

Calyptanthes grandifolia O.Berg (Myrtaceae) ethanolic extract inhibits TNF- α gene expression and cytokine release *in vitro*

GEÓRGIA MUCCILLO DEXHEIMER¹, LUCIANA KNABEN DE OLIVEIRA BECKER DELVING²,
HENRIQUE SULZBACH DE OLIVEIRA¹, VANDERLEI BIOLCHI²,
MÁRCIA INÊS GOETTERT¹ and ADRIANE POZZOBON²

¹Biotechnology Program; ²Molecular Biology Laboratory and Medical School,
Univates, Lajeado, RS 95900-000, Brazil

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Abstract. Anti-tumor therapies based on anti-inflammatory effects have been considered in cancer treatment. Survival, proliferation and, resultantly, invasion and metastasis of tumor cells are regulated by local inflammatory mediators. Primary inflammatory cytokines, such as tumor necrosis factor (TNF), are targets for anticancer therapy. Several anti-inflammatory agents isolated from natural products are becoming important chemopreventive and therapeutic agents for cancer. The present study aimed to investigate the expression of TNF- α , nuclear factor- κ B (NF- κ B) and p38 α mitogen-activated protein kinase (p38 α) genes, associated with proliferation and inflammation in the Caco-2 cell line treated with ethanolic and hexanic extracts of *Calyptanthes grandifolia* O.Berg (Myrtaceae). Caco-2 cells were cultured and treated with plant extract at different concentrations (25, 50, 100 and 200 μ g/ml) and stimulated with lipopolysaccharide (LPS). For gene expression, analysis was performed by total RNA extraction followed by synthesis of complementary DNA and analysis by quantitative polymerase chain reaction. The release of TNF- α cytokine was evaluated by ELISA in RAW 264.7 murine macrophages activated by LPS. Among the evaluated genes, there was a decrease in TNF- α expression at 100 and 200 μ g/ml concentrations only with the ethanolic extract ($P < 0.025$). The p38 α gene exhibited a tendency to increase expression only when treated with ethanolic extract and the NF- κ B gene did not significantly differ compared with the positive control when treated with either analyzed extract. The inhibition of TNF- α cytokine in the RAW 264.7 cell line was significant ($P < 0.05$) in ethanolic extract at 200 μ g/ml compared with the positive control (LPS 1 μ g/ml). In conclusion, the ethanolic

extract may exhibit an anti-inflammatory activity by inhibiting TNF- α . However, further studies are required to confirm its potential anti-inflammatory effects.

Introduction

Inflammation associated with cancer is a promising target for the development of anticancer therapies. Cytokines, chemokines and growth factors may have an important function in the interaction between tumor cells and infiltrating leukocytes from blood vessels. The existence of inflammatory components in the microenvironment of neoplastic tissues frequently leads to increased angiogenesis, resistance to hormones and inhibition of adaptive anti-tumor immunity. The survival, proliferation and subsequent invasion and metastasis of tumor cells is regulated by inflammatory mediators present at the tumor site (1).

Primary inflammatory cytokines, including interleukin-1 (IL-1) and tumor necrosis factor (TNF), expressed by leukocytes and tumor infiltrating cells, are targets for anticancer therapy. Anti-cytokine strategies against tumors have been investigated with TNF inhibitors in certain inflammatory diseases (2). Clinical trials with TNF- α antagonists, alone or combined with other therapies, have been performed in patients with cancer. In cases of advanced solid cancer, TNF- α antagonists were well tolerated and exhibited biological activity and partial response with renal carcinoma or stable disease (3,4). Permanent activation of NF- κ B contributes actively in tumorigenesis by promoting cell cycle progression and inhibiting apoptosis. NF- κ B is activated through TNF- α , thus inhibition of this factor may support cancer therapies that target apoptosis (5,6). Inhibition of NF- κ B is associated with apoptosis and reduced cell growth, and this inhibition may be beneficial in the treatment of cancer (7). The p38 α mitogen-activated protein kinase (p38 α MAPK) may have tumor suppressor activity through regulation of the p53 gene, which interferes with cell cycle progression and induces apoptosis. However, it also exhibits oncogenic activity associated with various processes, including invasion, inflammation and angiogenesis, which are essential in tumor development (8).

Plants are a major source of active substances that are used in therapeutic medicine as their metabolites have great

Correspondence to: Ms Geórgia Muccillo Dexheimer, Biotechnology Program, Univates, 171 Avenida Avelino Tallini, Lajeado, RS 95900-000, Brazil
E-mail: georgiamuccillo@gmail.com

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structural diversity (9). Anti-inflammatory compounds have been extracted from natural products, including fruits, vegetables, spices and traditional medicinal herbs, and these compounds have been gaining importance as potential chemopreventive or therapeutic agents for cancer (10). In the last 2 decades, herbal products have been important candidates in the discovery of novel drugs for cancer (11). Drugs derived from natural products that have antibacterial, anticoagulant, antiparasitic, immunosuppressive and anticancer properties are able to treat 87% of categorized human diseases (12). There were 12 natural products and 32 derivatives of natural products among the 128 anticancer drugs released to the market between 1981 and 2010. They were obtained from various sources, including plants and microorganisms. Between 1940 and 2010, 175 small molecules were released for the treatment of cancer and of those, 131 molecules were developed from natural products (13). The investigation of novel plants with anti-inflammatory, anti-tumor and anti-carcinogenic potential is important and may enable the development of novel drugs for cancer treatment. Several members of Myrtaceae family have been previously investigated and various activities were observed, including antioxidant (14), anti-tumor (15-21) and anti-inflammatory activities (22-27). To the best of our knowledge, there are no existing studies investigating the activity of *Calyptanthes grandifolia*. However, important activities have been demonstrated in other members of the *Calyptanthes* genus (28-35). Thus, this genus demonstrates potential and may be beneficial in the treatment of inflammatory and tumor processes. The objective of the present study was to investigate the expression of genes associated with proliferation and inflammation in cells of the Caco-2 cell line treated with extracts from *Calyptanthes grandifolia* O.Berg.

Materials and methods

Plant material. Leaves of *Calyptanthes grandifolia* O.Berg were collected in Lajeado, Rio Grande do Sul, in Southern Brazil and identified by Professor Elisete Maria de Freitas. From this material ethanol and hexane extracts were isolated according to the following methodology.

Hexanic extract preparation. The leaves of the plant were dried in an incubator with circulating air at 38°C for 24 h. Subsequently, leaves were reduced to small fragments to increase the contact surface with the extraction solution. The leaves of *Calyptanthes grandifolia* O.Berg were packed with hexane solvent in an amber bottle at room temperature for 72 h. Vacuum filtration was performed and the filtrate was stored in an amber jar at room temperature until the solvent was removed. Maceration of the leaves occurred for 2 weeks and the solvent was changed twice. Subsequently, vacuum filtration and removal of solvent was performed with the aid of a rotary evaporator at 40°C. Finally, the extract obtained was stored in amber bottles and refrigerated at 4±1°C until experiments were performed.

Ethanolic extract preparation. Plant leaves were dried in an oven with circulating air at 38°C for 24 h. Following this period, cold static soaking was performed on fragments of leaves with 90% ethanol and the material was placed in an amber bottle

and kept at room temperature for 7 days. Subsequent to the extraction period, vacuum filtration and removal of solvent was performed with the aid of a rotary evaporator at 40°C. The extract was placed in amber bottles and refrigerated at 4±1°C until experiments were performed.

Dilution of extracts. Solubilization of the extract was performed with dimethylsulfoxide (DMSO) so that the final concentration was ≤0.5%.

Cell culture. Caco-2 colorectal adenocarcinoma cell line (HTB-37; American Type Culture Collection, Manassas, VA, USA) and RAW 264.7 murine macrophage cell line (TIB-71; American Type Culture Collection) were cultivated in microwell plates (1x10⁵ cells/well) in an incubator (37°C, 5% CO₂). Subsequently, treatment with plant extract was performed at different concentrations (25, 50, 100 and 200 µg/ml), with incubation in culture medium Dulbecco's modified Eagle's medium (DMEM, Sigma-Aldrich; Merck KGaA, Darmstadt, Germany) supplemented with 10% fetal bovine serum (FBS, Sigma-Aldrich; Merck KGaA) for 1 h at 37°C. Subsequently, lipopolysaccharides (LPS; 1 µg/ml) were added, and incubation was performed at 37°C and 5% CO₂ for 24 h (36). Subsequent to 24 h incubation at different concentrations and treatments, extraction of total RNA was performed on the Caco-2 cell line and the supernatant of RAW 264.7 cells was collected for ELISA assay. In addition to treatment with extracts, each culture plate had a positive control, in which cells were stimulated with LPS only, and a negative control with no stimulation or treatment. A total of 5 different experiments were performed for gene expression analysis.

Cell viability assay by alamar Blue®. Caco-2 cells were plated at density of 1x10⁵ cells/ml in 96-well plates containing 200 µl of DMEM low glucose and 10% FBS, and incubated for 24 h in an atmosphere of 5% CO₂, 90% humidity and 37°C. Subsequently, cells were treated with concentrations of 25, 50, 100 or 200 µg/ml per well of extract, and incubated for 72 h. Following this period, treatment was removed and a solution of 10% alamar Blue® dye was added per well. The absorbance readings were performed following 6 h incubation at 37°C, at a wavelength of 540 nm (oxidized) and 620 nm (reduced) in an ELISA reader. As a negative control, cells were placed only in culture medium and DMSO. The percentage of cell viability was calculated using the following formula: % alamar Blue® reduction = absorbance at 540 nm - (absorbance at 630 nm x correction factor) x 100. Correction factor was calculated by staining the culture medium with no cells.

Cell viability assay by Trypan Blue. Cells were removed from the plates with the aid of a scraper and transferred to 15 ml centrifuge tubes (1x10⁵ cells/centrifuge tube; Corning Incorporated, Corning, NY, USA), which were centrifuged at 600 x g for 10 min at room temperature. Following centrifugation, the supernatant was discarded and the pellet was resuspended in 1 ml DMEM in a 1:10 dilution with Trypan Blue dye, which stains non-viable cells. Finally, a total count of viable cells was performed in a Neubauer chamber for subsequent plating of viable Caco-2 and RAW264.7 cells (37).

RNA extraction. Total RNA extraction was performed by the TRIzol[®] method (Invitrogen; Thermo Fisher Scientific, Inc.) according to the manufacturer's instructions and purification was performed by illustra RNAspin Mini kit (GE Healthcare Life Sciences, Chalfont, UK). Total RNA was quantified using aL-Quant[®] spectrophotometer (Loccus, São Paulo, Brazil) with a 2 μ l final product of RNA extraction, and absorbance was read at 260 nm.

Complementary DNA (cDNA) synthesis. Synthesis of cDNA was performed from 0.5 μ g total RNA, using poly-A tail complementary oligonucleotide primers and the Superscript[™] II Reverse Transcriptase kit according to the manufacturer's protocol (Invitrogen; Thermo Fisher Scientific, Inc.). To each sample tube, 1 μ l of dNTP mix and 1 μ l OligodT was added prior to incubation for 5 min at 65°C. Then, 9 μ l of a mix containing 10X PCR Buffer, 25 mM MgCl₂, 0.1 M DTT and RNase OUT was added and further incubated for 2 min at 42°C. A total of 1 μ l of Superscript II RT was added and incubated again at 42°C for 50 min and 70°C for 15 min. Finally, 1 μ l of RNase H was added and incubated at 37°C for 20 min. At the end of synthesis, the cDNA was stored at -20°C until amplification was performed by quantitative polymerase chain reaction (qPCR).

qPCR. Gene expression analysis was performed by qPCR. The results were normalized to β -actin (38,39) and the efficiency of reactions was evaluated using the standard curve of each gene analyzed. The qPCR results were expressed as the relative quantification of amplified cDNA with respect to the normalizer gene (40). Primers used for amplification of specific cDNA fragments were selected from the published sequence of each gene using online tool Primer3 v.0.4.0 (41). All primers were synthesized by Invitrogen (Thermo Fisher Scientific, Inc.; Table I). DNA amplification and relative quantification was performed using a StepOnePlus[™] Real-Time PCR system (Applied Biosystems; Thermo Fisher Scientific, Inc.) and the Platinum[®] SYBR[®] Green qPCR SuperMix-UDG kit (Invitrogen; Thermo Fisher Scientific, Inc.) in a total volume of 25 ml [12.5 μ l SuperMix, 0.5 μ l (50 μ mol/l) Rox reference dye, 0.3 μ l of each primer (10 μ mol/l forward and 10 μ mol/l reverse), 9.4 μ l H₂O and 2.0 μ l 1:20 diluted template cDNA] according to the manufacturer's (SYBR[®] Green kit) instructions. Amplification and reading of samples was performed in duplicate with the following protocol for all genes: Initial incubation for 3 min at 94°C; followed by 45 cycles of 30 sec denaturation at 94°C; 30 sec annealing at 55°C and 30 sec extension at 60°C. To confirm specificity of the reaction, a dissociation curve was performed for each primer pair with melting temperature analysis of each gene.

TNF- α cytokine release in RAW 264.7 cells by ELISA assay. Following 24 h incubation of RAW 264.7 cells with LPS stimulation, 0.4 ml supernatant was collected and stored at -80°C and used for quantification of the release of TNF- α pro-inflammatory cytokine using the Mouse TNF alpha ELISA Ready-SET-Go[®] ELISA kit (cat. no. 88-7324-86; e-Bioscience, Inc., San Diego, CA, USA) according to the manufacturer's instructions.

Table I. Oligonucleotide primers for quantitative polymerase chain reaction.

Gene	Sequence (5'-3')	Fragment size (bp)
NF- κ B		209
Sense	ACACCGTGTAACCAAAGCC	
Antisense	CAGCCAGTGTGTGATTGCT	
p38 α		243
Sense	CAGTGGGATGCATAATGGCC	
Antisense	GCATCTTCTCCAGCAAGTCG	
TNF- α		120
Sense	CCCTGGTATGAGCCCATCTATC	
Antisense	AAAGTAGACCTGCCAGACTCG	
β -actin		140
Sense	CTGGAACGGTGAAGGTGACA	
Antisense	AAGGGACTTCCTGTAACAATGCA	

NF- κ B, nuclear factor κ B; p38 α , p38 α mitogen-activated protein kinase; TNF- α , tumor necrosis factor- α .

Statistical analysis. Data were tabulated and analyzed with descriptive statistics using SPSS software (version 20.0; IBM SPSS, Armonk, NY, USA) and GraphPad Prism (version 5.0; GraphPad Software, Inc., La Jolla, CA, USA). Comparison between the controls was performed using an unpaired Student's *t*-test and the effects of extracts were analyzed by one-way analysis of variance followed by the Tukey test. *P*<0.05 was considered to indicate a statistically significant difference.

Results

Cell viability by alamar Blue[®]. It was observed that the ethanolic and hexane extracts of *Calypttranthes grandifolia* O.Berg did not affect the cell proliferation when compared with the negative control. The negative control exhibited a viability of 62.6% (\pm 4.367), and for ethanolic and hexane extracts at tested concentrations, the percentages of viability were similar to the control (Fig. 1).

Gene expression analysis. For the analysis of the genes of interest, the positive control consisted of Caco-2 cells that were stimulated only by LPS with no extract treatment, and the negative control consisted only of cells in culture. The comparison between negative and positive controls was performed in order to verify whether there was a difference in gene expression when stimulated by LPS. Increased expression of TNF- α was observed following treatment with LPS compared with the negative control, however, this was not significantly different. In addition, the expression of NF- κ B and p38 α were similar in negative and positive control groups; expression was not altered by LPS treatment. Thus, the expression of these genes was stable in the negative and positive control groups (Fig. 2). When evaluating the effect of the ethanolic extract of *Calypttranthes grandifolia* on gene expression, a decrease

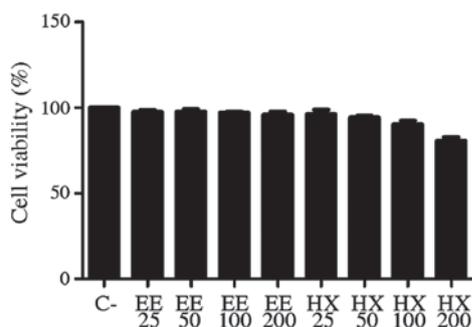


Figure 1. Percentage cell viability by alamar Blue[®] method. Results were analyzed by one-way analysis of variance followed by Tukey test ($P=0.775$; $n=3$). C-, negative control; EE, ethanolic extract; HX, hexanic extract.

in TNF- α expression was observed, which was significant at concentrations of 100 and 200 $\mu\text{g/ml}$, compared with the positive control ($P<0.025$; Fig. 3A). Regarding NF- κB gene expression, expression was similar for all treatments and there were no significant changes (Fig. 3B). The expression of p38 α increased with increasing concentrations of ethanolic extract, however, these increases were not significant (Fig. 3C). The expression of TNF- α , NF- κB and p38 α with hexane extract treatment exhibited no significant variation compared with the positive control (Fig. 3D-F).

TNF- α cytokine release in RAW 264.7 cells by ELISA.

In order to demonstrate the decrease in TNF- α in gene expression, an ELISA colorimetric assay was performed to evaluate the level of cytokine release. A macrophage lineage was used, characterized by the release of inflammatory cytokines. The inhibition of TNF- α was significant ($P<0.05$) with the ethanolic extract at a concentration of 200 $\mu\text{g/ml}$ (464.0 ± 36.7 pg/ml) compared with the positive control (1143.0 ± 118.0 pg/ml; Fig. 4) and also when compared with a concentration of 25 $\mu\text{g/ml}$ (1272 ± 293 pg/ml). Concentrations of 25 $\mu\text{g/ml}$ (1272 ± 293 pg/ml), 50 $\mu\text{g/ml}$ (1179 ± 261 pg/ml) and 100 $\mu\text{g/ml}$ (1185 ± 312 pg/ml) exhibited no significant difference compared with the positive control (Fig. 4). The negative control (53.6 ± 36.8 pg/ml) exhibited a significant difference in TNF- α cytokine release compared with the positive control ($P<0.0001$; Fig. 4) and compared with concentrations of 25, 50 and 100 $\mu\text{g/ml}$ ($P<0.001$; Fig. 4). There were no significant differences when treated with hexanic extract (data not shown).

Discussion

Stimulation by LPS increased TNF- α gene expression in the positive control (stimulated with LPS) compared with the negative control (no stimulation with LPS). The expression of p38 α and NF- κB did not change in the positive control compared with the negative control. Herath *et al* (42) demonstrated that different serotypes of LPS produce different responses upon cell stimulation. For example, in human gingival fibroblasts there was no activation of the NF- κB pathway by LPS 1435/1449. However, LPS1690 significantly activated the pathway. Other pro-inflammatory genes also exhibit modified activation by different LPS serotypes.

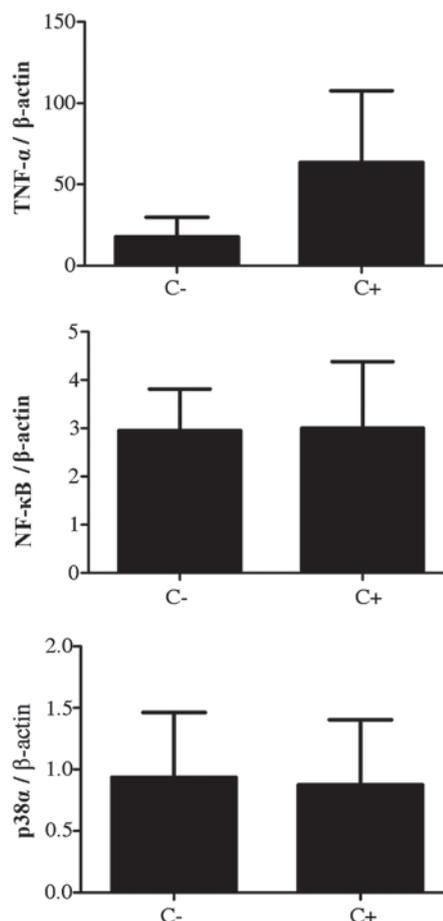


Figure 2. Effect of LPS on gene expression compared with the negative control. Results were analyzed by an unpaired Student's *t* test and are presented as the mean \pm standard error of the mean ($n=5$). P-values for TNF- α , NF- κB and p38 α are $P=0.337$, $P=0.97$ and $P=0.937$, respectively. LPS, lipopolysaccharide; TNF- α , tumor necrosis factor- α ; NF- κB , nuclear factor- κB ; p38 α , p38 α mitogen-activated protein kinase; C-, negative control; C+, positive control.

When evaluating the effect of *Calyptanthes grandifolia* O.Berg extract treatment, on TNF- α gene expression, the present study observed reduced expression compared with the positive control at certain concentrations in both extracts. However, only the ethanolic extract at concentrations of 100 and 200 $\mu\text{g/ml}$ significantly reduced TNF- α expression compared with the positive control. To the best of our knowledge, there are no previous studies that have evaluated the effect of this plant on TNF- α expression, however, Ferreira *et al* (43) evaluated the anti-inflammatory activity of *Campomanesia adamantium* extracts (also a member of the Myrtaceae family) and the production of nitric oxide (NO), TNF- α and IL-10 in J774.A1 macrophages stimulated with LPS/interferon- γ . Paw edema in mice was inhibited by oral administration of the extracts, it inhibited NO and TNF- α production, and increased IL-10 production. Thus, the anti-inflammatory activity of the extract may be associated with the inhibition of pro-inflammatory cytokines and increased IL-10. Li *et al* (44) evaluated the activity of flavonoids in RAW 264.7 cells and demonstrated the inhibition of NO, TNF- α , IL-6 and IL-1 β , indicating an important role for flavonoids in anti-inflammatory activity (44).

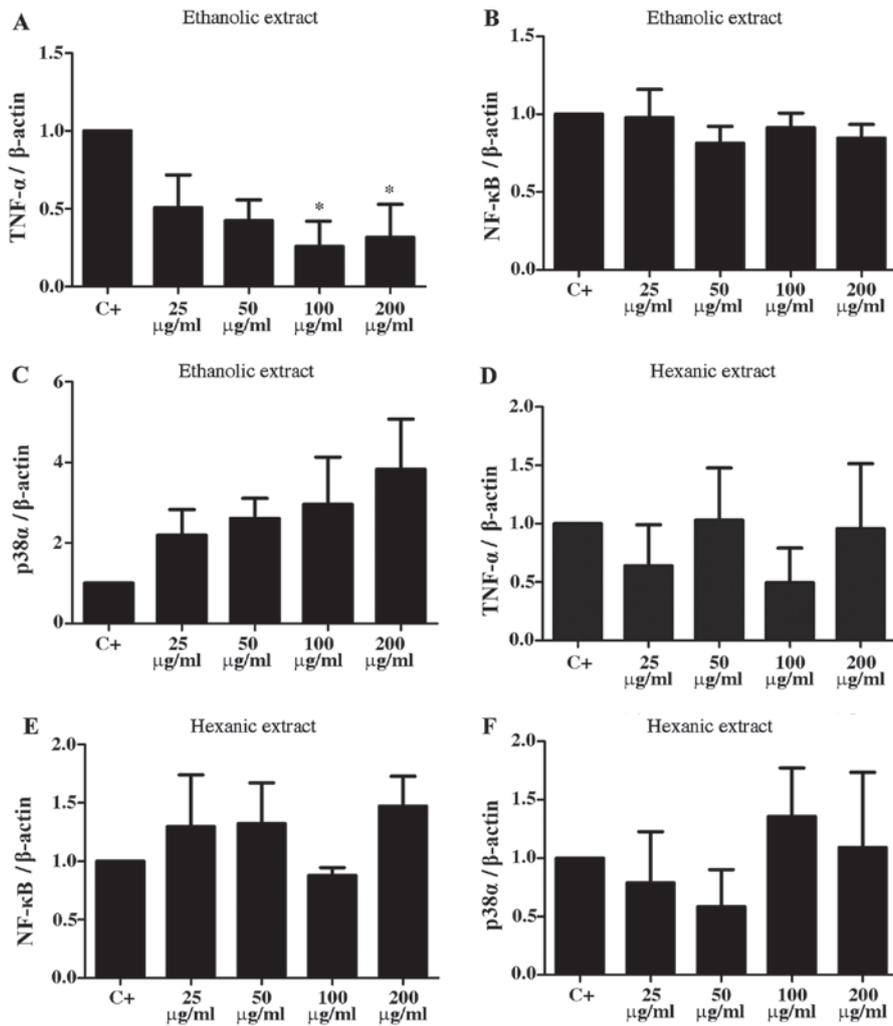


Figure 3. Representative plots of the effect of various concentrations of ethanol and hexane extracts on the gene expression of TNF- α , NF- κ B and p38 α compared with the positive control. Effect of ethanol extract on the gene expression of (A) TNF- α , (B) NF- κ B and (C) p38 α . Effect of hexane extract on the gene expression of (D) TNF- α , (E) NF- κ B and (F) p38 α . Data are presented as the mean + standard error of the mean, n=5. *P<0.025 vs. C+. TNF- α , tumor necrosis factor- α ; NF- κ B, nuclear factor- κ B; p38 α , p38 α mitogen-activated protein kinase; C+, positive control.

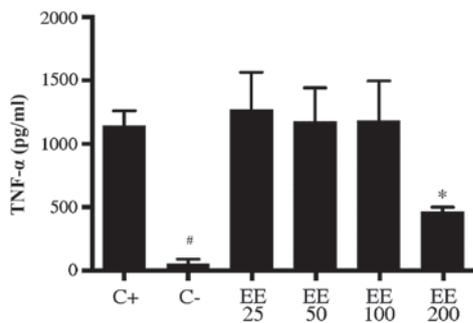


Figure 4. Evaluation of the inhibition of TNF- α cytokine release following treatment with ethanol extract in RAW264.7 cells stimulated with lipopolysaccharides. Data are presented as the mean \pm standard error of the mean with 3 replicates. #P<0.01 vs. C+. TNF- α , tumor necrosis factor- α ; C+, positive control; C-, negative control; EE, ethanol extract.

The present study observed no changes in NF- κ B expression between the different extracts and concentrations tested. The activation of NF- κ B in Caco-2 cells may be associated with the state of cell differentiation. NF- κ B signaling pathways or I κ B kinase do not exist in isolation, therefore, various

mechanisms integrate their activity with other signaling pathways (45). NF- κ B is a protein that modulates the apoptotic response as it is a transcription factor that protects against and contributes to apoptosis (46). The nuclear factor is a central regulator of immune responses and has an important role in the expression of cytokine genes, including IL-2, IL-6, C-C motif chemokine 2 and CD40 ligand. Furthermore, it is also involved in the expression of genes associated with cell survival and proliferation, including cyclin D1, cyclin D2, c-Myc, c-Myb, cyclooxygenase-2, Bcl-2 and Bcl-x1 (47,48). Thus, NF- κ B is considered as a tumor promoter and is frequently identified as constitutively active in tumors (49-52). NF- κ B may not have been activated due to the high expression of TNF- α , and thus by anti-inflammatory signals.

The present study observed no significant differences in p38 α gene expression between the various treatments and concentrations. However, its expression did increase with increasing concentrations of ethanol extract, with the highest expression at a concentration of 200 μ g/ml. The activation of p38 isoforms is specifically controlled by different regulators and they are co-activated by several combinations (53,54). The

p38 α MAPK activates other kinases and therefore regulates diverse cellular responses. Thus, p38 signaling may be associated with inflammation, the cell cycle, cell death, development, differentiation, senescence and tumorigenesis (55). The anti-tumor effect of various chemotherapeutic drugs is based on apoptosis through p38 α activation. However, it is important to note that this kinase is associated with various responses, and may also be involved in resistance to chemotherapy in certain types of tumors (56). Campbell *et al* (57) demonstrated that a modification of p38 α MAPK inhibited TNF- α production in macrophages induced by LPS, post-transcriptionally. Furthermore, p38 negatively regulated the expression of NF- κ B, which allows transcriptional control of TNF- α as the nuclear factor is required for its expression (57). Despite the pro-apoptotic role of p38 α , p38 α MAPK has also been previously associated with an anti-apoptotic activity (58-61). Comes *et al* (62) demonstrated that p38 α induced the survival of colorectal cancer cells by inhibiting autophagy. Another mechanism involved in p38 α -induced survival is the activation of the activating transcription factor 6 α -Ras homolog enriched in brain-mechanistic target of rapamycin (ATF6 α -Rheb-mTOR) pathway, which promoted the survival of dormant tumor cells *in vivo* (63). Although the present study observed no significant differences in the mRNA levels of p38 α and NF- κ B, further studies are required with larger sample sizes, as well as studies that investigate the expression at the protein level, to verify the presence and activation of these proteins. The increase in mRNA levels is not directly proportional to the amount of protein translated, due to transcriptional and post-translational modifications.

The extracts included in the present study did not affect the cell viability when compared with the negative control. According to the US National Cancer Institute, an extract may be considered active or cytotoxic when it presents cytotoxicity with IC₅₀ values <30 μ g/ml (half maximal inhibitory concentration) (64). In this aspect, cytotoxic extracts maybe potential candidates for anti-carcinogenic studies (65). However, it is important to note that these statements should be tested in other cell lines in order to observe potential selective cytotoxicity.

The antioxidant activity was also determined in ethanolic and hexane extracts of *Calyptanthes grandifolia* by the research group (data not shown). This activity was evaluated by antioxidant activity testing, by capturing the free radical 2,2-diphenyl-1-picryl-hidrazila (66). The results demonstrated that the hexane extract exhibited no antioxidant activity, however, the ethanolic extract exhibited a dose dependent antioxidant activity. The observed antioxidant activity may be associated with a potential anti-inflammatory activity of the extract, based on the results observed for gene expression and TNF- α cytokine release. The ethanolic extract had significant antioxidant activity compared with hexane, which corresponds with the decrease of TNF- α gene expression.

Based on the significant reduction of TNF- α when Caco-2 cells were treated with the ethanolic extract of *Calyptanthes grandifolia* O.Berg, the significant inhibition of TNF- α cytokine release in RAW 264.7 murine macrophages, antioxidant activity of the extract and the lack of effects on cell viability, it is indicated that this extract may

have anti-inflammatory potential. However, further studies are required to elucidate the signaling pathway that may be activated. Given the variation between experiments and potential post-transcriptional regulation, the analysis of other genes is required in order to assess the potential pathways implicated. Furthermore, it is important to analyze protein expression in order to confirm that the expressed genes in Caco-2 cells are translated and determine the respective levels of translation.

In conclusion, *Calyptanthes grandifolia* ethanolic extract at concentrations of 100 and 200 μ g/ml significantly reduced TNF- α gene expression in the Caco-2 cell line. There were no significant differences in p38 α and NF- κ B gene expression. Cells treated with hexane extract exhibited no significant variations in the expression of the genes investigated at any of the concentrations. Ethanolic extract at 200 μ g/ml significantly inhibited TNF- α pro-inflammatory cytokine release. The extracts were not considered to be cytotoxic and are not candidates for anti-carcinogenic studies in this lineage. However, other studies using different cell lines maybe performed to identify selective cytotoxicity. The results of the present study indicate that ethanolic extract has an anti-inflammatory potential by decreasing expression of TNF- α . Thus, it is important to investigate its genotoxicity and to conduct *in vivo* analysis to confirm its anti-inflammatory potential.

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