Dexon, Shanghai, China) were sterilized using 75% alcohol and then cut into 0.5 $\times$ 0.5 $\times$ 0.5  $\text{cm}^2$  pieces. The PGA scaffolds were soaked in DMEM for 2 h, then 2 ml BMSCs ( $5.0 \times 10^7$  cells/ml) were seeded into the scaffolds as the negative control group. The following treatment groups were seeded onto the scaffolds: i) BMSCs + PGA + CPB50; ii) BMSCs + PGA + CPB100; and iii) BMSCs + PGA + CPB200. The PGA scaffolds were incubated for 5 days to ensure that cells and carriers had adhered successfully. The scaffolds were then implanted into the dorsal muscles of rats in order to determine the ectopic bone formation. Three scaffolds were implanted per treatment group. After 2 months, the 4 rats were euthanized by cervical dislocation and the regions with the implanted scaffolds were dissected. The tissues were then subjected to hematoxylin and eosin staining.

**Statistical analysis.** Data are presented as the mean  $\pm$  standard error. One-way analysis of variance was performed using SPSS version 17.0 software (SPSS, Inc., Chicago, IL, USA).  $P < 0.05$  was considered to indicate statistically significant difference.

## Results

**Transfection efficiency of CNPBs.** Following 24 h transfection of BMSCs no green fluorescence was observed in the blank control BMSC (untreated) group. However, green fluorescence was observed in the pBMP2 (positive control) and CNPB at different concentrations groups. This indicated that chitosan nanoparticles delivered the BMP2 gene into BMSCs and successfully displayed green fluorescence. The fluorescence of the pBMP2 and CPB50 groups was weaker compared with the CPB100 and CPB200 groups and particle analysis revealed that there was a greater quantity of fluorescence particles and in greater total areas in the CPB100 and CPB200 groups (Table II, and Figs. 1 and 2). Statistical analysis revealed that there was a significant difference in particle quantity among the groups ( $P < 0.0001$ ). Pairwise comparisons determined that there was a significant difference in particle quantity between any two groups, with the exception of the CPB50 and pBMP2 groups ( $P=0.243$ ). There was also a significant

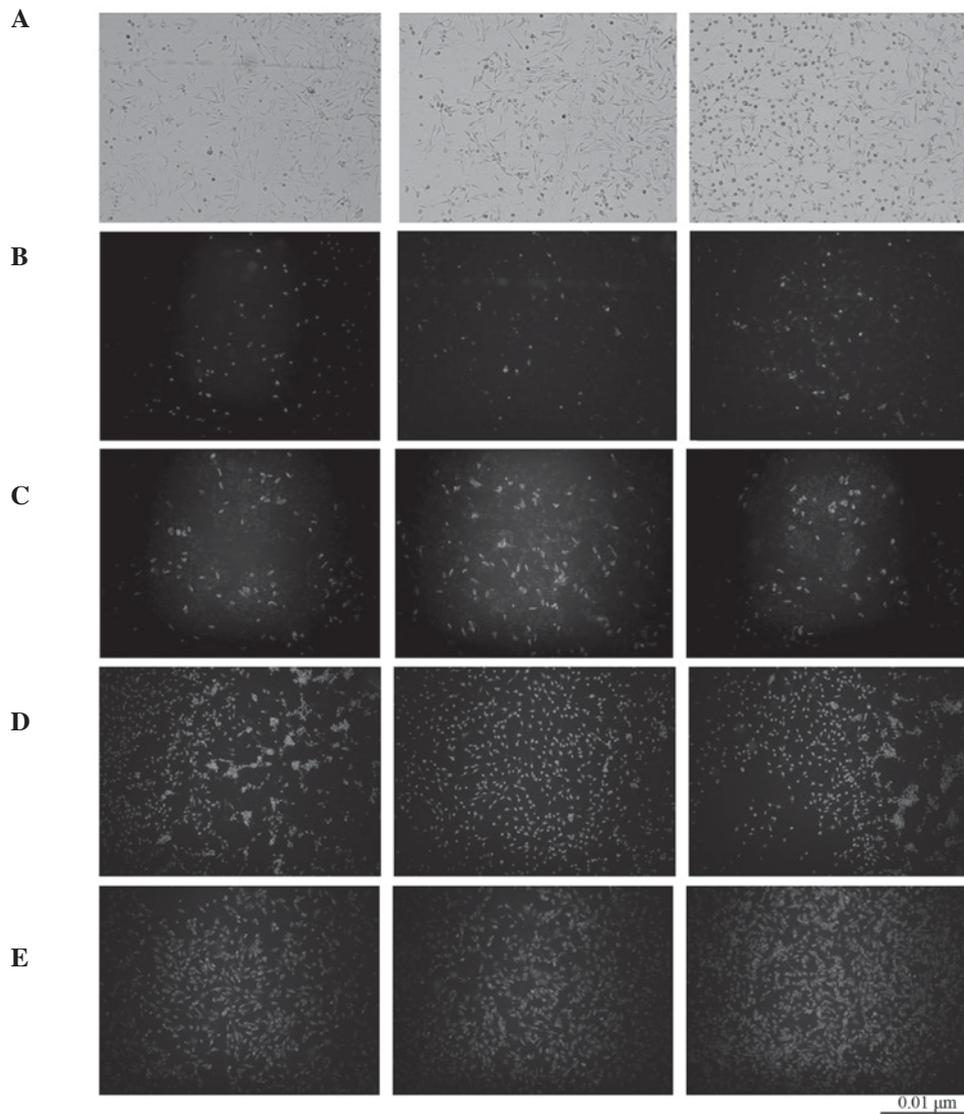


Figure 1. BMSCs with CNPBs transfection. (A) Blank control, (B) positive control, (C) CPB50, (D) CPB100 and (E) CPB200 groups. No fluorescence particles were observed in the untreated BMSC (blank control) group. A small number of fluorescence particles were observed in the positive control group (BMSCs exposed to pBMP2 only). There were many fluorescence particles observed in the CPB100 and CPB200 groups. BMSCs, bone marrow stem cells; pBMP2, plasmid bone morphogenetic protein 2; CPB50/100/200, chitosan nanoparticles containing plasmid-bone morphogenetic protein 2 at concentrations of 50, 100 and 200  $\mu\text{g}/\text{ml}$ .

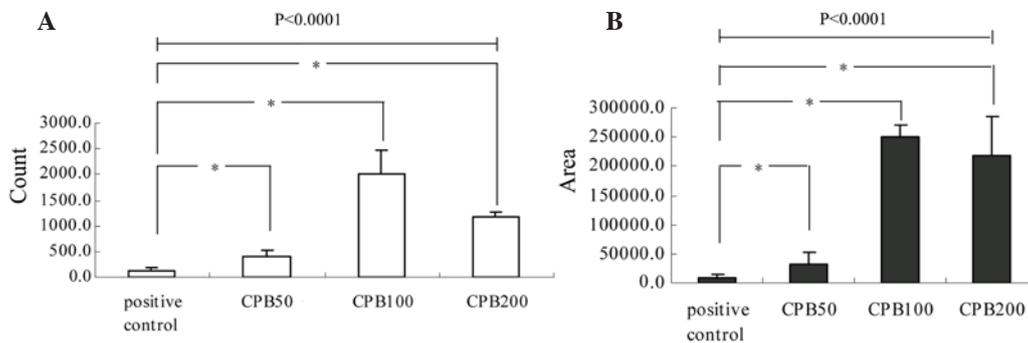


Figure 2. Histograms of the (A) quantity and (B) total area of fluorescent particles in bone marrow stem cells treatment groups. A larger quantity and larger total area of particles were observed in the CPB100 and CPB200 groups. CPB50/100/200, chitosan nanoparticles containing plasmid-bone morphogenetic protein 2 at concentrations of 50, 100 and 200  $\mu\text{g}/\text{ml}$ .  $P < 0.00001$ .

difference in total areas with fluorescence particles among the groups ( $P < 0.0001$ ). The difference in total areas between any

two groups was significant, except between the CPB100 and CPB200 treatment groups ( $P = 0.321$ ; Table II, Figs. 1 and 2).

Table III. Expression levels of ALP, OPG, OC, and OPN in osteoblast-like cells differentiated from BMSCs transfected with CNPBs.

Groups Objective genes	mRNA expression		Protein expression	
	ALP	OPG	OC	OPN
Blank control	148.8±4.5	238.1±7.1	116.0±4.3	24.2±2.2
CPB50	314.0±9.5	372.7±9.8	147.7±4.3	53.3±2.4
CPB100	467.0±6.3	529.3±2.5	262.2±5.4	88.7±1.8
CPB200	485.2±15.5	666.0±3.9	292.8±6.6	117.6±5.2
Negative control	151.3±6.2	186.2±6.3	88.7±2.1	16.6±0.7

Statistical analysis determined that there was a significant difference between any two groups ( $P < 0.0001$ ), except between the blank control and negative control groups ( $P = 0.751$ ). ALP, alkaline phosphatase; OPG, osteoprotegerin; OC, osteocalcin; OPN, osteopontin; BMSCs, bone marrow stem cells; CNPBs, chitosan nanoparticles containing plasmid-bone morphogenetic protein 2 sequences; CPB50/100/200, chitosan nanoparticles containing plasmid-bone morphogenetic protein 2 at concentrations of 50, 100 and 200  $\mu\text{g/ml}$ .

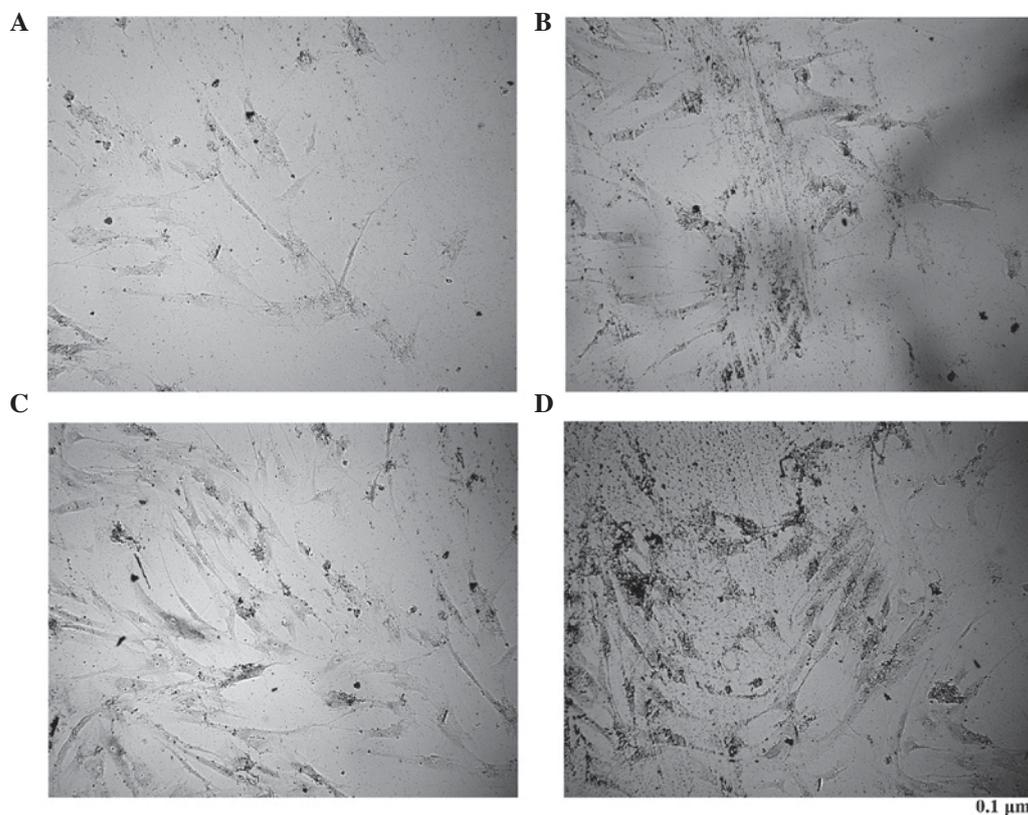


Figure 3. Alkaline phosphatase staining of osteoblast-like cells. (A) Control; (B) CPB50; (C) CPB100; and (D) CPB200. No brownish-black particles were observed in the control group. The number of brownish-black particles increased in the CPB200 group. CPB50/100/200, chitosan nanoparticles containing plasmid-bone morphogenetic protein 2 at concentrations of 50, 100 and 200  $\mu\text{g/ml}$ .

*Alkaline phosphatase staining of osteoblast-like cells differentiated from transfected BMSCs with CNPBs.* There were no obvious brownish-black particles observed in the control group; however, there were numerous brownish-black particles in the CPB50, CPB100 and CPB200 groups. In addition, more particles were observed in the CPB200 group compared with the CPB50 and CPB100 groups (Fig. 3).

*Wright's staining of osteoblast-like cells differentiated from BMSCs transfected with CNPBs.* The staining revealed that

BMSCs of a smaller size were fibriform in the control group (Fig. 4A). However, osteoblast-like cells that differentiated from BMSCs were observed to be larger in size, with inflated cell bodies and nucleoli stained dark blue in the CPB50, CPB100 and CPB200 treatment groups. In addition, osteoblast-like cells also had characteristic stick-like prominences (Fig. 4).

*Von Kossa staining of osteoblast-like cells differentiated from BMSCs transfected with CNPBs.* No staining was observed in the control group (Fig. 5A). Black particles, indicating

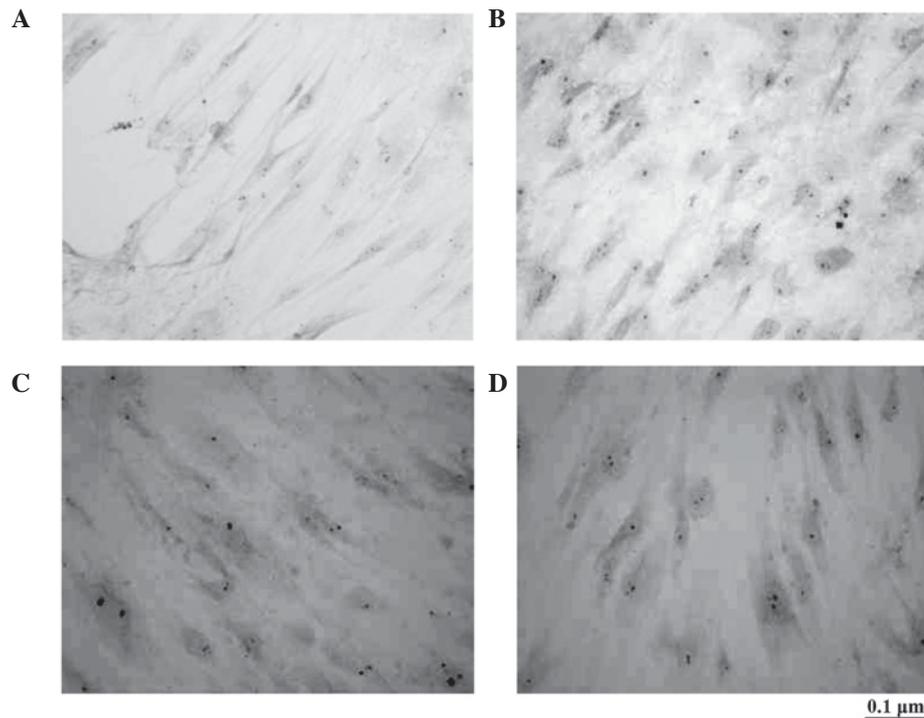


Figure 4. Wright's staining of BMSCs transfected with CNPBs of (A) control, (B) CPB50, (C) CPB100 and (D) CPB200 groups. Differences among the treatment groups were identified. Osteoblast-like cells with characteristic stick-like prominences differentiated from BMSCs were larger, with inflated cell bodies and their nucleoli had dark blue staining in the CPB200 group. BMSCs, bone marrow stem cells; pBMP2, plasmid bone morphogenetic protein 2; CNPBs, chitosan nanoparticles containing plasmid-bone morphogenetic protein 2 sequences; CPB50/100/200, chitosan nanoparticles containing plasmid-bone morphogenetic protein 2 at concentrations of 50, 100 and 200  $\mu\text{g/ml}$ .

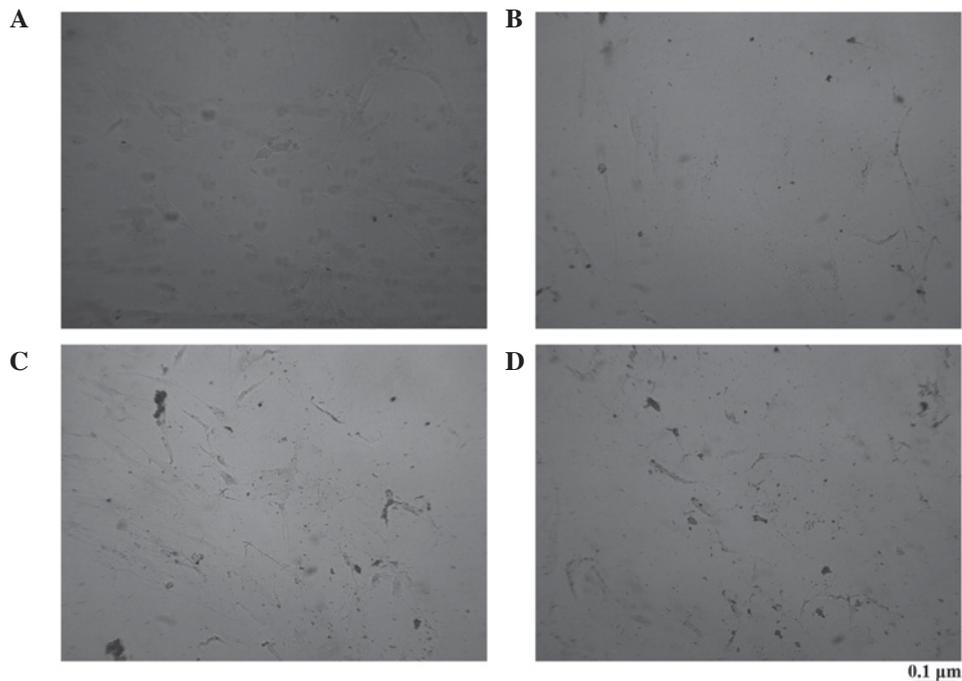


Figure 5. Von Kossa staining of BMSCs transfected with CNPBs. (A) Control; (B) CPB50; (C) CPB100; and (D) CPB200. Black nodes observed in the CNPB groups; however, they were not observed in the control group. BMSCs, bone marrow stem cells; pBMP2, plasmid bone morphogenetic protein 2; CNPBs, chitosan nanoparticles containing plasmid-bone morphogenetic protein 2 sequences; CPB50/100/200, chitosan nanoparticles containing plasmid-bone morphogenetic protein 2 at concentrations of 50, 100 and 200  $\mu\text{g/ml}$ .

a positive reaction of calcium phosphate in the mineralized extracellular matrix of osteoblasts, were observed in the CPB50, CPB100 and CPB200 treatment groups (Fig. 5B-D).

*Expression levels of ALP, OPG, OC and OPN in osteoblast-like cells differentiated from BMSCs transfected with CNPBs.* The RT-PCR determined that the mRNA expression levels of

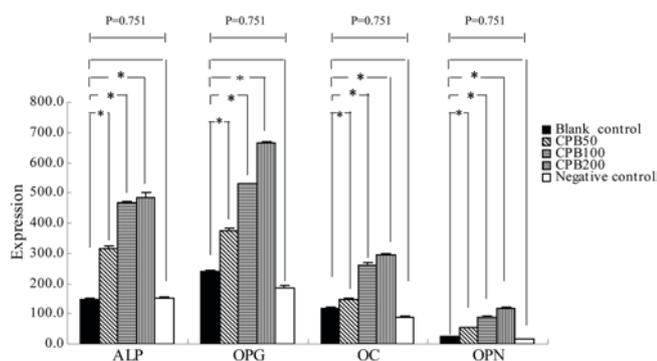


Figure 6. Expression levels of ALP, OPG, OC and OPN. The highest mRNA expression levels were found in the CPB200 group. Significantly lower expressions of genes were observed in the control groups. ALP, alkaline phosphatase; OPG, osteoprotegerin; OC, osteocalcin; OPN, osteopontin; CNPBs, chitosan nanoparticles containing plasmid-bone morphogenetic protein 2 sequences; CPB50/100/200, chitosan nanoparticles containing plasmid-bone morphogenetic protein 2 at concentrations of 50, 100 and 200  $\mu\text{g/ml}$ . \* $P < 0.0001$ .

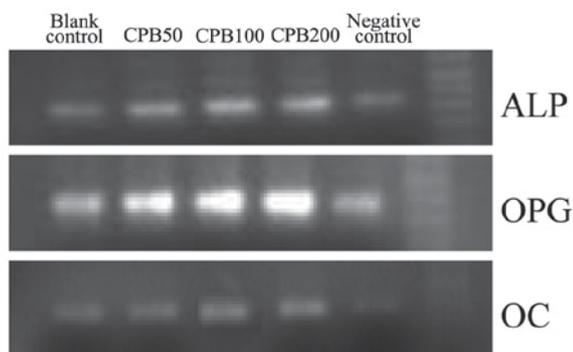


Figure 7. Gel electrophoresis of ALP, OPG, and OC. mRNA expression levels of ALP, OPG, and OC were evident in the CPB200 group. ALP, alkaline phosphatase; OPG, osteoprotegerin; OC, osteocalcin; CPB50/100/200, chitosan nanoparticles containing plasmid-bone morphogenetic protein 2 at concentrations of 50, 100 and 200  $\mu\text{g/ml}$ .

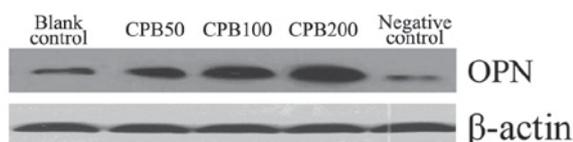


Figure 8. Western blotting was performed in order to determine the protein expression levels of OPN in the various treatment groups. The CPB200 group had the highest expression levels. OPN, osteopontin; CPB50/100/200, chitosan nanoparticles containing plasmid-bone morphogenetic protein 2 at concentrations of 50, 100 and 200  $\mu\text{g/ml}$ .

ALP, OPG, OC and OPN were increased in the CNPB treatment groups compared with the blank and negative control groups (Fig. 6). The highest mRNA expression levels of ALP, OPG, OC and OPN were identified in the CPB200 treatment group (Table III; Figs. 6-8). Statistical analysis revealed that there was a significant difference between any two groups ( $P < 0.0001$ ), except between the blank and negative control groups ( $P = 0.751$ ).

**Ectopic bone formation.** Two months after the PGA scaffolds that were integrated with BMSCs transfected with CNPBs

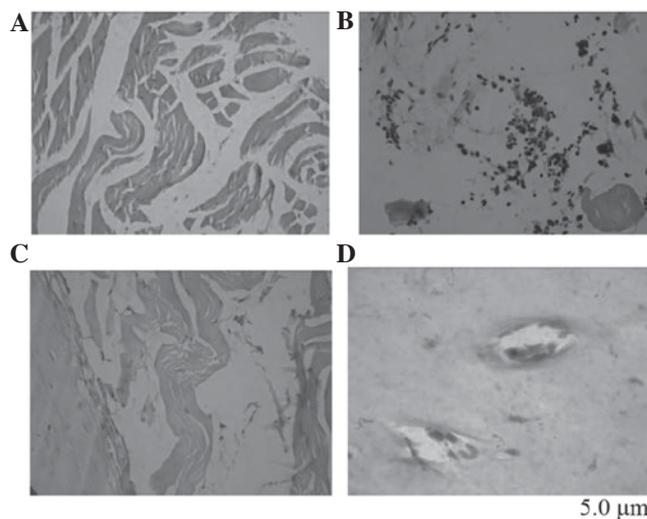


Figure 9. Histological staining of ectopic bone formation test. (A) Red staining of muscle fibers was evident in the control group. (B) CPB200 group had aggregating lymphocytes in the implanted regions. (C) A classic bone tissue image in the CPB200 group. (D) Bone lacunae were evident in the CPB200 group. CPB200, chitosan nanoparticles containing plasmid-bone morphogenetic protein 2 at concentration of 200  $\mu\text{g/ml}$ .

were implanted into the muscles of rats, by touch, the sensation of a 'string-like' object in the subdermal implanted regions of the control and CPB50 groups was identified. However, a node-like object was identified in the subdermal implanted regions of the CPB100 and CPB200 groups. After the implanted regions were excised, numerous muscle fibers were observed in the control and CPB50 groups. In the CPB100 and CPB200 groups, a dark-red node with white spots was identified, which was surrounded by fibrous tissues. Histological staining revealed that there was novel bone formation had occurred in the CPB100 and CPB200 treatment groups (Fig. 9). This bone formation was more evident in the CPB200 group. In the positions of white spots were determined to be cartilage islands (Fig. 9). The PGA scaffolds were absorbed in all treatment groups.

## Discussion

Chitosan is considered to be an effective non-viral carrier and is extensively used for tissue engineering (13,16). In our previous study (15), chitosan nanoparticles had excellent biocompatibility. On the basis of these results, the present study used chitosan nanoparticles as carriers of genetic material. CNPBs had excellent transfection efficiency to BMSCs and induced the differentiation and proliferation of osteoblast-like cells. In addition, CNPBs induced ectopic formation of new bone in rats. Therefore, the present study determined that CNPBs are promising carriers of genetic material.

BMSCs have been used extensively in bone tissue engineering. As BMSCs are capable of multi-directional differentiation, bone growth factors and osteogenesis are important factors to consider for successful bone tissue engineering. Bone growth factors primarily include the BMP family, transforming growth factor- $\beta$ , dexamethasone, the active form of vitamin D [1,25-(OH) 2D3], vitamin C and sodium glycerophosphate. The exogenous BMP family is a group of acid

polypeptides with low molecular weights. BMP2 may promote cell proliferation and induce seed cells to differentiate into osteoblast-like cells, which may result in the formation of new bone. Hou *et al* (16) determined that BMP2 had potent osteo-inductive properties in bone regeneration (16). The composite scaffold that Hou *et al* (16) constructed of recombinant human BMP2 (rhBMP2)-loaded collagen/chitosan microspheres bridged bone defects and recanalized the bone-marrow cavity. The results of a previous study indicated that BMP2 alone had a positive effect on bone regeneration (17). Yilgor *et al* (18) revealed that treatment with BMP2 alone resulted in a higher ALP activity compared with treatment with BMP7 (18).

In the present study, BMSCs were transfected with CNPBs. BMP2 was released continuously during osteogenesis to promote formation of new bone. Therefore, the present study determined that BMSCs transfected with CNPBs at a higher efficiency. The phenotype of osteoblast-like cells was confirmed using alkaline phosphatase, Wright's, von Kossa staining. In addition, the mRNA and protein expression levels of ALP, OPG, OC and OPN in osteoblast-like cells differentiated from the transfected BMSCs were recorded at higher levels compared with the control group, which had untreated cells. In order to determine the extent of ectopic bone formation, CNPBs were attached to PGA scaffolds to induce new bone formation following implantation into the dorsal muscles of rats.

Chitosan gels loaded with BMP2 enhanced ALP activity in BMSCs by 3.6-fold, and increased the calcium mineral deposition of mesenchymal cells by 2.8-fold (19). In addition, chitosan gels loaded with BMP2 induced synthesis of OC in BMSCs (19). The present study was consistent with the findings of previous studies (20-22). Zhao *et al* (20) determined that a calcium phosphate-chitosan fibrous scaffold delivery system with BMP2 promoted osteo-differentiation and resulted in increased gene expression levels of ALP and OC (20). Shi *et al* (21) demonstrated that carboxymethyl chitosan-BMP2 modified substrates significantly promoted ALP activity and calcium mineral deposition of osteoblasts and human bone marrow-derived mesenchymal stem cells (21). Additionally, bone formation was observed in the quadriceps muscles of rats following the implantation of rhBMP2-loaded chitosan carriers (22).

The BMP2-induced differentiation of BMSCs into osteoblast-like cells ensures that BMP2 is able to recruit BMSCs and promote their proliferation. During bone formation, BMP2 promotes cellular differentiation and proliferation through autocrine and paracrine secretion. Following BMP2 binding to the BMP2-specific receptor on the cell surface, Smad protein becomes activated through signal transmission. Subsequently, Smad protein is transported into the cell nuclei. Next, the nuclear factor, runt-related transcription factor 2 (Runx2; also known as core-binding factor subunit- $\alpha$ 1), is activated through the mitogen-activated protein kinase pathway. Following the combination of Runx2 with Smad, specific phenotypes in osteoblast-like cells are activated sequentially (16-22). Finally, expression levels of ALP, OPG, OC and OPN were upregulated, and BMSCs were differentiated into osteoblast-like cells.

Scaffold materials may primarily include natural and synthesis polymer and bioceramics (23). Natural polymers such as collagen, chitosan-alginate gel and hyaluronic acid

may be used. Synthesized polymers that are biodegradable include polyactide, polyviol, polyacrylic acid and polyethylene glycol. The primary bioceramics used are calcium phosphate ceramics, hydroxyapatite and calcium carbonate (24,25). Chen *et al* (26) used bilayered integrated scaffolds in their study. Their findings revealed that mesenchymal stem cells seeded in each layer of the bilayered gene-activated osteochondral scaffold had a higher expression of BMP2 protein (26). Park *et al* (27) determined that chitosan-alginate gel/mesenchymal stem cell/BMP2 composites were able to stimulate novel bone formation. The primary aim of tissue engineering is to construct a three-dimensional complex composed of seed cells and biomaterials. Therefore, PGA scaffolds with the same excellent biocompatibility and biodegradability have been used extensively in bone tissue engineering. The present study involved the transfection of BMSCs with CNPBs, and they were subsequently incubated with PGA scaffolds. Cells adhered to, and proliferated on, the PGA scaffolds. The histological staining revealed that the PGA scaffolds had successfully degraded, and novel bone formation was observed. The present study determined that PGA scaffolds with three-dimensional structures provided a larger surface area, and simulated the formation of natural bone structures. This allowed for the slow release of BMP2 in the CNPBs, providing an excellent 'shelter' for osteoblast-like cells differentiated from BMSCs. This promoted the osteogenic functions of osteoblast-like cells.

In conclusion, in the present study CNPBs were successfully transfected into BMSCs with a high efficiency, and BMSCs were promoted to differentiate into osteoblast-like cells *in vitro*. Additionally, CNPBs upregulated the expression levels of ALP, OPG, OC and OPN in osteoblast-like cells and induced the formation of new ectopic bone *in vivo*.

## Acknowledgements

The present study was supported by the Department of Education Foundation of Zhejiang Province of China (grant nos. 20080180, Y201018976 and N20110143), Zhejiang Provincial Natural Science Foundation of China (grant nos. Y2080340 and LY12H14004) and Zhejiang Provincial and Ministry Joint Project (grant no. WKJ2011-2-009).

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