Expression of microRNA 638 and sex-determining region Y-box 2 in hepatocellular carcinoma: Association between clinicopathological features and prognosis

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Abstract. The aim of the present study was to determine the expression profile of microRNA 638 (miR-638) and sex-determining region Y-box 2 (SOX2) in hepatocellular carcinoma (HCC), and to investigate their association with clinicopathological features and survival. Reverse transcription-quantitative polymerase chain reaction analysis was used to investigate miR-638 and SOX2 expression in 78 patients with HCC. Western blot and immunohistochemical analyses were performed in order to determine SOX2 protein expression in HCC samples. Combined with the clinical postoperative follow-up data, the expression of miR-638 and SOX2 and the association between this and the prognostic values of patients with HCC were statistically analyzed. The results of the present study confirmed that miR-638 expression in tumor tissues was significantly downregulated (P<0.001), while SOX2 expression was significantly increased, compared with healthy control tissues (P<0.05). In addition, a significant inverse correlation between miR-638 and SOX2 expression was also observed in the HCC tissues (r=-0.675; P<0.05). Clinicopathological correlation analysis demonstrated that reduced miR-638 and elevated SOX2 expression was significantly associated with the Tumor-Node-Metastasis stage and portal vascular invasion (P<0.05). However, no significant differences were observed in other clinicopathological features, including age, sex, tumor size, tumor differentiation and hepatitis status (P>0.05). Notably, follow-up analysis revealed that patients with HCC with low miR-638 expression and high SOX2 expression tended to have a significantly shorter postoperative survival time (P<0.001). It was concluded that miR-638 may serve

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a vital role in the occurrence and progression of HCC by regulating SOX2 expression and thus, that miR-638 and SOX2 may be critical as novel diagnostic and prognostic biomarkers for HCC.

Introduction

Hepatocellular carcinoma (HCC) has one of the highest cancer-associated mortality rates worldwide, with a higher incidence observed more frequently in males than females (1). Although the prognosis for HCC has improved during the last two decades, HCC remains the sixth most common type of cancer globally and the second leading cause of cancer-associated mortality among males globally, with China alone accounting for ~50% of the total number of cases and mortalities (1). The traditional treatment method for HCC is surgery, including partial hepatectomy, transarterial chemoembolization and liver transplantation, followed by systemic postoperative chemotherapy; however, its clinical benefits remain uncertain (2-4). As the majority of patients with HCC are diagnosed at an advanced stage, with lymphatic or hematogenous metastasis to distal organs, curative surgical treatment at this time is no longer beneficial (5). Consequently, it is an ongoing effort to identify the metastatic behavior of HCC in clinical studies and the specific molecular biomarkers that may serve as potential diagnostic and prognostic indicators.

MicroRNAs (miRNAs/miRs) are a small class of endogenous non-coding RNAs (length, 19-22 nucleotides) that regulate the expression of protein-coding genes. In general, miRNAs suppress mRNA translation or degradation by binding to the 3'-untranslated region (3'-UTR) of target mRNAs (6). Numerous studies have demonstrated that miRNAs serve pivotal regulatory functions in cell cycle control (7), proliferation (8), differentiation (9), metastasis (10) and carcinogenesis (11). In human cancer, it has been observed that miRNAs function as tumor oncogenes or suppressor genes in the occurrence and progression of tumors (12,13). Dysregulation of miR-638 has been reported in several different types of cancer. For example, miR-638 has been demonstrated to be involved in colorectal carcinoma (14), gastric cancer (15), breast cancer (16) and osteosarcoma (17),

through dysregulation of its target genes as a tumor suppressor. Furthermore, miR-638 has also been demonstrated to promote melanoma progression and metastasis by suppressing tumor protein p53-mediated apoptosis pathways and autophagy as an oncogene (18). These inconsistent observations suggested that the function of miR-638 in tumorigenesis is cancer-specific.

The sex-determining region Y (SRY)-box 2 (SOX2) gene is a key transcriptional regulator associated with the maintenance of cell pluripotency and self-renewal in embryonic stem cells, encoding a member of the SRY-associated high mobility group (HMG)-box (SOX) family of transcription factors (19). Over time, accumulating evidence has established that the pro-oncogenic roles of SOX2 vary among different types of human malignant tumors, including breast (20), colorectal (21), prostate (22) and lung cancer (23). However, there has been contradictory evidence regarding SOX2 in certain types of tumor, with a number of studies suggesting that SOX2 may suppress tumors and that increased SOX2 expression inhibits cell proliferation and metastasis (24-26). In line with this, several studies have reported that miR-638 suppressed cell invasion, proliferation and epithelial-mesenchymal transition by downregulating SOX2 gene expression in non-small cell lung cancer (NSCLC) and colorectal carcinoma cells (27,28). As for HCC, there are limited studies that have investigated miR-638 and SOX2 expression and the clinical significance of this (29,30). Notably, whether or not an association exists between these two molecules remains to be elucidated.

The aim of the present study was to investigate miR-638 and SOX2 expression in human HCC tissues and their matched non-cancerous tissues, and to assess their association with clinicopathological features. The results of the present study demonstrated that miR-638 was downregulated and that SOX2 was upregulated in HCC, with a significant inverse correlation. Furthermore, reduced miR-638 expression and elevated SOX2 expression was associated with tumor stage, portal vascular invasion and poor postoperative survival. Consequently, they may be regarded as potential biomarkers for predicting HCC progression and prognosis.

Materials and methods

Clinical specimens and follow-up data. A total of 78 fresh cancerous tissues and their matched adjacent non-cancerous controls (≥3 cm from the tumor margin) were obtained from patients with HCC who had undergone routine curative surgical removal of the tumor at The First Affiliated Hospital of Wenzhou Medical University (Zhejiang, China) between January 2011 and July 2012. The histological diagnoses of the HCC specimens were independently confirmed by two senior pathologists at the First Affiliated Hospital of Wenzhou Medical University (Wenzhou, China), and the relevant clinicopathological information and follow-up data were retrieved from patient hospital records. Some of the surgically resected specimens were stored in liquid nitrogen at -80°C for further reverse transcription-quantitative polymerase chain reaction (RT-qPCR) or western blot analysis. The remaining specimens were fixed in a fixation solution containing 4% paraformaldehyde for 24 h at 4°C and were embedded in paraffin blocks for immunohistochemical analysis. Ethical approval was obtained from the Ethics Committee of The First Affiliated Hospital of Wenzhou Medical University, and written informed consent was obtained from all patients. Clinical specimens were collected from 67 males and 11 females aged between 30 and 83 years (mean, 57 years). Tumor size was classified according to the maximum size of the tumor detected by magnetic resonance imaging. Tumors were staged between I and IV, according to the latest edition of Tumor-Node-Metastasis (TNM) classification system of the American Joint Committee Cancer and Union for International Cancer Control (31). Tumor differentiation was assigned using the World Health Organization classification and grading system (32,33). The detailed clinicopathological data regarding the specimens are summarized in Table I.

RT-qPCR. Total RNA was extracted from tumor tissues and adjacent non-cancerous tissues using the E.Z.N.A miRNA kit (Omega Bio-Tek, Inc., Norcross, GA, USA), according to the manufacturer's protocol. miRNA expression analysis, synthesis of first strand cDNA and RT-qPCR were performed using the All-in-One™ miR RT-qPCR detection kit (GeneCopoeia, Inc., Rockville, MD, USA), according to the manufacturer's protocol. The reverse transcriptase reaction mixture (25 μ l), containing 2 µg total RNA, was applied by incubating mixtures at 37°C for 60 min, 85°C for 5 min, and 4°C for 30 min. The 20 µl PCR reaction mixture contained 1.2 µl RT product, 10 μl 2x All-in-One qPCR Master mix, 2 μl of each primer and 4.8 μ l ddH₂O. The primer sequences for miR-638 (cat. no. HmiR0295) and the reference gene small nuceloular RNA, C/D box 44 (RNU44; cat. no. HmiRQP9011) were designed and purchased from GeneCopoeia Inc. Following an initial denaturation step at 95°C for 10 min, the PCR samples were run for 40 cycles of 95°C for 10 sec, 60°C for 20 sec and 72°C for 10 sec. The relative expression of miR-638 was calculated using the $2^{-\Delta\Delta Cq}$ method (34), based upon the quantification cycle (Cq) method with RNU44 small nuclear RNA molecule as an endogenous reference.

For the measurement and quantification of SOX2 mRNA, cDNA was synthesized using the Hiscript® Q RT SuperMix for qPCR (Vazyme, Piscataway, NJ, USA). The PCR reaction (20 μ l) contained 2 μ l reverse transcriptase product, 10 μ l AceQ® qPCR SYBR®-Green Master mix (Vazyme), and 0.4 μ l of each primer. The PCR samples were subsequently incubated at 95°C for 3 min, followed by 49 cycles at 95°C for 15 sec, 60°C for 1 min and 50°C for 30 sec. Relative expression levels of SOX2 were calculated based on the $2^{-\Delta\Delta Cq}$ method and GAPDH was used as an internal control. All PCR analyses were performed using a CFX96TM Real-Time PCR detection system (Bio-Rad Laboratories, Inc., Hercules, CA, USA).

The primers for SOX2 and GAPDH were as follows: SOX2 forward, 5'-CGAGATAAACATGGCAATCAAAAT-3'; and reverse, 5'-AATTCAGCAAGAAGCCTCTCCTT-3'; GAPDH forward, 5'-TGCACCACCAACTGCTTAGC-3' and reverse, 5'-GGCATGGACTGTGGTCATGAG-3'. The primer sequences for miR-638 (cat. no. HmiR0295) and RNU44 (cat. no. HmiRQP9011) were purchased from GeneCopoeia Inc.

Immunohistochemistry (IHC) and assessment of IHC. Briefly, paraffin-embedded tissue blocks were sectioned (5 μ m thick) using a microtome, transferred onto tissue anti-off slides and heated at 60°C for 4 h. Tissue sections were dewaxed with

Table I. Association between miR-638 or SOX2 expression and clinicopathological characteristics in 78 patients with hepatocellular carcinoma.

| Variable | | Expression | n of miR-638 | Expression of SOX2 | | | |
|--------------------------|------|---------------|----------------|--------------------|-----------------|-----------------|-------------|
| | n=78 | Low (n=39) | High (n=39) | P-value | Negative (n=33) | Positive (n=45) | P-value |
| Age, years | | | | 0.820 | | | 0.710 |
| <57 | 35 | 17 | 18 | | 14 | 21 | |
| ≥57 | 43 | 22 | 21 | | 19 | 24 | |
| Sex | | | | 0.104 | | | 0.188 |
| Male | 67 | 36 | 31 | | 26 | 41 | |
| Female | 11 | 3 | 8 | | 7 | 4 | |
| Tumor size, cm | | | | 0.712 | | | 0.009^{b} |
| <5 | 51 | 23 | 28 | | 27 | 24 | |
| ≥5 | 27 | 11 | 16 | | 6 | 21 | |
| Tumor number | | | | 0.711 | | | 0.456 |
| 1 | 70 | 34 | 36 | | 31 | 39 | |
| ≥2 | 8 | 5 | 3 | | 2 | 6 | |
| Hepatitis B virus | | | | 0.411 | | | 0.224 |
| Positive | 61 | 29 | 32 | | 28 | 33 | |
| Negative | 17 | 10 | 7 | | 5 | 12 | |
| Cirrhosis | | | | 0.745 | | | 0.751 |
| Yes | 67 | 34 | 33 | | 29 | 38 | |
| No | 11 | 5 | 6 | | 4 | 7 | |
| AFP | | | | 0.784 | | | 0.915 |
| <400 ng/ml | 61 | 31 | 30 | 51, 51 | 26 | 35 | 0,010 |
| ≥400 ng/ml | 17 | 8 | 9 | | 7 | 10 | |
| Tumor differentiation | | | | 0.751 | | | 0.675 |
| Well | 31 | 16 | 15 | 57,51 | 15 | 16 | 0,0,0 |
| Moderately | 31 | 14 | 17 | | 12 | 19 | |
| Poorly | 16 | 9 | 7 | | 6 | 10 | |
| TNM stage | | | | $0.001^{\rm b}$ | | | 0.002^{b} |
| I-II | 48 | 17 | 31 | 0.001 | 27 | 21 | 0.002 |
| III-IV | 30 | 22 | 8 | | 6 | 24 | |
| Portal vascular invasion | | | | $0.005^{\rm b}$ | | | 0.012a |
| Yes | 21 | 16 | 5 | 31332 | 4 | 17 | 0.012 |
| No | 57 | 23 | 34 | | 29 | 28 | |
| Distant metastasis | | | | 0.156 | | | 0.071 |
| Yes | 9 | 7 | 2 | 0.100 | 1 | 8 | 3.071 |
| No | 69 | 32 | 37 | | 32 | 37 | |

^aP<0.05, ^bP<0.01. miR, microRNA; SOX2, sex-determining region Y-box 2; AFP, α-fetoprotein; TNM, Tumor-Node-Metastasis.

dimethylbenzene and rehydrated in a descending ethanol series (100, 95, 85 and 75%). Endogenous peroxidase activity was blocked with 0.3% hydrogen peroxide for 10 min at room temperature, and the sections were subsequently boiled by microwave in 0.01 mol/l citrate antigen retrieval solution (pH 6.0) for 15 min at 100°C. Following 3 washes with phosphate-buffered saline, the sections were blocked with 10% goat serum (cat. no C0265; Beyotime Institute of Biotechnology, Haimen, China) in phosphate-buffered

saline (PBS) for 15 min at 37°C, and incubated with a rabbit anti-human SOX2 polyconal antibody (dilution, 1:200; cat. no. ab97959; Abcam, Cambridge, UK) in a humidified chamber overnight at 4°C. The sections were subsequently incubated with a horseradish peroxidase (HRP)-labeled goat anti-rabbit secondary antibody (dilution, 1:100; cat. no. pv-6001; OriGene Technologies, Inc., Rockville, MD, USA) for 30 min at 37°C and washed with PBS to remove excess antibody. The sections were stained with 3,3'-Diaminobenzidine tetrahydrochloride

for 1 min at room temperature and observed using an optical microscope at a magnification, x40. Finally, the sections were counterstained with hematoxylin (OriGene Technologies, Inc.) for 3 min at room temperature, dehydrated, cleared, mounted and examined. In the present study, a PBS-only stained liver sample was used as a negative control, and human gastric cancer tissue was used as a positive control.

Semi-quantitative analysis. SOX2 expression in tumor tissues was semi-quantitatively evaluated and scored on the basis of the staining extent of positively stained cells, as described previously by Chen et al (24). Only nuclei stained brown were defined as SOX2-positive. For all samples, the staining intensity and percentage of positive tumor cells were evaluated and classified under double-blind conditions. Briefly, the staining intensities were scored as follows: 0, negative staining; 1, weak staining; 2, medium staining; or 3, strong staining. The staining percentage score (positively-stained cells/total tumor cells x100%) was defined as follows: 0, 0; 1, 1-9; 2, 10-29; 3, 30-49; or 4, 50-100%. Consequently, a final semi-quantitative immunoreactivity score (IRS) of SOX2 staining was obtained by multiplying the staining percentage score by the staining intensity score, resulting in scores ranging between 0 and 12. For statistical analyses, tumors with a final IRS of <5 were categorized as a low protein expression group and those with a final IRS of ≥6 were categorized as a high protein expression group.

Western blot analysis. The cancerous liver tissue samples and their adjacent non-cancerous tissue samples were homogenized using a radioimmunoprecipitation acid lysis buffer kit (Beyotime Institute of Biotechnology), and the lysates were subsequently centrifuged at 13,400 x g for 15 min at 4°C. The total protein concentration was measured by using the BCA method (Beyotime Institute of Biotechnology). Protein samples (40 μ g of each sample) were separated by 12% SDS-PAGE, prior to being transferred onto polyvinylidene difluoride membranes. Following blocking with 5% skimmed milk buffer for 120 min at room temperature, the membrane was incubated with anti-SOX2 (dilution, 1:2,000; cat. no. ab97957; Abcam, Cambridge, UK) and GAPDH (dilution, 1:2,000; cat. no. 5174; Cell Signaling Technology, Inc., Danvers, MA, USA) primary antibodies overnight at 4°C. Following three washes with 0.1% TBST for 5 min each time, the membranes were incubated with a HRP-conjugated goat anti-rabbit IgG secondary antibody (dilution, 1:5,000; cat. no 774P2; Cell Signaling Technology, Inc.) for 2 h at room temperature, prior to being washed again with 0.1% TBST 3 times. Finally, the protein complexes on the band were detected using a SuperSignal Western Femto kit (Pierce; Thermo Fisher Scientific, Inc., Waltham, MA, USA), and were subsequently quantified using the ChemiDoc XRS⁺ system with Image Lab™ 3.0 Software (cat. no. 1708265; Bio-Rad Laboratories, Inc.).

Statistical analysis. Data are presented as the mean \pm standard deviation, and were analyzed using SPSS 20.0 software (IBM Corp., Armonk, NY, USA) and GraphPad Prism 5.01 statistical software (GraphPad Software, Inc., La Jolla, CA, USA). The Pearson χ^2 and Fisher's exact tests were performed in order to test the significance of observed clinicopathological variables

among different groups. Student's t-test was performed to compare the miR-638 and SOX2 expression levels between different groups. Comparisons among multiple groups were performed using one-way analysis of variance and a Tukey's test. The association between miR-638 and SOX2 in the matched HCC tumor specimens was determined using Pearson's correlation analysis, and r-values represent the Pearson's correlation coefficient. The Kaplan-Meier method, followed by the log-rank test, and Cox proportional hazards regression model analysis were performed in order to plot survival curves and to calculate the hazard ratios (HRs) and the 95% confidence intervals (CIs), respectively. P<0.05 was considered to indicate a statistically significant difference.

Results

Patient characteristics. With regards to tumor size, 51 cases (65.4%) were classified as small (<5 cm in maximum diameter) and 27 cases (34.6%) were classified as large (≥5 cm in maximum diameter). With respect to tumor grades, 31 cases (39.7%) were categorized as well-differentiated, 31 cases (39.7%) as moderately-differentiated, and 16 cases (20.6%) as poorly-differentiated (Table I). In terms of clinical TNM stage, 48 cases (61.5%) exhibited TNM stage I-II disease and 30 cases (38.5%) exhibited TNM stage III-IV disease. With regards to tumor vascular invasion, 21 cases (26.9%) exhibited portal vascular invasion and 57 cases (73.1%) exhibited no vascular invasion (Table I). There were 61 cases (78.2%) of hepatitis B (HBV) positivity and 9 cases (11.5%) of distant metastasis. The overall survival period was defined as the time period between the date of surgical resection of HCC to the last follow-up or patient mortality. Mortalities caused by other events were considered to be censored (Table I).

Expression of miR-638 and SOX2 mRNA in human HCC. To assess miR-638 and SOX2 expression in HCC tumor tissues and paired adjacent normal tissues, RT-qPCR was performed. In the present study, it was demonstrated that miR-638 expression in tumor tissues was significantly lower than that in paired pericancerous healthy tissues (Fig. 1A; P<0.001). However, SOX2 mRNA expression was significantly upregulated (Fig. 1B; P<0.001). Furthermore, miR-638 expression in HCC tissues were negatively correlated with that of SOX2 mRNA (r=-0.675; Fig. 1C; P<0.001). The median expression of miR-638 and SOX2 were used as cut-off points to divide 78 samples into two groups. For statistical analysis, samples with miR-638 expression levels equal to or above the cut-off point were categorized as the high expression group (n=39) and samples with miR-638 expression levels below the cut-off point were categorized as the low expression group (n=39).

SOX2 protein expression is inversely associated with miR-638 expression. In order to study the association between miR-638 and SOX2 protein in HCC, SOX2 protein levels were measured in 78 pairs of tumor tissues and their adjacent non-cancerous tissues using western blot and immunohistochemical analyses. Western blot analysis demonstrated that SOX2 protein levels were significantly increased in 15 paired HCC samples, compared with adjacent non-cancerous samples (Fig. 2A; P<0.001). Immunohistochemical staining results revealed

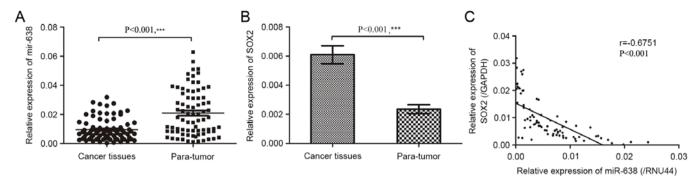


Figure 1. miR-638 is downregulated and SOX2 is overexpressed in patients with HCC. (A) miR-638 mRNA expression in 78 pairs of human HCC tissues and corresponding para-tumor tissues. (B) SOX2 mRNA expression in cancer tissues and para-tumor tissues was examined by reverse transcription-quantitative polymerase chain reaction (P<0.05). (C) An inverse correlation was observed between miR-638 and SOX2 mRNA expression levels in HCC tissues (r=-0.675; P<0.001). **P<0.005, ***P<0.001. miR, microRNA; SOX2, sex-determining region Y-box 2; HCC, hepatocellular carcinoma; para-tumor, paracancerous tissues.

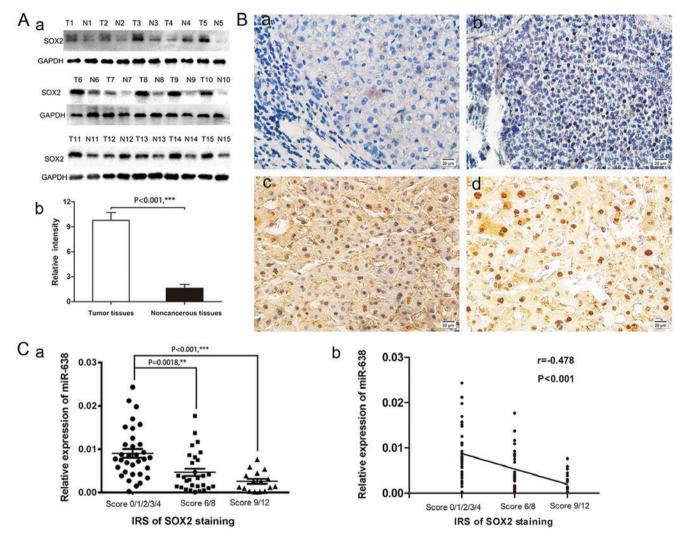


Figure 2. SOX2 was overexpressed in HCC tissues. (A) Expression of a) SOX2 protein in HCC tissues and matched adjacent non-cancerous tissues in 15 randomly selected patients with HCC was detected by b) western blot analysis. (B) Representative images of immunohistochemical staining of SOX2 in HCC tissues and adjacent non-cancerous tissues are presented in a-d. (Ba) No staining was detected for SOX2 in the blank control group; (Bb) weak nuclear staining of SOX2 in tumor cells; (Bc) medium nuclear staining of SOX2 in tumor cells; and (Bd) strong nuclear staining of SOX2 in tumor cells. (C) The a) semi-quantitative IRS of SOX2 staining was b) negatively correlated with the miR-638 levels in HCC tissues (one-way analysis of variance, P<0.05; Spearman's, r=-0.478; P<0.001).

P<0.05, *P<0.001.SOX2, sex-determining region Y-box 2; HCC, hepatocellular carcinoma; IRS, immunoreactivity score; miR, microRNA.

that SOX2-positive staining was only evident in the nuclei of tumor cells but not in peritumoral tissues. The representative images of SOX2 immunostaining in HCC tissues are presented in Fig. 2B. Of the 78 samples, 45 (57.7%) displayed high expression of SOX2 in tumor tissues compared with 21 (26.9%) paracancerous tissues samples exhibiting high

Table II. Expression of SOX2 in HCC tissues and paracancerous tissues.

| Group | | | Expression of | | | |
|-----------------------|-------|----------|---------------|-------------------|----------|---------|
| | Total | Negative | Positive | Positive rate (%) | χ^2 | P-value |
| HCC tissues | 78 | 33 | 45 | 57.7 | 15.127 | 0.000 |
| Paracancerous tissues | 78 | 57 | 21 | 26.9 | | |

SOX2, sex-determining region Y-box 2; HCC, hepatocellular carcinoma.

Table III. The association between miR-638 and the IRS of SOX2 staining expression in 78 pairs of hepatocellular carcinoma samples.

| | IRS | | | | | |
|--------------------|-----------|-----|------|-------|----------|---------|
| Group | 0/1/2/3/4 | 6/8 | 9/12 | Total | χ^2 | P-value |
| miR-638 expression | | | | | | |
| High | 8 | 19 | 12 | 39 | 15.551 | 0.000 |
| Low | 25 | 10 | 4 | 39 | | |
| Total | 33 | 29 | 16 | 78 | | |

miR, microRNA; IRS, immunoreactivity score; SOX2, sex-determining region Y-box 2.

expression (χ^2 =15.127, Table II; P<0.001). Furthermore, in order to statistically analyze the correlation between miR-638 and SOX2, a cross analysis was performed and is presented in Table III (χ^2 =15.551; P<0.0001), and the results revealed that the semi-quantitative immunoreactivity scores (IRS) of SOX2 staining in HCC tissues were negatively associated with miR-638 expression level (one-way analysis of variance, P<0.05; r=-0.478, P<0.001; Fig. 2C). The aforementioned results indicated that increased SOX2 expression in HCC may be due to miR-638 underexpression, suggesting a potential functional link between these two molecules.

Correlation between miR-638 or SOX2 expression and clinicopathological features. To observe whether miR-638 and SOX2 expression levels were correlated with clinicopathological features of HCC, clinicopathological analysis was performed (Table I). The data indicated that miR-638 and SOX2 were not significantly correlated with age, sex, tumor size, tumor number, serum a-fetoprotein (AFP) level, tumor differentiation (all P>0.05). By contrast, a significant association was observed between miR-638 or SOX2 expression and TNM staging (P=0.001 and 0.002, respectively). It was also demonstrated that patients with HCC with lower miR-638 expression and higher SOX2 expression were more likely to be at a higher portal vascular rate (P=0.005 and 0.012, respectively). The results from the present study suggested that miR-638 and SOX2 expression may serve a vital role in HCC progression and may have potential as novel prognosis biomarkers for HCC.

Association between miR-638 or SOX2 expression and the prognosis of patients with HCC. In order to further evaluate

whether there was an association between miR-638 or SOX2 expression and the overall prognosis of patients with HCC, the Kaplan-Meier method and Cox regression analysis were performed. Kaplan-Meier survival analysis suggested that miR-638 expression was significantly associated with a higher overall survival rate [median survival time (determined from the date of operation to the last visit or death), 50.20 vs. 19.10 months; Fig. 3A; P<0.001]. Consistently, the overall survival rate of patients with exhibiting SOX2-positive expression was significantly lower than in those with SOX2-negative expression (median survival time, 19.80 vs. 50.75 months; Fig. 3B; P<0.001). Furthermore, Univariate Cox proportional hazards regression model analysis demonstrated that tumor size, tumor number, HBV status, TNM stage, portal vascular invasion, distant metastasis, and miR-638 and SOX2 expression were statistically significant risk factors for the overall survival of patients with HCC (Table IV). However, age, sex, cirrhosis, serum AFP level and tumor differentiation had no prognostic value. Multivariate analysis, stratified for the known prognostic variables, revealed that portal vascular invasion (P=0.029), expression of SOX2 (P=0.013) and expression of miR-638 (P=0.008) were independent prognostic factors of HCC. These observations suggested that miR-638 and SOX2 may serve as novel prognostic biomarkers for patients diagnosed with HCC.

Discussion

HCC remains one of the leading causes of cancer-associated mortality worldwide (1). Due to the fact that there are no effective methods for prevention or early diagnosis, the lack of effective treatment strategies means that patients

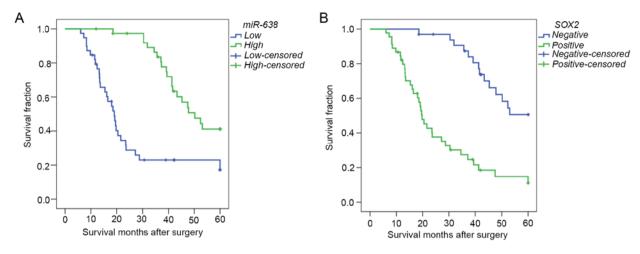


Figure 3. Kaplan-Meier curves for overall survival in patients with HCC. (A) Low miR-638 expression vs. high miR-638 expression and (B) negative SOX2 expression vs. positive SOX2 expression. HCC, hepatocellular carcinoma; miR, microRNA; SOX2, sex-determining region Y-box 2.

diagnosed with HCC exhibit a high mortality rate and a poor prognosis (2-5). In view of this, identifying sensitive and specific biomarkers for early diagnosis, therapy guidance and predicting prognosis is imperative. In the present study, the data indicated that miR-638 may negatively regulate SOX2 expression, and may be associated with advanced TNM stages and portal vascular invasion. The results provided evidence for the regulatory roles of miR-638 and SOX2 in HCC progression, and suggested that miR-638 and SOX2 may be potential biomarkers for predicting the prognosis of patients with HCC.

Dysregulation of miR-638 has been described to be involved in tumorigenesis and tumor progression in various types of human cancer, by targeting specific genes. Notably, the roles of miR-638 in different tumor types remains controversial as it is able to behave either as a tumor suppressive gene or an oncogene. For instance, Tan et al (16) demonstrated that expression of miR-638 reduced cell proliferation and decreased triple-negative breast cancer cell invasion, which in turn contributed to esophageal squamous carcinoma proliferation in vivo (35). Zhao et al (36) also identified that miRNA-638 acts as an anti-oncogene in human gastric cancer cells. By contrast, it has been observed that miR-638 promotes tumorigenesis and tumor development in human colon carcinoma cells and human osteosarcoma cells (37), and contributes to DNA damage in the benzo (a) pyrene-induced human cell transformation (38). Although inconsistent findings regarding miR-638 have been observed in different types of tumors, the role of miR-638 in hepatocarcinogenesis has not been clearly elucidated. Only one previous report demonstrated that underexpression of miRNA-638 promoted the angiogenesis and proliferation of HCC cells by targeting vascular endothelial growth factor (29), indicating that downregulation of miR-638 may serve a vital role in HCC progression. In agreement with this observation, the present study also confirmed that miR-638 expression levels were downregulated in a large number of HCC clinical samples. In addition, it was revealed that the reduced miR-638 expression was negatively correlated with advanced TNM stages and portal vascular invasion. Notably, it was confirmed that patients with tumors with a low expression of miR-638 were significantly more likely to exhibit a poorer overall survival. In line with this, a Cox proportional hazards model, adjusted for the possible prognostic variables, revealed that miR-638 may serve as an independent and favorable prognostic factor for HCC.

SOX2 is a highly conserved transcriptional regulator, which contributes an important role to the maintenance of embryonic stem cell pluripotency and self-renewal (19). It is well documented that SOX2 is expressed in various tissues and serves an important function in the differentiation and morphogenesis of the esophagus and stomach epithelial (39,40). The roles of SOX2 in these biological processes implicates that it has the potential to modulate the progression of cellular malignant transformation (41). Therefore, numerous studies have demonstrated that SOX2 was aberrantly expressed in a wide variety of solid malignant tumors, including breast, colorectal, prostate and lung cancer (20-23), indicating that SOX2 may act as a key factor in tumorigenesis and tumor development. In addition, previous studies have demonstrated that the overexpression of SOX2 in tumor tissues was associated with a strong invasiveness and a poor prognosis (42,43). Furthermore, SOX2 exhibits a close interaction with numerous miRNAs and, in previous studies, SOX2 activity was attributable to regulation by several miRNAs, including miR-126, miR-429 and miR-625 (44-46). In 2014, Ma et al (27) demonstrated a functional association between miR-638 and SOX2. Their study identified that downregulation of miR-638 promotes colorectal carcinoma cell (CRC) invasion and proliferation by influencing SOX2 expression. It was further demonstrated that miR-638 expression levels were negatively correlated with SOX2 expression, and the invasive and differentiative potential of CRC cells. In the same year, Xia et al (28) reported that the expression levels of miR-638 and SOX2 were inversely associated in non-small-cell lung cancer (NSCLC) tissues. The expression levels of miR-638 in the highly aggressive NSCLC cells were much lower than those in normal lung tissue cells. It was also observed that low miR-638 and high SOX2 expression in NSCLC tissues was significantly associated with tumor size, TNM stage and distant metastasis. Gain and loss of function experiments revealed that miR-638 downregulated SOX2 protein expression in NSCLC cells and inhibited cell invasive potential. Additionally, the regulation of SOX2 by

Table IV. Cox regression analysis of overall survival for patients with hepatocellular carcinoma (n=78).

| | | Univariate analysi | Multivariate analysis | | | |
|---------------------------------------|-------|--------------------|-----------------------|-------|-------------|-------------|
| Variable | HR | (95% CI) | P-value | HR | (95% CI) | P-value |
| Age, years (≥57 vs. <57) | 1.018 | 0.990-1.046 | 0.216 | | | |
| Sex (female vs. male) | 0.770 | 0.327-1.812 | 0.549 | | | |
| Tumor size, cm (≥5 vs. <5 cm) | 2.615 | 1.475-4.634 | $0.001^{\rm b}$ | 1.560 | 0.907-1.824 | 0.347 |
| Tumor number (≥2 vs. 1) | 2.257 | 1.047-4.865 | 0.038^{a} | | | |
| Hepatitis B virus | 0.471 | 0.240-0.926 | 0.029^{a} | | | |
| (positive vs. negative) | | | | | | |
| Cirrhosis (yes vs. no) | 0.813 | 0.365-1.811 | 0.612 | | | |
| AFP, U/l (≥400 vs. <400) | 1.581 | 0.822-3.040 | 0.170 | | | |
| Tumor differentiation | 1.407 | 0.975-2.030 | 0.068 | | | |
| (poorly vs. moderately vs. well) | | | | | | |
| TNM stage (III+IV vs. I+II) | 4.565 | 2.536-8.220 | <0.001° | 2.016 | 0.806-5.041 | 0.134 |
| Portal vascular invasion (yes vs. no) | 5.672 | 3.049-10.549 | <0.001° | 3.172 | 1.126-8.931 | 0.029^{a} |
| Distant metastasis (yes vs. no) | 4.030 | 1.893-8.579 | <0.001° | 1.505 | 0.589-3.846 | 0.394 |
| SOX2 (positive vs. negative) | 4.732 | 2.499-8.959 | <0.001° | 2.812 | 1.896-3.780 | 0.013^{a} |
| miR-638 expression (high vs. low) | 0.269 | 0.269-0.149 | <0.001° | 0.338 | 0.152-0.751 | 0.008^{b} |

^aP<0.05, ^bP<0.01, ^cP<0.001. HR, hazard ratio; CI, confidence interval; AFP, α-fetoprotein; TNM, Tumor-Node-Metastasis; miR, microRNA; SOX2, sex-determining region Y-box 2.

miR-638 may influence the epithelial-mesenchymal transition in NSCLC cells. An investigation into SOX2 expression and its clinical significance in HCC is therefore important for the management of the disease. The present study confirmed that SOX2 was overexpressed in liver cancer tissues and that SOX2 expression was positively correlated with tumor size, tumor stage and portal vascular invasion, as compared with negative controls. In addition, it was demonstrated that high SOX2 expression levels were associated with a poorer prognosis in patients with HCC, and served as an independent and unfavorable prognostic factor for HCC. The results of the present study supported the hypothesis that SOX2 is a key regulator in tumorigenesis and tumor development, which is in accordance with the results of previous studies on multiple types of cancer (42,43).

In the present retrospective study of patients with HCC, it was observed that miR-638 expression was markedly downregulated and SOX2 presented significantly higher expression in HCC tissue, compared with expression in adjacent non-cancerous controls. It was further validated that reduced miR-638 expression was negatively associated with overexpression of SOX2 protein in HCC. These observations were, in part, consistent with the conclusion of Zhang et al (30), that miR-638 may influence HCC progression by negatively regulating SOX2 expression. In addition, the present study demonstrated that miR-638 and SOX2 expression were significantly associated with tumor stage, portal vascular invasion and postoperative survival in patients with HCC. However, the failure to validate the molecular rationale for the involvement of miR-638 and SOX2 in the progression of HCC in vitro is one limitation of the present study. Another limitation is that the present study was retrospective and therefore, the results require further validation with more extensive tests in future prospective studies.

In summary, to the best of our knowledge, the present study was the first to provide evidence regarding the detailed expression pattern and clinical significance of miR-638 and SOX2 in a large number of patients with HCC. Furthermore, the results of the present study suggested that miR-638 may serve an important role in the occurrence and progression of HCC by downregulating SOX2 expression. Consequently, these proteins may serve as potential novel biomarkers, and may also be beneficial to the currently available HCC indicators for predicting HCC progression and poor prognosis. By identifying the patients who are more likely to have a higher risk of mortality, there is a possibility of implementing a more aggressive therapeutic regimen. Present and future studies regarding miR-638 and SOX2 expression in HCC progression may provide novel insights into the diagnosis and prognosis of this devastating disease.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

WYe, XC and JL carried out the experiments, and the data collection and interpretation. GF and YZ participated in the design and coordination of experimental work, and data acquisition. CZ and LC significantly contributed to analysis of data and preparation of the manuscript. WYa designed the study, analyzed and interpreted the data, and drafted the manuscript.

Ethics approval and consent to participate

Ethical approval was obtained from the Ethics Committee of The First Affiliated Hospital of Wenzhou Medical University (Zhejiang, China), and written informed consent was obtained from all patients.

Consent for publication

Written informed consent was obtained from all patients.

Competing interests

The authors declare that they have no competing interests.

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