

Molecular mechanisms of autophagy in cardiac ischemia/reperfusion injury (Review)

XIAO-LONG LIN^{1*}, WEI-JIN XIAO^{2*}, LE-LE XIAO³ and MI-HUA LIU⁴

¹Department of Pathology, Hui Zhou Third People's Hospital, Guangzhou Medical University, Huizhou, Guangdong 516002; ²Department of Pathology, The Central Hospital of Shaoyang, Hunan 422000;

³School of Medicine, Huzhou University, Huzhou, Zhejiang 313000; ⁴Department of Infectious Diseases, Centre for Lipid Research and Key Laboratory of Molecular Biology for Infectious Diseases (Ministry of Education), Institute for Viral Hepatitis, The Second Affiliated Hospital, Chongqing Medical University, Chongqing 400016, P.R. China

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Abstract. Autophagy is a maintenance process for recycling long-lived proteins and cytoplasmic organelles. The level of this process is enhanced during ischemia/reperfusion (I/R) injury. Autophagy can trigger survival signaling in myocardial ischemia, whereas defective autophagy during reperfusion is detrimental. Autophagy can be regulated through multiple signaling pathways in I/R, including Beclin-1/class III phosphatidylinositol-3 kinase (PI-3K), adenosine monophosphate activated protein kinase/mammalian target of rapamycin (mTOR), and PI-3K/protein kinase B/mTOR pathways, which consequently lead to different functions. Thus, autophagy has both protective and detrimental functions, which are determined by different signaling pathways and conditions. Targeting the activation of autophagy can be a promising new therapeutic strategy for treating cardiovascular disease.

Contents

1. Introduction
2. Autophagy
3. Autophagy and cardioprotection
4. Mechanism of autophagy in cardioprotection
5. Autophagy as a therapeutic target for I/R injury
6. Conclusions

Correspondence to: Dr Mi-Hua Liu, Department of Infectious Diseases, Centre for Lipid Research and Key Laboratory of Molecular Biology for Infectious Diseases (Ministry of Education), Institute for Viral Hepatitis, The Second Affiliated Hospital, Chongqing Medical University, 1 Yixueyuan Road, Chongqing 400016, P.R. China
E-mail: mihualiu@163.com

*Contributed equally

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1. Introduction

Myocardial ischemia was one of the main causes of sudden cardiac death in the past decades (1). Acute myocardial infarction is a leading cause of death worldwide (2). Strategies for reducing ischemia/reperfusion (I/R)-induced injury in cardiomyocytes are receiving considerable attention due to the failure of cardiomyocytes to regenerate (1). Coronary reperfusion is the most effective treatment for ischemic diseases. However, it may initially aggravate cellular damage during the ischemic period (3).

Cardiac ischemic preconditioning (IPC), which is achieved through repeated brief I/R periods, is one of the well-known protective strategies of the myocardium against I/R injury (4). Recently, autophagy has been linked to IPC-mediated cardioprotection (5). In addition, a number of pharmaceutical therapies targeting I/R injury have been developed to orchestrate multiple protein complexes and signaling pathways in autophagy (6,7). In this review, we aim to draw attention to the role of autophagy in cardioprotection.

2. Autophagy

Autophagy is a self-protective mechanism of living cells under various stress conditions (8). During autophagy, cellular cytoplasm constituents are delivered to lysosomes for degradation and recycling (9). Autophagy limits the production of reactive oxygen species and excessive protein aggregation to maintain intracellular or extracellular homeostasis (10). Autophagy has emerged as a potential drug target for numerous diseases including cancer, neurodegenerative diseases, and cardiovascular disease (11,12).

Autophagy plays multifaceted roles in heart and diseases (13). Under basal conditions, autophagy is a maintenance process for recycling long-lived proteins and cytoplasmic organelles in the heart (14). Furthermore, autophagy plays significant roles in starvation, aging, inflammation, and reverse cardiac remodeling by maintaining cellular homeostasis (15). Autophagy can be regarded as an end effector in hypoxic and ischemic conditions to eliminate superfluous, damaged, or aged cells or organelles (16-18). However, this process will

cause detrimental autophagic cell death when triggered by severe ischemia or in cardiovascular diseases (19).

3. Autophagy and cardioprotection

Myocardial I/R injury is a complex process that destroys proteins, DNA, and plasma membrane, thereby resulting in cell death and decreased cardiac output (20,21). Many studies have reported an increase in the number of autophagosomes in the heart during I/R in animal models (5,22). Autophagy induced by ischemia was subsequently enhanced by reperfusion in isolated rabbit hearts (23) and in mouse hearts (24). The activation of autophagy is reflected in the abundance of autophagy-related protein pathways, such as light chain 3 (LC3), Beclin-1, autophagy-related gene (ATG) 5-12 complex, and p62 (25-27). Hu *et al* reported that approximately 20 min of aortic clamping with hyperkalemic cold blood cardioplegia to achieve total autophagy, which in accordance with previous evidence (28). The abundance of autophagic proteins will actually decrease with the progress of autophagy because of self-degradation (25,26). In particular, in biopsies from the right atrial appendage of patients undergoing valve surgery or coronary artery bypass grafting, the expression of autophagy-related proteins, including LC3-I, LC3-II, ATG5-12, Beclin-1, and p62, is reduced during reperfusion (26).

Until recently, the debate continues whether autophagy plays a protective or deleterious role in the I/R injury process. On the one hand, modest levels of autophagy triggered by mild to moderate hypoxia/ischemia are protective and seem to prevent the activation of apoptosis (23,29). On the other hand, high levels of autophagy induced by severe hypoxia or I/R may cause self-digestion and eventual cell death (30). Therefore, autophagic flux induced by ischemia during the early stage of I/R has been speculated to be beneficial; however, it is harmful during reperfusion at the later stage of I/R (15,19).

Autophagy may play an alternative role in I/R, which determines cell fate. The extent of autophagy in response to ischemia is considered based on the severity and duration of ischemic insults (31). Nutrient and oxygen deprivation in the heart threatens cellular survival during I/R, and increased autophagy may provide at least a temporary reprieve for a threatened myocardium by serving as a source of intracellular nutrients (32). Oxidative stress, calcium overload, endoplasmic reticulum (ER) stress, and mitochondrial dysfunction maintain a high level of autophagy during reperfusion (33). However, high levels or long-term upregulation of autophagy can lead to excessive degradation of essential proteins and organelles (34). If intracellular energy sources become inadequate, then autophagic processes will be a particular form of cell death, called type II or autophagic cell death (35). In fact, aware that necrosis and apoptosis are not the only mechanisms of cell death is increasing (36). Autophagic cell death has been identified as a cell death phenotype via electron microscope observations; it has a morphological term characterized by abundant autophagic vacuoles in the cytoplasm (37,38).

Moreover, increased autophagy after I/R is not due to increased autophagosome formation, but instead, to impaired clearance of autophagosomes (39); this assumption is derived from the concept of autophagic flux (40). Furthermore, a rapid decline induced by reperfusion in LAMP2, which is

a critical protein for autophagosome-lysosome fusion, can impair autophagosome processing and mitochondrial permeabilization, thereby increasing ROS generation and triggering cardiomyocyte death (41). In addition, when the engulfed targets or autophagosomes cannot fuse with lysosomes and digest their contents, a cell may eject the autophagosomes as a response, which induces an acute and significant inflammatory response (42).

4. Mechanism of autophagy in cardioprotection

Autophagy is a complex and dynamic multi-step process that depends on strict regulation and coordination through multiple signaling pathways (43). To date, several cellular signaling pathways are considered to trigger autophagy in I/R. In addition, autophagy has been shown to be regulated by several signaling pathways (44), including Beclin-1/class III phosphatidylinositol-3 kinase (PI-3K), AMPK/mammalian target of rapamycin (mTOR), and PI-3K/Akt/mTOR pathways.

Beclin-1/class III PI-3K pathway. Beclin-1, which is a phylogenetically conserved protein, the mammalian homologue of the yeast Atg6, and the interacting protein of the anti-apoptotic protein Bcl-2, is a key molecule involved in mediating autophagy (45,46). It plays a crucial role in engaging class III PI-3K to positively modulate autophagy in mammalian cells (47,48). Autophagy in mammalian cells is reported to be activated by the class III PI-3K complex, which contains Vps34 and Beclin-1 (29,49). Moreover, a coiled-coil domain (aa 140-268) is present in this 450 amino acid-long protein in Beclin-1; this domain can mediate binding to class III PI-3K Vps34 by interacting with an evolutionarily conserved domain (ECD; aa 244-337) (50). RNA interference of Beclin-1, which inhibits autophagy, will subsequently enhance cardiac cell survival (51).

Autophagy is involved in delayed cardioprotection induced by sevoflurane preconditioning (52). Sevoflurane preconditioning reduces the autophagy induced by H/R by decreasing the Beclin-1 expression (52). Accordingly, IPC protects the rat heart against MI/R injury by inhibiting Bcl-2 dissociation from Beclin-1 during the reperfusion phase *in vivo*, although IPC-induced autophagy reflects a compensatory pro-survival response to I/R injury (53). Bcl-2 is the prototype of a protein family, which contains at least one Bcl-2 homology (BH) region (54). Bcl-2 binding molecules have been recently shown to regulate autophagy activation (55). Transgenic mice with a cardiac-specific overexpression of Bcl-2 are protected from I/R injury (56,57). Autophagy is disrupted when Bcl-2 binds to Beclin-1 (58). In addition, when a mutant of Beclin-1 that lacks the Bcl-2 binding domain is overexpressed in cells, excessive autophagy and cell death are induced (47). Bcl-2 can also inhibit Beclin-1/Vsp34 PI-3K complex formation and the activity of Beclin-1-associated class III PI-3K (53). Furthermore, the class III PI-3K autophagic pathway is inhibited by combining the BH3 hydrophobic groove in Bcl-2 and the BH3-like amphipathic α -helix in Beclin-1 (59). However, the interaction with Bcl-2 (and Bcl-xL) in the ER, rather than in the mitochondria, inhibits the Beclin-1 activity in autophagy (60). The interaction between Bcl-2 and Beclin-1 maintains

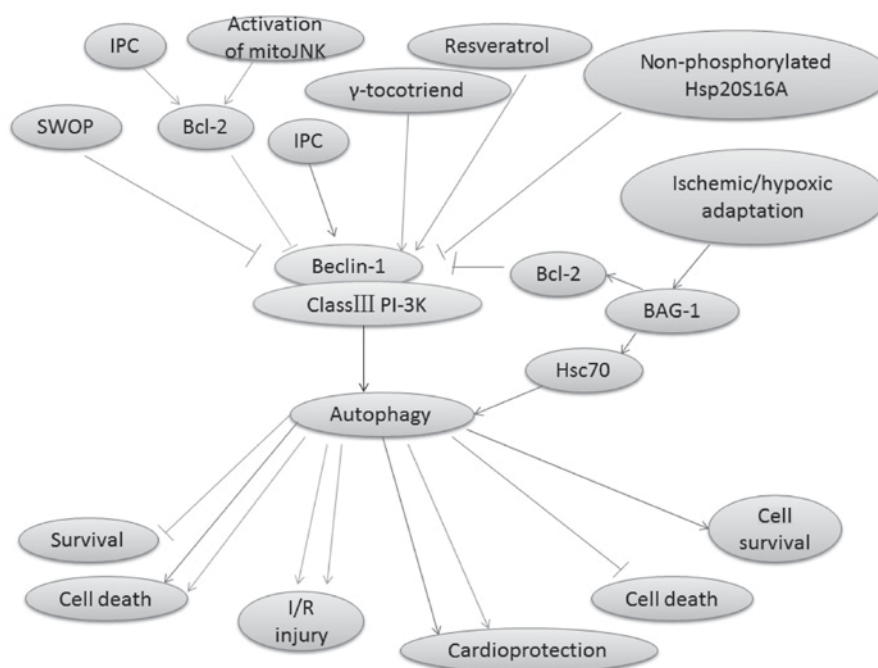


Figure 1. Beclin-1/class III PI-3K pathway-regulated autophagy and autophagy-mediated function in I/R injury. Under I/R condition, autophagy is activated by the class III PI-3K complex, which contains class III PI-3K Vps34 and Beclin-1. SWOP induces cardioprotection by reducing Beclin-1 expression, increasing the survival rate of cells, and reducing their apoptosis percentage. IPC protects rat heart against myocardial I/R injury by inhibiting Bcl-2 dissociation from Beclin-1. MitoJNK activation, instead of JNK mitochondrial localization, induces triggering of Bcl2-regulated autophagy, which further causes cell death and aggravates myocardial I/R injury. However, IPC can exert cardioprotection via autophagy through the activation of PI-3K. Resveratrol and γ -tocotrienol can enhance autophagy through the induction survival pathway, which depends on class III PI-3K, thereby synergistically providing an increased degree of cardioprotection. Non-phosphorylated Hsp20S16A increases cell death by suppressing autophagy, and Beclin-1 is a potential target of phosphorylated Hsp20 in regulating autophagy. Ischemic/hypoxic adaptation induces improvement in cardiac cell survival mediated by BAG-1. BAG-1 can bind to both Bcl-2 and Hsc70 molecules, and may activate autophagy via Hsc70. PI-3K, phosphatidylinositol-3 kinase; I/R, ischemia/reperfusion; JNK, C-Jun N-terminal kinase; IPC, ischemic preconditioning; Hsp20, heat shock protein.

autophagy at levels (47). Blocking the interaction between the BH3 domains of Beclin-1 and Bcl-2 increases autophagic activity (53). Recent studies indicate that the increase in the interaction between Beclin-1 and Bcl-2 is caused by IPC (53). C-Jun N-terminal kinase (JNK), which is a member of an evolutionarily conserved subfamily of mitogen-activated protein kinases, is critical for the cellular responses of multiple environmental and cellular stimuli (61,62). I/R can trigger Bcl2-regulated autophagy by inducing a dominant increase in mitoJNK activation, which causes cell death (63). Xu *et al* reported that mitoJNK activation, and not JNK mitochondrial localization, induced autophagy, which further aggravates I/R injury (63). In addition, the mitoJNK phosphorylate Bcl2, which antagonizes Bcl2 anti-apoptotic and anti-autophagic activities, may contribute to the deleterious role of mitoJNK in I/R injury (64,65).

Heat shock protein (Hsp20) is the only member of the sHsps family that contains the consensus peptide motif RRAS for protein kinase A-/protein kinase G-dependent phosphorylation at Ser16 (66). Qian *et al* demonstrated that non-phosphorylated Hsp20S16A is detrimental in I/R injury because it suppresses autophagy and further increases cell death (36). Ischemic/hypoxic adaptation improves cardiac cell survival by suppressing the BAG-1 protein expression (67). BAG-1 can bind with both Bcl-2 and Hsc70 molecules (67). Autophagosomal membrane contains a significantly higher amount of Hsc70 proteins (68). BAG-1 has been shown exhibit numerous functions through its interaction with Hsc70 (69). The treatment of rats with wortmannin, an

inhibitor of class III PI-3K, has been used to suppress autophagy in many studies (70,71), and attenuates both the LC3-II and BAG-1 protein expressions (67). Zheng *et al* (72) reported that the activated PI3K/Akt pathway contributes to the berberine postconditioning-induced cardioprotection through modulating autophagy. The Beclin-1/class III PI-3K pathway-regulated autophagy and autophagy-mediated function in I/R injury are shown in Fig. 1.

AMPK/mTOR pathway. AMPK, which is activated in response to stress that exhaust cellular ATP supplies, such as ischemia and hypoxia, plays a crucial role as a master regulator of cellular energy homeostasis (73). AMPK is ubiquitously expressed in metabolically active tissues, such as cardiac muscles, and activated upon the depletion of energy stores by functioning as an intracellular fuel sensor (74). Ischemia has been proposed to stimulate autophagy via an AMPK-dependent mechanism (53), which is one of the most significant approaches in upregulating autophagy (75,76). During I/R injury, intracellular ATP stores are rapidly consumed and cannot be supplemented with decreasing glucose supply (77). AMPK signaling can positively regulate autophagy by activating Ulk1 via the phosphorylation of Ser 317 and Ser 777 or indirectly by inhibiting mTOR signaling (78,79). Moreover, AMPK functions as a master regulator of the autophagy pathway through inactivating mTOR (80).

High mTOR activity negatively regulates autophagy by inhibiting the activation of Ulk1, which is one of the

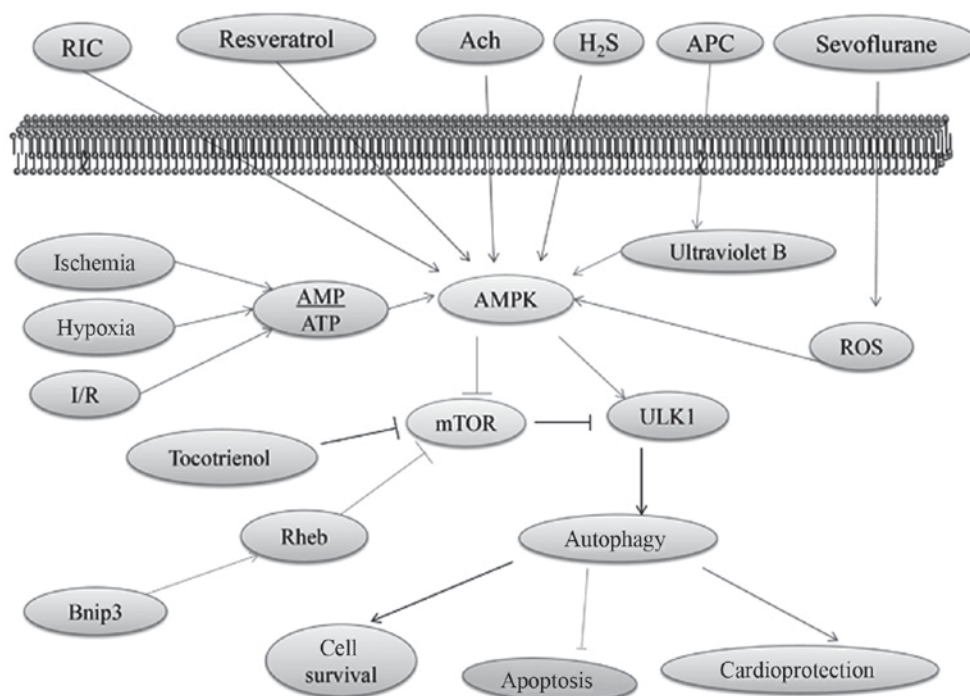


Figure 2. AMPK/mTOR pathway-regulated autophagy and autophagy-mediated function in I/R injury. Tocotrienol induces autophagy through the mTOR pathway, which subsequently leads to cell survival and cardioprotection. mTOR activity negatively regulates autophagy by inhibiting Ulk1 activation, which initiates the nucleation of the autophagic membrane. Bnip3 can inhibit the mTOR pathway and induce autophagy by directly binding to Rheb and protect cardiac myocytes against I/R injury-related apoptosis. Under the condition of ischemia, hypoxia, or I/R, AMPK is activated by the increased levels of AMP/ATP. Then, autophagy triggered by AMPK can resist cardiac injury, and AMPK signaling can positively regulate autophagy by activating Ulk1, or indirectly, by inhibiting mTOR signaling. RIC, resveratrol, Ach, and H₂S can induce the autophagic AMPK pathway, which is cardioprotective. Ultraviolet B is a critical mediator of cardioprotection via APC, and ultraviolet B-induced autophagy activates AMPK by inhibiting the phosphorylation of GSK3 β . Sevoflurane provides cardioprotection against I/R injury via ROS-mediated upregulation of autophagy. mTOR, mammalian target of rapamycin; I/R, ischemia/reperfusion; Rheb, Ras homolog that is enriched in the brain.

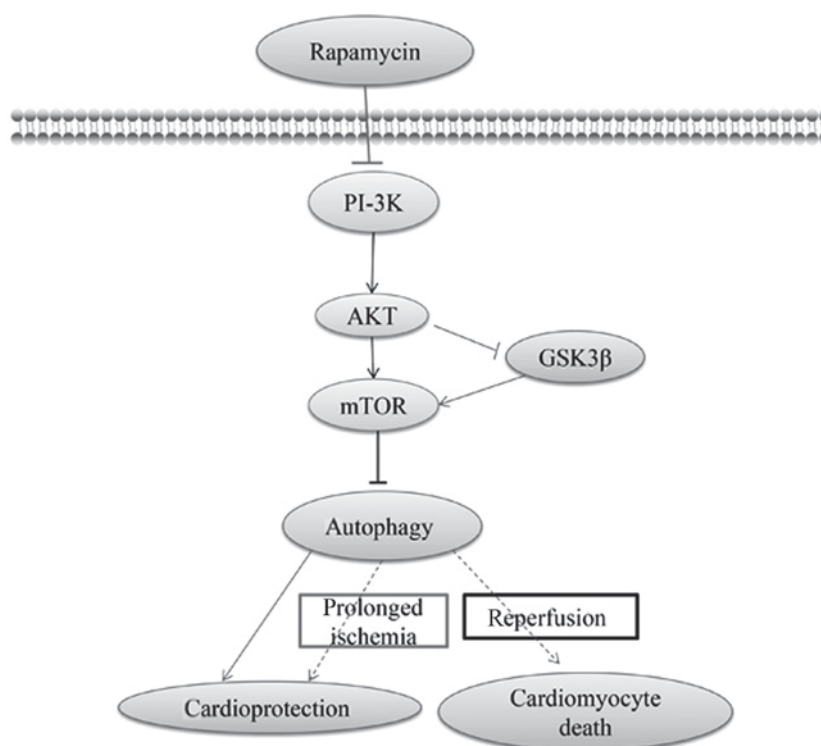


Figure 3. PI-3K/Akt/mTOR pathway-regulated autophagy and autophagy-mediated function in I/R injury. Class I PI-3K (PI-3K) will inhibit the induction of autophagy via the phosphorylation of Akt and mTOR, and achieve additional benefit against I/R injury. Rapamycin exerts beneficial effects against cardiomyocyte I/R injury through autophagy that depends on the PI-3K-Akt signaling pathway. GSK-3 β is the downstream of PI-3K/Akt, and GSK-3 β causes cardiomyocyte death during reperfusion but mediates the survival of cardiomyocytes during prolonged ischemia through mTOR-dependent autophagy attenuation. PI-3K, phosphatidylinositol-3 kinase; mTOR, mammalian target of rapamycin.

Table I. Summary of various autophagy modulators on cardiac ischemia/reperfusion injury.

Author, year	Drugs	Mechanism of action	Effect on cardiac ischemia/reperfusion injury	(Refs.)
Huang <i>et al</i> , 2010	Sulfaphenazole	Activate protein kinase C	Protects against myocardial I/R and reduces infarct size	(39)
Xie <i>et al</i> , 2015	Sevoflurane	Inhibition Beclin 1-mediated autophagic cell death	Delayed cardioprotection	(52)
Zheng <i>et al</i> , 2017	Berberamine	Activate PI3K/Akt pathway	Improved post-ischemic myocardial function and attenuated cell death	(72)
Lekli <i>et al</i> , 2010	Tocotrienol	Activate the mTOR pathway	Reduces cardiomyocyte apoptosis	(84)
Gurusamy <i>et al</i> , 2010	Resveratrol	Activate mTOR-Rictor survival pathway	Attenuates myocardial I/R injury and reduces infarct size	(91)
Zhao <i>et al</i> , 2013	Acetylcholine	Activate AMPK-mTOR pathway	Reduces cardiomyocyte death	(92)
Xie <i>et al</i> , 2015	H ₂ S	Activate AMP-activated protein kinase	Protects against myocardial I/R injury	(93)
Zhong <i>et al</i> , 2017	Trimetazidine	Activate AMPK-mTOR pathway	Reduces hypoxia/reoxygenation injury	(99)
Wang <i>et al</i> , 2015	Rapamycin	Activate PI3k/Akt pathway	Attenuates anoxia/reoxygenation injury	(103)

mTOR, mammalian target of rapamycin; I/R, ischemia/reperfusion.

mammalian autophagy-initiating kinases that is important for membrane nucleation via the phosphorylation of Ulk1 Ser 757 (1,81,82). The association between ATG1 and ATG13 is negatively regulated by mTOR, which inhibits autophagy (83). In addition, the activation of Ulk1 and its combination with other molecules, such as ATG13 and FIP200, initiate the nucleation of the autophagic membrane (1). Lekli *et al* proposed that tocotrienol could induce autophagy through the mTOR pathway, which would consequently lead to cell survival and cardioprotection (84). The overexpression of Bnip3, a hypoxia-inducible Bcl-2 homology 3 domain-containing protein (85) and the pro-apoptotic molecule present in the mitochondrial membrane; can upregulate autophagy and protect cardiac myocytes against I/R injury-related apoptosis (86). The high-mobility group box 1 protein (HMGB1)-mediated activation of mTOR inhibits hypoxia and reoxygenation injury in rat cardiomyocytes (87,88). Moreover, Bnip3 can inhibit the mTOR pathway and induce autophagy by directly binding to the Ras homolog that is enriched in the brain (Rheb), which is a Ras-related small guanosine triphosphatase (85,89).

The cardioprotection effect of resveratrol has been shown to induce autophagy by facilitating AMPK activation (90,91). In addition, AMPK expression is elevated with ACh during H/R (92). ACh activates cytoprotective autophagy through the AMPK-mTOR-dependent pathway that is activated by a muscarinic receptor (92). Xie *et al* found that the post-reperfusion AMPK activation induced by a slow-releasing organic H₂S donor that could restore I/R impaired autophagic flux; is critical to H₂S cardioprotection (93). Accordingly, studies have proven that autophagy activation through the AMPK/mTOR pathway plays a cardioprotection role (94). Recently, ultraviolet B-induced autophagy has been found to activate AMPK by inhibiting the phosphorylation of GSK3 β (95), which is a

critical mediator of cardioprotection via anesthetic preconditioning (96). Sevoflurane provides cardioprotection against I/R injury via the ROS-mediated upregulation of autophagy (97). Hariharan *et al* reported that oxidative stress triggers autophagic flux during MI/R injury (98). In addition, trimetazidine (99) and thioredoxin-2 (100) protect against hypoxia/reoxygenation injury by promoting the AMPK-dependent autophagic flux in H9c2 cardiomyocytes. The AMPK/mTOR pathway-regulated autophagy and autophagy-mediated function in I/R injury are illustrated in Fig. 2.

PI-3K/Akt/mTOR pathway. PI-3K/Akt/mTOR signaling may also provide an additional benefit against I/R injury (101). In addition, class III PI-3K focuses on the formation of autophagosomes, whereas class I PI-3K will inhibit the induction of autophagy through the phosphorylation of Akt and mTOR (70). Thus, the interaction of Akt with mTOR is multifaceted and bidirectional (101). Moreover, the self-regulation of autophagy has been postulated to be regulated by the autophagy-induced inhibition of mTOR (102). Furthermore, rapamycin provided a strong beneficial effect against cardiomyocyte anoxia/reoxygenation injury, which would mediate cardioprotection via autophagy that probably depended on the PI-3K/Akt signaling pathway (103). During prolonged ischemia and I/R, the differential effects of GSK-3 β , which is the downstream of PI-3K/Akt, on myocardial injury has been suggested to be determined by changes in autophagy (104). GSK-3 β inhibition modulates mTOR-dependent attenuation of autophagy, thereby causing the death of cardiomyocytes during prolonged ischemia while mediating their survival during reperfusion (104). In addition, mTOR activation via GSK-3 β has been suggested to provide cardioprotection via autophagy (104). The PI-3K/Akt/mTOR pathway-regulated

autophagy and autophagy-mediated function in I/R injury are illustrated in Fig. 3.

Others. The p53 transcription factor is a major regulator of cellular response to acute stress (105). Knockdown of p53 can activate autophagy in cardiomyocytes, thereby protecting the myocardium against ischemic injury (106). Autophagy is inhibited with STAT1 to modulate stress response to I/R in STAT1^{-/-} null hearts (32). In I/R, STAT1 can interact directly with p53 and regulate its functional activity (107), thereby suggesting that STAT1 can act with p53 to modulate autophagy (32). The mitochondrial permeability transition pore (MPTP) plays an important role in myocardial I/R injury, and the opening of MPTP has also been shown to trigger autophagy (108). Making the heart more tolerant to subsequent I/R injury is a crucial step in transient MPTP opening before prolonged ischemia (109,110). Moreover, PKC has been reported to trigger the phosphorylation of a regulatory sub-unit of VPATPase, which subsequently induces autophagy (111-113).

5. Autophagy as a therapeutic target for I/R injury

Microarray analysis showed that autophagy-associated genes and the unfolded protein response were upregulated under the condition of repetitive coronary occlusion achieved during chronic local ischemic conditioning in mice (114,115). In another study, inhibiting mTOR decreased infarct size in mice (116). Rapamycin (116), caloric restriction (117), exercise (118), nitric oxide (119), and lipopolysaccharide (120) has been identified as cardioprotective interventions for triggering autophagy. Gurusamy *et al* used isolated rat heart models and demonstrated that the induction of IPC via repeated I/R cycles immediately enhanced the expression of LC3-II and Beclin-1 (67). *In vivo* swine models, infarct size was limited after chloramphenicol succinate was used before ischemia (121). Han *et al* found that cardioprotection induced by remote limb ischemic postconditioning was associated with elevated autophagy 3 h post-reperfusion (2). A similar phenomenon was observed by Hamacher-Brady *et al* in HL-1 myocytes; they found that simulated I/R-mediated cell death was prevented by strengthening autophagy, whereas its inhibition caused cell death (40). Other researchers have observed that blocking autophagy via cell-permeable Tat-Atg5K130R concurrently increased infarct size in hearts when treated with SUL (39). Moreover, Tibetan patients with coronary heart disease resist I/R injury during cardiac surgery better than patients living at sea level, which is possibly correlated with the upregulation of basal autophagy resulting from chronic hypoxia (28).

Autophagy has been determined as a significant element of the endogenous defense mechanisms activated by various preconditioning types. Induction of autophagy may represent a novel therapeutic approach to myocardial protection in humans (39). The identification of agents that can rapidly induce autophagy can contribute to the discovery of new cardioprotective drugs (122). In addition, induction of autophagy can preserve heart function during I/R injury (91,121,123). Other studies have suggested that autophagy is detrimental because it contributes to cell death (15,124). The beneficial or detrimental role of autophagy may be a consequence of balance, depending

on the extent of autophagy (7). Thus, for autophagy to be effective, searching for a candidate cardioprotective drug that can induce autophagy in a target population is important, together with the appropriate timing and response magnitude (16). Various autophagy modulators on cardiac ischemia/ injury are summarized in Table I.

6. Conclusions

Recent studies have shown that autophagy plays an important role in I/R injury. Moreover, evidence has emerged that autophagy plays various roles in I/R through multiple mechanisms. Autophagy can trigger a survival signal in the case of myocardial ischemia, whereas defective autophagy during reperfusion is detrimental. Although we have obtained substantial knowledge about the function of autophagy in I/R injury, the autophagy pathway is highly complex and remains far from being understood completely. Additional studies are necessary to identify the molecular components of the autophagy pathway, characterize the role of autophagy in I/R injury, elucidate the diverse processes that regulate autophagy expression and activity, and determine the contribution of autophagy to myocardial infarction protection in humans. Such studies will provide additional insights into the role of autophagy in I/R injury and potentially discover novel therapeutic strategies for treating the diseases.

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Authors' contributions

X-LL and M-HL conceived and designed this review. X-LL, L-LX and W-JX contributed the central idea, analyzed most of the data, and wrote the initial draft of the paper. L-LX and M-HL revised the manuscript.

Ethics approval and consent to participate

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Consent for publication

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Competing interests

The authors declare that they have no competing interests.

References

- Yang K, Xu C, Li X and Jiang H: Combination of D942 with curcumin protects cardiomyocytes from ischemic damage through promoting autophagy. *J Cardiovasc Pharmacol Therap* 18: 570-581, 2013.
- Han Z, Cao J, Song D, Tian L, Chen K, Wang Y, Gao L, Yin Z, Fan Y and Wang C: Autophagy is involved in the cardioprotection effect of remote limb ischemic preconditioning on myocardial ischemia/reperfusion injury in normal mice, but not diabetic mice. *PLoS One* 9: e86838, 2014.
- Murphy E and Steenbergen C: Mechanisms underlying acute protection from cardiac ischemia-reperfusion injury. *Physiol Rev* 88: 581-609, 2008.
- Das DK and Maulik N: Preconditioning potentiates redox signaling and converts death signal into survival signal. *Arch Biochem Biophys* 420: 305-311, 2003.
- Aoyagi T, Kusakari Y, Xiao CY, Inouye BT, Takahashi M, Scherrer-Crosbie M, Rosenzweig A, Hara K and Matsui T: Cardiac mTOR protects the heart against ischemia-reperfusion injury. *Am J Physiol Heart Circ Physiol* 303: H75-H85, 2012.
- Li Y, Xiang Y, Zhang S, Wang Y, Yang J, Liu W and Xue F: Intramyocardial injection of thioredoxin 2-expressing lentivirus alleviates myocardial ischemia-reperfusion injury in rats. *Am J Transl Res* 9: 4428-4439, 2017.
- Sasaki Y, Ikeda Y, Iwabayashi M, Akasaki Y and Ohishi M: The impact of autophagy on cardiovascular senescence and diseases. *Int Heart J* 58: 666-673, 2017.
- Yang L, Wang H, Shen Q, Feng L and Jin H: Long non-coding RNAs involved in autophagy regulation. *Cell Death Dis* 8: e3073, 2017.
- Green DR, Galluzzi L and Kroemer G: Mitochondria and the autophagy-inflammation-cell death axis in organismal aging. *Science* 333: 1109-1112, 2011.
- Sica V, Galluzzi L, Bravo-San Pedro JM, Izzo V, Maiuri MC and Kroemer G: Organelle-specific initiation of autophagy. *Mol Cell* 59: 522-539, 2015.
- Mowers EE, Sharifi MN and MacLeod KF: Functions of autophagy in the tumor microenvironment and cancer metastasis. *FEBS J*, 2018 doi: 10.1111/febs.14388.
- Pellacani C and Costa LG: Role of autophagy in environmental neurotoxicity. *Environ Pollut* 235: 791-805, 2018.
- Yang X, Cohen MV and Downey JM: Mechanism of cardioprotection by early ischemic preconditioning. *Cardiovasc Drugs Ther* 24: 225-234, 2010.
- Levine B and Klionsky DJ: Development by self-digestion: Molecular mechanisms and biological functions of autophagy. *Dev Cell* 6: 463-477, 2004.
- Ma H, Guo R, Yu L, Zhang Y and Ren J: Aldehyde dehydrogenase 2 (ALDH2) rescues myocardial ischaemia/reperfusion injury: Role of autophagy paradox and toxic aldehyde. *Eur Heart J* 32: 1025-1038, 2011.
- Huang C, Yitzhaki S, Perry CN, Liu W, Giricz Z, Mentzer RM Jr and Gottlieb RA: Autophagy induced by ischemic preconditioning is essential for cardioprotection. *J Cardiovasc Transl Res* 3: 365-373, 2010.
- Shintani T and Klionsky DJ: Autophagy in health and disease: A double-edged sword. *Science* 306: 990-995, 2004.
- Rubinshtein DC, Gestwicki JE, Murphy LO and Klionsky DJ: Potential therapeutic applications of autophagy. *Nat Rev Drug Discov* 6: 304-312, 2007.
- Sciarretta S, Hariharan N, Monden Y, Zablocki D and Sadoshima J: Is autophagy in response to ischemia and reperfusion protective or detrimental for the heart? *Pediatr Cardiol* 32: 275-281, 2011.
- Go AS, Mozaffarian D, Roger VL, Benjamin EJ, Berry JD, Blaha MJ, Dai S, Ford ES, Fox CS, Franco S, *et al*: Executive summary: Heart disease and stroke statistics-2014 update: A report from the American Heart Association. *Circulation* 129: 399-410, 2014.
- Bulluck H, Yellon DM and Hausenloy DJ: Reducing myocardial infarct size: Challenges and future opportunities. *Heart* 102: 341-348, 2016.
- Hamacher-Brady A, Brady NR, Logue SE, Sayen MR, Jinno M, Kirshenbaum LA, Gottlieb RA and Gustafsson AB: Response to myocardial ischemia/reperfusion injury involves Bnip3 and autophagy. *Cell Death Differ* 14: 146-157, 2007.
- Decker RS and Wildenthal K: Lysosomal alterations in hypoxic and reoxygenated hearts. I. Ultrastructural and cytochemical changes. *Am J Pathol* 98: 425-444, 1980.
- Matsui Y, Takagi H, Qu X, Abdellatif M, Sakoda H, Asano T, Levine B and Sadoshima J: Distinct roles of autophagy in the heart during ischemia and reperfusion: Roles of AMP-activated protein kinase and Beclin 1 in mediating autophagy. *Circ Res* 100: 914-922, 2007.
- Kassiotis C, Ballal K, Wellnitz K, Vela D, Gong M, Salazar R, Frazier OH and Taegtmeier H: Markers of autophagy are downregulated in failing human heart after mechanical unloading. *Circulation* 120 (11 Suppl): S191-S197, 2009.
- Jahania SM, Sengstock D, Vaitkevicius P, Andres A, Ito BR, Gottlieb RA and Mentzer RM Jr: Activation of the homeostatic intracellular repair response during cardiac surgery. *J Am Coll Surg* 216: 719-729, 2013.
- Schiattarella GG and Hill JA: Therapeutic targeting of autophagy in cardiovascular disease. *J Mol Cell Cardiol* 95: 86-93, 2016.
- Hu Y, Sun Q, Li Z, Chen J, Shen C, Song Y and Zhong Q: High basal level of autophagy in high-altitude residents attenuates myocardial ischemia-reperfusion injury. *J Thorac Cardiovasc Surg* 148: 1674-1680, 2014.
- Hamacher-Brady A, Brady NR and Gottlieb RA: The interplay between pro-death and pro-survival signaling pathways in myocardial ischemia/reperfusion injury: Apoptosis meets autophagy. *Cardiovasc Drugs Ther* 20: 445-462, 2006.
- Gustafsson AB and Gottlieb RA: Eat your heart out: Role of autophagy in myocardial ischemia/reperfusion. *Autophagy* 4: 416-421, 2008.
- Song X, Kusakari Y, Xiao CY, Kinsella SD, Rosenberg MA, Scherrer-Crosbie M, Hara K, Rosenzweig A and Matsui T: mTOR attenuates the inflammatory response in cardiomyocytes and prevents cardiac dysfunction in pathological hypertrophy. *Am J Physiol Cell Physiol* 299: C1256-C1266, 2010.
- McCormick J, Suleman N, Scarabelli TM, Knight RA, Latchman DS and Stephanou A: STAT1 deficiency in the heart protects against myocardial infarction by enhancing autophagy. *J Cell Mol Med* 16: 386-393, 2012.
- Gustafsson AB and Gottlieb RA: Autophagy in ischemic heart disease. *Circ Res* 104: 150-158, 2009.
- House SL, Branch K, Newman G, Doetschman T and Schultz Jel J: Cardioprotection induced by cardiac-specific overexpression of fibroblast growth factor-2 is mediated by the MAPK cascade. *Am J Physiol Heart Circ Physiol* 289: H2167-H2175, 2005.
- Baehrecke EH: Autophagy: Dual roles in life and death? *Nat Rev Mol Cell Biol* 6: 505-510, 2005.
- Qian J, Ren X, Wang X, Zhang P, Jones WK, Molkentin JD, Fan GC and Kranias EG: Blockade of Hsp20 phosphorylation exacerbates cardiac ischemia/reperfusion injury by suppressed autophagy and increased cell death. *Circ Res* 105: 1223-1231, 2009.
- Tsujimoto Y and Shimizu S: Another way to die: Autophagic programmed cell death. *Cell Death Differ* 12 (Suppl 2): S1528-S1534, 2005.
- Galluzzi L, Maiuri MC, Vitale I, Zischka H, Castedo M, Zitvogel L and Kroemer G: Cell death modalities: Classification and pathophysiological implications. *Cell Death Differ* 14: 1237-1243, 2007.
- Huang C, Liu W, Perry CN, Yitzhaki S, Lee Y, Yuan H, Tsukada YT, Hamacher-Brady A, Mentzer RM Jr and Gottlieb RA: Autophagy and protein kinase C are required for cardioprotection by sulfaphenazole. *Am J Physiol Heart Circ Physiol* 298: H570-H579, 2010.
- Hamacher-Brady A, Brady NR and Gottlieb RA: Enhancing macroautophagy protects against ischemia/reperfusion injury in cardiac myocytes. *J Biol Chem* 281: 29776-29787, 2006.
- Huber SM, Misovic M, Mayer C, Rodemann HP and Dittmann K: EGFR-mediated stimulation of sodium/glucose cotransport promotes survival of irradiated human A549 lung adenocarcinoma cells. *Radiother Oncol* 103: 373-379, 2012.
- Gottlieb RA and Mentzer RM: Autophagy during cardiac stress: Joys and frustrations of autophagy. *Annu Rev Physiol* 72: 45-59, 2010.
- Sciarretta S, Zhai P, Shao D, Zablocki D, Nagarajan N, Terada LS, Volpe M and Sadoshima J: Activation of NADPH oxidase 4 in the endoplasmic reticulum promotes cardiomyocyte autophagy and survival during energy stress through the protein kinase RNA-activated-like endoplasmic reticulum kinase/eukaryotic initiation factor 2 α /activating transcription factor 4 pathway. *Circ Res* 113: 1253-1264, 2013.
- Wei L, Wu RB, Yang CM, Zheng SY and Yu XY: Cardioprotective effect of a hemoglobin-based oxygen carrier on cold ischemia/reperfusion injury. *Cardiology* 120: 73-83, 2011.

45. Boya P and Kroemer G: Beclin 1: A BH3-only protein that fails to induce apoptosis. *Oncogene* 28: 2125-2127, 2009.
46. Zalckvar E, Berissi H, Eisenstein M and Kimchi A: Phosphorylation of Beclin 1 by DAP-kinase promotes autophagy by weakening its interactions with Bcl-2 and Bcl-XL. *Autophagy* 5: 720-722, 2009.
47. Pattingre S, Tassa A, Qu X, Garuti R, Liang XH, Mizushima N, Packer M, Schneider MD and Levine B: Bcl-2 antiapoptotic proteins inhibit Beclin 1-dependent autophagy. *Cell* 122: 927-939, 2005.
48. Itakura E, Kishi C, Inoue K and Mizushima N: Beclin 1 forms two distinct phosphatidylinositol 3-kinase complexes with mammalian Atg14 and UVRAG. *Mol Biol Cell* 19: 5360-5372, 2008.
49. Martinet W, Knaapen MW, Kockx MM and De Meyer GR: Autophagy in cardiovascular disease. *Trends Mol Med* 13: 482-491, 2007.
50. Furuya N, Yu J, Byfield M, Pattingre S and Levine B: The evolutionarily conserved domain of Beclin 1 is required for Vps34 binding, autophagy and tumor suppressor function. *Autophagy* 1: 46-52, 2005.
51. Valentim L, Laurence KM, Townsend PA, Carroll CJ, Soond S, Scarabelli TM, Knight RA, Latchman DS and Stephanou A: Urocortin inhibits Beclin1-mediated autophagic cell death in cardiac myocytes exposed to ischaemia/reperfusion injury. *J Mol Cell Cardiol* 40: 846-852, 2006.
52. Xie H, Liu Q, Qiao S, Jiang X and Wang C: Delayed cardioprotection by sevoflurane preconditioning: A novel mechanism via inhibiting Beclin 1-mediated autophagic cell death in cardiac myocytes exposed to hypoxia/reoxygenation injury. *Int J Clin Exp Pathol* 8: 217-226, 2015.
53. Peng W, Liu Y, Xu WJ and Xia QH: Role of Beclin 1-dependent autophagy in cardioprotection of ischemic preconditioning. *J Huazhong Univ Sci Technol Med Sci* 33: 51-56, 2013.
54. Levine B, Sinha S and Kroemer G: Bcl-2 family members: Dual regulators of apoptosis and autophagy. *Autophagy* 4: 600-606, 2008.
55. Shimizu S, Kanaseki T, Mizushima N, Mizuta T, Arakawa-Kobayashi S, Thompson CB and Tsujimoto Y: Role of Bcl-2 family proteins in a non-apoptotic programmed cell death dependent on autophagy genes. *Nat Cell Biol* 6: 1221-1228, 2004.
56. Brocheriou V, Hagege AA, Oubenaissa A, Lambert M, Mallet VO, Duriez M, Wassef M, Kahn A, Menasché P and Gilgenkrantz H: Cardiac functional improvement by a human Bcl-2 transgene in a mouse model of ischemia/reperfusion injury. *J Gene Med* 2: 326-333, 2000.
57. Imahashi K, Schneider MD, Steenbergen C and Murphy E: Transgenic expression of Bcl-2 modulates energy metabolism, prevents cytosolic acidification during ischemia and reduces ischemia/reperfusion injury. *Circ Res* 95: 734-741, 2004.
58. Liang XH, Kleeman LK, Jiang HH, Gordon G, Goldman JE, Berry G, Herman B and Levine B: Protection against fatal Sindbis virus encephalitis by beclin, a novel Bcl-2-interacting protein. *J Virol* 72: 8586-8596, 1998.
59. Ke J, Yao B, Li T, Cui S and Ding H: A2 Adenosine receptor-mediated cardioprotection against reperfusion injury in rat hearts is associated with autophagy downregulation. *J Cardiovasc Pharmacol* 66: 25-34, 2015.
60. Maiuri MC, Le Toumelin G, Criollo A, Rain JC, Gautier F, Juin P, Tasdemir E, Pierron G, Troulinaki K, Tavernarakis N, *et al*: Functional and physical interaction between Bcl-X(L) and a BH3-like domain in Beclin-1. *EMBO J* 26: 2527-2539, 2007.
61. Weston CR and Davis RJ: The JNK signal transduction pathway. *Curr Opin Cell Biol* 19: 142-149, 2007.
62. Zhao Y and Herdegen T: Cerebral ischemia provokes a profound exchange of activated JNK isoforms in brain mitochondria. *Mol Cell Neurosci* 41: 186-195, 2009.
63. Xu J, Qin X, Cai X, Yang L, Xing Y, Li J, Zhang L, Tang Y, Liu J, Zhang X and Gao F: Mitochondrial JNK activation triggers autophagy and apoptosis and aggravates myocardial injury following ischemia/reperfusion. *Biochim Biophys Acta* 1852: 262-270, 2015.
64. Madesh M, Antonsson B, Srinivasula SM, Alnemri ES and Hajnoczky G: Rapid kinetics of tBid-induced cytochrome c and Smac/DIABLO release and mitochondrial depolarization. *J Biol Chem* 277: 5651-5659, 2002.
65. Yang J, Liu X, Bhalla K, Kim CN, Ibrado AM, Cai J, Peng TI, Jones DP and Wang X: Prevention of apoptosis by Bcl-2: Release of cytochrome c from mitochondria blocked. *Science* 275: 1129-1132, 1997.
66. Chu G, Egnaczyk GF, Zhao W, Jo SH, Fan GC, Maggio JE, Xiao RP and Kranias EG: Phosphoproteome analysis of cardiomyocytes subjected to beta-adrenergic stimulation: Identification and characterization of a cardiac heat shock protein p20. *Circ Res* 94: 184-193, 2004.
67. Gurusamy N, Lekli I, Gorbunov NV, Gherghiceanu M, Popescu LM and Das DK: Cardioprotection by adaptation to ischaemia augments autophagy in association with BAG-1 protein. *J Cell Mol Med* 13: 373-387, 2009.
68. Overbye A, Fengsrud M and Seglen PO: Proteomic analysis of membrane-associated proteins from rat liver autophagosomes. *Autophagy* 3: 300-322, 2007.
69. Townsend PA, Stephanou A, Packham G and Latchman DS: BAG-1: A multi-functional pro-survival molecule. *Int J Biochem Cell Biol* 37: 251-259, 2005.
70. Petiot A, Ogier-Denis E, Blommaert EF, Meijer AJ and Codogno P: Distinct classes of phosphatidylinositol 3'-kinases are involved in signaling pathways that control macroautophagy in HT-29 cells. *J Biol Chem* 275: 992-998, 2000.
71. Gutierrez MG, Master SS, Singh SB, Taylor GA, Colombo MI and Deretic V: Autophagy is a defense mechanism inhibiting BCG and Mycobacterium tuberculosis survival in infected macrophages. *Cell* 119: 753-766, 2004.
72. Zheng Y, Gu S, Li X, Tan J, Liu S, Jiang Y, Zhang C, Gao L and Yang HT: Berbamine postconditioning protects the heart from ischemia/reperfusion injury through modulation of autophagy. *Cell Death Dis* 8: e2577, 2017.
73. Herzig S and Shaw RJ: AMPK: Guardian of metabolism and mitochondrial homeostasis. *Nat Rev Mol Cell Biol* 19: 121-135, 2018.
74. Hardie DG and Sakamoto K: AMPK: A key sensor of fuel and energy status in skeletal muscle. *Physiology (Bethesda)* 21: 48-60, 2006.
75. Meley D, Bauvy C, Houben-Weerts JH, Dubbelhuis PF, Helmond MT, Codogno P and Meijer AJ: AMP-activated protein kinase and the regulation of autophagic proteolysis. *J Biol Chem* 281: 34870-34879, 2006.
76. Samari HR and Seglen PO: Inhibition of hepatocytic autophagy by adenosine, aminoimidazole-4-carboxamide riboside and N6-mercaptopurine riboside. Evidence for involvement of amp-activated protein kinase. *J Biol Chem* 273: 23758-23763, 1998.
77. Rohailla S, Clarizia N, Sourour M, Sourour W, Gelber N, Wei C, Li J and Redington AN: Acute, delayed and chronic remote ischemic conditioning is associated with downregulation of mTOR and enhanced autophagy signaling. *PLoS One* 9: e111291, 2014.
78. Kandadi MR, Hu N and Ren J: ULK1 plays a critical role in AMPK-mediated myocardial autophagy and contractile dysfunction following acute alcohol challenge. *Curr Pharm Des* 19: 4874-4887, 2013.
79. Park CW, Hong SM, Kim ES, Kwon JH, Kim KT, Nam HG and Choi KY: BNIP3 is degraded by ULK1-dependent autophagy via MTORC1 and AMPK. *Autophagy* 9: 345-360, 2013.
80. Przyklenk K, Undyala VV, Wider J, Sala-Mercado JA, Gottlieb RA and Mentzer RM Jr: Acute induction of autophagy as a novel strategy for cardioprotection: Getting to the heart of the matter. *Autophagy* 7: 432-433, 2011.
81. Dunlop EA, Hunt DK, Acosta-Jaquez HA, Fingar DC and Tee AR: ULK1 inhibits mTORC1 signaling, promotes multi-site Raptor phosphorylation and hinders substrate binding. *Autophagy* 7: 737-747, 2011.
82. Kim J, Kundu M, Viollet B and Guan KL: AMPK and mTOR regulate autophagy through direct phosphorylation of Ulk1. *Nat Cell Biol* 13: 132-141, 2011.
83. Fiordaliso F, Li B, Latini R, Sonnenblick EH, Anversa P, Leri A and Kajstura J: Myocyte death in streptozotocin-induced diabetes in rats is angiotensin II-dependent. *Lab Invest* 80: 513-527, 2000.
84. Lekli I, Ray D, Mukherjee S, Gurusamy N, Ahsan MK, Juhasz B, Bak I, Tosaki A, Gherghiceanu M, Popescu LM and Das DK: Co-ordinated autophagy with resveratrol and gamma-tocotrienol confers synergistic cardioprotection. *J Cell Mol Med* 14: 2506-2518, 2010.
85. Li Y, Wang Y, Kim E, Beemiller P, Wang CY, Swanson J, You M and Guan KL: Bnip3 mediates the hypoxia-induced inhibition on mammalian target of rapamycin by interacting with Rheb. *J Biol Chem* 282: 35803-35813, 2007.

86. Hamacher-Brady A, Brady NR, Gottlieb RA and Gustafsson AB: Autophagy as a protective response to Bnip3-mediated apoptotic signaling in the heart. *Autophagy* 2: 307-309, 2006.
87. Xu W, Jiang H, Hu X and Fu W: Effects of high-mobility group box 1 on the expression of Beclin-1 and LC3 proteins following hypoxia and reoxygenation injury in rat cardiomyocytes. *Int J Clin Exp Med* 7: 5353-5357, 2014.
88. Ouyang F, Huang H, Zhang M, Chen M, Huang F and Zhou S: HMGB1 induces apoptosis and EMT in association with increased autophagy following H/R injury in cardiomyocytes. *Int J Mol Med* 37: 679-689, 2016.
89. Sciarretta S, Zhai P, Shao D, Maejima Y, Robbins J, Volpe M, Condorelli G and Sadoshima J: Rheb is a critical regulator of autophagy during myocardial ischemia: Pathophysiological implications in obesity and metabolic syndrome. *Circulation* 125: 1134-1146, 2012.
90. Shi WY, Xiao D, Wang L, Dong LH, Yan ZX, Shen ZX, Chen SJ, Chen Y and Zhao WL: Therapeutic metformin/AMPK activation blocked lymphoma cell growth via inhibition of mTOR pathway and induction of autophagy. *Cell Death Dis* 3: e275, 2012.
91. Gurusamy N, Lekli I, Mukherjee S, Ray D, Ahsan MK, Gherghiceanu M, Popescu LM and Das DK: Cardioprotection by resveratrol: A novel mechanism via autophagy involving the mTORC2 pathway. *Cardiovasc Res* 86: 103-112, 2010.
92. Zhao M, Sun L, Yu XJ, Miao Y, Liu JJ, Wang H, Ren J and Zang WJ: Acetylcholine mediates AMPK-dependent autophagic cytoprotection in H9c2 cells during hypoxia/reoxygenation injury. *Cell Physiol Biochem* 32: 601-613, 2013.
93. Xie H, Xu Q, Jia J, Ao G, Sun Y, Hu L, Alkayed NJ, Wang C and Cheng J: Hydrogen sulfide protects against myocardial ischemia and reperfusion injury by activating AMP-activated protein kinase to restore autophagic flux. *Biochem Biophys Res Commun* 458: 632-638, 2015.
94. Takagi H, Matsui Y, Hirotsu S, Sakoda H, Asano T and Sadoshima J: AMPK mediates autophagy during myocardial ischemia in vivo. *Autophagy* 3: 405-407, 2007.
95. Yang Y, Wang H, Wang S, Xu M, Liu M, Liao M, Frank JA, Adhikari S, Bower KA, Shi X, *et al*: GSK3 β signaling is involved in ultraviolet B-induced activation of autophagy in epidermal cells. *Int J Oncol* 41: 1782-1788, 2012.
96. Onishi A, Miyamae M, Kaneda K, Kotani J and Figueredo VM: Direct evidence for inhibition of mitochondrial permeability transition pore opening by sevoflurane preconditioning in cardiomyocytes: Comparison with cyclosporine A. *Eur J Pharmacol* 675: 40-46, 2012.
97. Shiomi M, Miyamae M, Takemura G, Kaneda K, Inamura Y, Onishi A, Koshinuma S, Momota Y, Minami T and Figueredo VM: Sevoflurane induces cardioprotection through reactive oxygen species-mediated upregulation of autophagy in isolated guinea pig hearts. *J Anesth* 28: 593-600, 2014.
98. Hariharan N, Zhai P and Sadoshima J: Oxidative stress stimulates autophagic flux during ischemia/reperfusion. *Antioxid Redox Signal* 14: 2179-2190, 2011.
99. Zhong Y, Zhong P, He S, Zhang Y, Tang L, Ling Y, Fu S, Tang Y, Yang P, Luo T, *et al*: Trimetazidine protects cardiomyocytes against hypoxia/reoxygenation injury by promoting AMP-activated protein kinase-dependent autophagic flux. *J Cardiovasc Pharmacol* 69: 389-397, 2017.
100. Li YY, Xiang Y, Zhang S, Wang Y, Yang J, Liu W and Xue FT: Thioredoxin-2 protects against oxygen-glucose deprivation/reperfusion injury by inhibiting autophagy and apoptosis in H9c2 cardiomyocytes. *Am J Transl Res* 9: 1471-1482, 2017.
101. Shiomi M, Miyamae M, Takemura G, Kaneda K, Inamura Y, Onishi A, Koshinuma S, Momota Y, Minami T and Figueredo VM: Induction of autophagy restores the loss of sevoflurane cardiac preconditioning seen with prolonged ischemic insult. *Eur J Pharmacol* 724: 58-66, 2014.
102. He C and Klionsky DJ: Regulation mechanisms and signaling pathways of autophagy. *Annu Rev Genet* 43: 67-93, 2009.
103. Wang LQ, Cheng XS, Huang CH, Huang B and Liang Q: Rapamycin protects cardiomyocytes against anoxia/reoxygenation injury by inducing autophagy through the PI3K/Akt pathway. *J Huazhong Univ Sci Technol Med Sci* 35: 10-15, 2015.
104. Zhai P, Sciarretta S, Galeotti J, Volpe M and Sadoshima J: Differential roles of GSK-3 β during myocardial ischemia and ischemia/reperfusion. *Circ Res* 109: 502-511, 2011.
105. Horn HF and Vousden KH: Coping with stress: Multiple ways to activate p53. *Oncogene* 26: 1306-1316, 2007.
106. Hoshino A, Matoba S, Iwai-Kanai E, Nakamura H, Kimata M, Nakaoka M, Katamura M, Okawa Y, Ariyoshi M, Mita Y, *et al*: p53-TIGAR axis attenuates mitophagy to exacerbate cardiac damage after ischemia. *J Mol Cell Cardiol* 52: 175-184, 2012.
107. Townsend PA, Scarabelli TM, Davidson SM, Knight RA, Latchman DS and Stephanou A: STAT-1 interacts with p53 to enhance DNA damage-induced apoptosis. *J Biol Chem* 279: 5811-5820, 2004.
108. Halestrap AP, Clarke SJ and Javadov SA: Mitochondrial permeability transition pore opening during myocardial reperfusion—a target for cardioprotection. *Cardiovasc Res* 61: 372-385, 2004.
109. Hausenloy D, Wynne A, Duchon M and Yellon D: Transient mitochondrial permeability transition pore opening mediates preconditioning-induced protection. *Circulation* 109: 1714-1717, 2004.
110. Saotome M, Katoh H, Yaguchi Y, Tanaka T, Urushida T, Satoh H and Hayashi H: Transient opening of mitochondrial permeability transition pore by reactive oxygen species protects myocardium from ischemia-reperfusion injury. *Am J Physiol Heart Circ Physiol* 296: H1125-H1132, 2009.
111. Nanda A, Gukovskaya A, Tseng J and Grinstein S: Activation of vacuolar-type proton pumps by protein kinase C. Role in neutrophil pH regulation. *J Biol Chem* 267: 22740-22746, 1992.
112. Nordstrom T, Grinstein S, Brisseau GF, Manolson MF and Rotstein OD: Protein kinase C activation accelerates proton extrusion by vacuolar-type H(+)-ATPases in murine peritoneal macrophages. *FEBS Lett* 350: 82-86, 1994.
113. Voss M, Vitavska O, Walz B, Wiczorek H and Baumann O: Stimulus-induced phosphorylation of vacuolar H (+)-ATPase by protein kinase A. *J Biol Chem* 282: 33735-33742, 2007.
114. Shen YT, Depre C, Yan L, Park JY, Tian B, Jain K, Chen L, Zhang Y, Kudej RK, Zhao X, *et al*: Repetitive ischemia by coronary stenosis induces a novel window of ischemic preconditioning. *Circulation* 118: 1961-1969, 2008.
115. Depre C, Park JY, Shen YT, Zhao X, Qiu H, Yan L, Tian B, Vatner SF and Vatner DE: Molecular mechanisms mediating preconditioning following chronic ischemia differ from those in classical second window. *Am J Physiol Heart Circ Physiol* 299: H752-H762, 2010.
116. Khan S, Salloum F, Das A, Xi L, Vetrovec GW and Kukreja RC: Rapamycin confers preconditioning-like protection against ischemia-reperfusion injury in isolated mouse heart and cardiomyocytes. *J Mol Cell Cardiol* 41: 256-264, 2006.
117. Marzetti E, Wohlgenuth SE, Anton SD, Bernabei R, Carter CS and Leeuwenburgh C: Cellular mechanisms of cardioprotection by calorie restriction: State of the science and future perspectives. *Clin Geriatr Med* 25: 715-732, 2009.
118. Kavazis AN, Alvarez S, Talbert E, Lee Y and Powers SK: Exercise training induces a cardioprotective phenotype and alterations in cardiac subsarcolemmal and intermyofibrillar mitochondrial proteins. *Am J Physiol Heart Circ Physiol* 297: H144-H152, 2009.
119. Jones SP and Bolli R: The ubiquitous role of nitric oxide in cardioprotection. *J Mol Cell Cardiol* 40: 16-23, 2006.
120. Ha T, Hua F, Liu X, Ma J, McMullen JR, Shioi T, Izumo S, Kelley J, Gao X, Browder W, *et al*: Lipopolysaccharide-induced myocardial protection against ischemia/reperfusion injury is mediated through a PI3K/Akt-dependent mechanism. *Cardiovasc Res* 78: 546-553, 2008.
121. Sala-Mercado JA, Wider J, Undyala VV, Jahania S, Yoo W, Mentzer RM Jr, Gottlieb RA and Przyklenk K: Profound cardioprotection with chloramphenicol succinate in the swine model of myocardial ischemia-reperfusion injury. *Circulation* 122: S179-S184, 2010.
122. Shirakabe A, Ikeda Y, Sciarretta S, Zablocki DK and Sadoshima J: Aging and autophagy in the heart. *Circ Res* 118: 1563-1576, 2016.
123. Gottlieb RA and Mentzer RM Jr: Cardioprotection through autophagy: Ready for clinical trial? *Autophagy* 7: 434-435, 2011.
124. Kanamori H, Takemura G, Goto K, Maruyama R, Tsujimoto A, Ogino A, Takeyama T, Kawaguchi T, Watanabe T, Fujiwara T, *et al*: The role of autophagy emerging in postinfarction cardiac remodeling. *Cardiovasc Res* 91: 330-339, 2011.