A novel molecular probe $^{131}$I-K237 targeting tumor angiogenesis in human prostate cancer xenografts

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Abstract. Specific molecular probes are essential for the early diagnosis of prostate cancer. In addition, peptides have been shown to have numerous uses as diagnostic and therapeutic molecular probes. The K237 peptide binds to the vascular endothelial growth factor receptor with high affinity and specificity, and was predicted to have potential use as a probe in tumor angiogenesis. The overall aim of the present study was to assess the diagnostic potential of $^{131}$I-K237 as a molecular probe for prostate cancer. The K237 peptide was radiolabeled with $^{131}$I using an iodogen method. The radiolabeling efficiency and radiochemical purity were found to be 73.7±3.2 and 96.7±0.6%, respectively, which were determined using thin layer chromatography and high performance liquid chromatography in vitro. Cellular uptake and competition binding experiments were used to identify the affinity of $^{131}$I-K237 to LNCaP prostate cancer cells. The binding ratio of $^{131}$I-K237 to LNCaP cells in the experimental group was 95.8±1.5%, whereas the binding ratios in the 5 kBq Na$^{131}$I, 10 kBq Na$^{131}$I, 15 kBq Na$^{131}$I and PBS groups were 8.2±0.4, 8.3±0.2, 8.5±0.2 and 0.0%, respectively. In addition, the binding ratio of $^{131}$I-K237 to LNCaP significantly decreased with the increased dose of unlabeled K237. A total of 40 male BALB/c mice with LNCaP xenografts were used for biodistribution and have a low risk of inducing immunogenic responses and rapid blood clearance (9).

Small peptides, which target tumor angiogenesis, may be labeled with radionuclide and used in the targeted diagnosis and therapy of PCa. Through targeted radiation, the formation of novel tumor vessels may be inhibited, which may therefore prevent further cancer progression. Hetian et al (10) reported that the K237 peptide, which was isolated from a phage-displayed peptide library, was able to bind to the vascular endothelial growth factor (VEGF) receptor kinase insert domain receptor/fetal liver kinase 1 (KDR) with high affinity and specificity (10).

However, there have been limited studies into diagnostic imaging and therapeutic experiments on prostate cancer with radioiodine-labeled ($^{131}$I)-K237.

In the present study, to the best of our knowledge, $^{131}$I-K237 was used in the LNCaP prostate cancer cell line, for the first time. In addition, SPECT/CT imaging was used to characterize biodistribution of $^{131}$I-K237 in prostate cancer xenografts in vivo. Therefore, the present study aimed to determine the
diagnostic potential of $^{131}$I-K237 as a molecular probe for prostate cancer.

**Materials and methods**

**Ethics statement.** All animal experiments were approved by Ningxia Medical University Animal Studies Committee (Ningxia, China), according to the Guidelines for the Care and Use of Research Animals (Ningxia Medical University). Mice were fed a standard diet and were housed in a standard specific pathogen-free environment, with free access to food and drinking water according to the guidelines. Mice were finally sacrificed by cervical dislocation under anesthesia (0.1 ml Chloral Hydrate; Yulong Co., Ltd., Qingdao, China) to limit suffering.

**Radiosynthesis of $^{131}$I-K237.** All chemicals used were of analytical grade and commercially available. The K237 peptide (H-His-Thr-Met-Tyr-Tyr-His-His-Tyr-Gln-His-His-Leu-OH) was synthesized through the solid-phase peptide synthesis (SPPS) method (11). K237 was radiolabeled with iodine-131 using the Iodogen method as described previously (12). The radiolabeling yields and radiochemical purity of $^{131}$I-K237 were determined using thin layer chromatography (TLC) analysis using silica gel GF$_254$ (Qingdao Jitai Silica Gel Desiccant, Qingdao, China) coated on glass plates. Following development and drying, the plates were cut into sections and the radioactivity of each section was counted using a $\gamma$-well counter (Wizard2 Automatic Gamma Counter; PerkinElmer, Waltham, MA, USA).

A total of 30 µl of 0.1% trifluoroacetic acid (TFA; mobile phase A) and 10 µl of $10^{-5}$ M diethylene trimine pentaacetic acid were added together with 2 µl labeled peptide solution. High performance liquid chromatography (HPLC) analysis involved a gradient elution performed on the Agilent HPLC system 1100 Series (Agilent Technologies, Santa Clara, CA, USA) equipped with a LichroCART 250-3 LiChrospher 100 RP-18 (5 µm; Merck KGaA, Darmstadt, Germany) with a UV monitor (LB506C; Bio-Rad Laboratories, Inc., Hercules, CA, USA) and a radioactivity monitoring analyzer (B3110; Shimadzu, Tokyo, Japan) in 0.1% TFA in water as a mobile phase A and 0.1% TFA in CH$_3$CN as phase B. The flow rate was 1 ml/min.

**Cell culture.** The human LNCaP prostate cancer cell line (No. CRL-1740; American Type Culture Collection, Manassas, VA, USA) was used in this study, which was provided by the Department of Urology, General Hospital of Ningxia Medical University. LNCaP cells were grown in RPMI-1640 medium (Sigma-Aldrich, St. Louis, MO, USA) supplemented with 10% fetal bovine serum (Sigma-Aldrich) and 1 mM glutamine (Sigma-Aldrich), and cultured in a humidified atmosphere with 95% air and 5% CO$_2$ at 37°C. The experiments were performed with cells in the logarithmic phase of growth. The cell growth status was monitored using inverted microscopy with phase contrast (CKX31; Olympus Corp., Tokyo, Japan).

$^{131}$I-K237 binding experiment. Cellular uptake and competition binding experiments were used to identify the affinity of $^{131}$I-K237 to LNCaP cells. Cells were separated into groups as follows: Experimental group, treated with $^{131}$I-K237; negative groups, treated with different doses of Na$^{131}$I; blocking groups, treated with $^{131}$I-K237 and various doses of K237; and the blank group, treated with phosphate-buffered saline (PBS). LNCaP cells were cultured in 96-well plates (5x10$^4$ per well). Cells were starved for 16 h in serum-free medium, then each group was co-cultured with different media as follows: Experimental group contains 15 kBq $^{131}$I-K237 per well, the negative groups were treated with 5, 10 and 15 kBq Na$^{131}$I, and the blank group was treated with PBS only. Blocking groups treated with various doses of unlabeled K237 (0, 1, 2, 4, 8 and 16 µg/µl) were also added into the wells. The medium (non-uptake $^{131}$I-K237) and cells (uptake $^{131}$I-K237) of each well were then harvested at 48 h, and radioactivity was measured using the $\gamma$-well counter along with a standard, which contained 15 kBq $^{131}$I. Treatment for each well was repeated three times. The cellular binding ratios were then calculated separately. The binding ability of $^{131}$I-K237 was represented by the ratio of mean uptake as follows: Uptake ratio=uptake counts/mean total counts (uptake + non-uptake counts) x 100.

**Biodistribution.** Male BALB/c nu/nu mice (18±2 g, three weeks old; Vital River Laboratory Animal Technology Co. Ltd, Beijing, China) were used in the present study. The mice were inoculated with 1x10$^7$ LNCaP cells in their right upper limb and the tumors were allowed to grow to ~0.7 cm diameter, as measured using calipers.

A total of 35 BALB/c mice with LNCaP xenografts were randomly divided into seven groups (n=5). $^{131}$I-K237 was purified and isolated using HPLC analysis, as described above. A dose of 2.96 MBq $^{131}$I-K237 was injected into each mouse via the lateral tail vein; all injections were successful with no leakage. The mice were then sacrificed by cervical dislocation at 30 min, 1, 2, 4, 8, 12 and 24 h following injection.

Mice were dissected and tissues of interest (blood, heart, liver, spleen, lung, kidneys, muscle and tumor) were weighed, and their radioactivity was measured using the $\gamma$-well counter (6H4/5), which was equipped with a NaI(Tl) crystal detector and coupled to a high gain photomultiplier for a maximum efficiency of 80%, along with a standard solution of the injection. Radioactivity results were recorded as the percentage injected activity per gram (%ID/g) of tissue corrected for background and decay.

**SPECT imaging.** A total of five BALB/c mice with LNCaP xenografts were used for SPECT; 5.55 MBq $^{131}$I-K237 was injected into each mouse via the lateral tail vein. SPECT imaging was performed using a Symbia SPECT/CT imaging system equipped with a high-energy, high-resolution, parallel-hole collimator (Siemens Healthcare, Erlangen, Germany) at 1, 2, 4 and 8 h following injection; these procedures were performed at the Department of Nuclear Medicine, General Hospital of Ningxia Medical University. Images were acquired at 200,000 counts with a zoom factor of 2.0, and were digitally stored in a 64x64 pixel matrix size.

**Statistical analysis.** SPSS 17.0 software (SPSS Inc., Chicago, IL, USA) was used for statistical analysis. Values are presented
as the mean ± standard deviation and a one-way analysis of variance was used to determine differences among groups. P<0.05 was considered to indicate a statistically significant difference.

Results

Radiosynthesis of $^{131}$I-K237. TLC was used to monitor the radiolabeling yields of $^{131}$I-K237. The radiolabeling yield was found to be 73.7±3.2% and the optimum radiolabeling conditions occurred when cells were treated with 100 µg K237 and 50 µg Iodogen with a reaction duration and temperature of 40 min and 37˚C, respectively. Radiochemical purities of >98.3% were obtained following purification. A representative example of HPLC analysis of the selected radiolabeled peptide is shown in Fig. 1.

$^{131}$I-K237 binding affinity for LNCaP cells. The binding ratio of $^{131}$I-K237 to LNCaP cells in the experimental group was 95.8±1.5%, whereas the binding ratio in the 5 kBq Na$^{131}$I, 10 kBq Na$^{131}$I, 15 kBq Na$^{131}$I and PBS groups were 8.2±0.4, 8.3±0.2, 8.5±0.2 and 0±0%, respectively. Conversely, the binding ratio of $^{131}$I-K237 to LNCaP cells significantly declined with the increased dose of unlabeled K237 (Fig. 2).

Biodistribution of $^{131}$I-K237 in mice bearing LNCaP xenografts. Biodistribution data are shown in Table I. The biodistribution of $^{131}$I-K237 was found to be characterized by rapid blood clearance, with 15.97 %ID/g remaining at 30 min following injection of 2.96 MBq $^{131}$I-K237, which

Table I. Biodistribution of $^{131}$I-K237 in mice bearing LNCaP xenografts.

<table>
<thead>
<tr>
<th>Organ/tissue</th>
<th>30 min</th>
<th>1 h</th>
<th>2 h</th>
<th>4 h</th>
<th>8 h</th>
<th>12 h</th>
<th>24 h</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blood</td>
<td>15.97±0.99</td>
<td>9.48±1.00</td>
<td>8.26±0.62</td>
<td>5.40±0.38</td>
<td>1.85±0.22</td>
<td>1.33±0.21</td>
<td>0.42±0.08</td>
</tr>
<tr>
<td>Heart</td>
<td>6.73±0.74</td>
<td>4.35±0.70</td>
<td>2.92±0.77</td>
<td>2.19±0.51</td>
<td>1.22±0.10</td>
<td>0.82±0.04</td>
<td>0.61±0.04</td>
</tr>
<tr>
<td>Spleen</td>
<td>4.67±0.37</td>
<td>3.74±0.27</td>
<td>3.39±0.34</td>
<td>2.33±0.23</td>
<td>1.36±0.21</td>
<td>0.78±0.02</td>
<td>0.54±0.02</td>
</tr>
<tr>
<td>Liver</td>
<td>7.15±0.60</td>
<td>5.61±0.80</td>
<td>5.36±0.91</td>
<td>3.13±0.79</td>
<td>1.20±0.16</td>
<td>0.79±0.03</td>
<td>0.56±0.04</td>
</tr>
<tr>
<td>Lung</td>
<td>5.90±0.61</td>
<td>4.26±0.43</td>
<td>2.89±0.50</td>
<td>1.61±0.33</td>
<td>1.16±0.20</td>
<td>0.68±0.06</td>
<td>0.27±0.02</td>
</tr>
<tr>
<td>Kidney</td>
<td>24.03±1.78</td>
<td>14.13±1.82</td>
<td>12.14±2.30</td>
<td>5.51±1.34</td>
<td>3.17±0.55</td>
<td>2.41±0.46</td>
<td>2.22±0.33</td>
</tr>
<tr>
<td>Skeletal muscle</td>
<td>2.52±0.30</td>
<td>1.53±0.24</td>
<td>1.27±0.25</td>
<td>0.94±0.15</td>
<td>0.58±0.06</td>
<td>0.33±0.03</td>
<td>0.12±0.01</td>
</tr>
<tr>
<td>Tumor</td>
<td>5.25±0.63</td>
<td>3.49±0.50</td>
<td>3.12±0.69</td>
<td>2.51±0.73</td>
<td>1.76±0.92</td>
<td>1.31±0.47</td>
<td>0.96±0.27</td>
</tr>
<tr>
<td>Tumor/muscle</td>
<td>2.08±0.15</td>
<td>2.28±0.10</td>
<td>2.45±0.13</td>
<td>2.68±0.18</td>
<td>3.04±0.26</td>
<td>3.97±0.26</td>
<td>8.01±0.33</td>
</tr>
</tbody>
</table>

Values are presented as the mean ± standard deviation (n=5) and were corrected for background radioactivity and decay. %ID/g, percentage of injected radioactivity per gram organ or tissue; $^{131}$I-K237, $^{131}$I-radiolabeled K237 peptide.

Figure 2. Binding ratio of $^{131}$I-K237 to LNCaP cells. Increasing doses of unlabeled K237 induced a significant decrease in the binding ratio of $^{131}$I-K237. Values are presented as the mean ± standard deviation (n=5; *P<0.05 compared with groups 1-4). 1-6, 0, 1, 2, 4, 8 and 16 µg/µl, respectively. $^{131}$I-K237, $^{131}$I-radiolabeled K237 peptide.
was reduced to 1.85 %ID/g by 8 h post injection. In addition, the uptake of $^{131}$I-K237 in the tumor remained at a relatively high level until 24 h following injection. As a result, the ratio of tumor-to-muscle (T/M) $^{131}$I-K237 accumulation markedly increased in a time-dependent manner. Notably, the T/M ratio reached 8.01±0.33 by 24 h post injection.

SPECT imaging of LNCaP xenografts in mice following treatment with $^{131}$I-K237. As shown in Fig. 3, nude mice bearing LNCaP xenografts were injected with 5.55 MBq $^{131}$I-K237 and SPECT imaging was performed at 1, 2, 4 and 8 h following injection. Tumors were observed at 2 h and were clearly visible by 4 h post injection of $^{131}$I-K237; this therefore indicated that the concentration of $^{131}$I-K237 in tumors gradually increased in a time-dependent manner.

**Discussion**

An increasing number of studies have started to focus on the targeted diagnosis and therapy of PCa (13-15) and it is essential to identify effective and specific targets involved in the formation and metastasis of PCa. It has been well-documented that tumor angiogenesis has a vital role in tumor growth and the initiation of metastasis (16-19). VEGF has been demonstrated to act as an endothelial cell-specific mitogen and has been confirmed to have a critical role in tumor angiogenesis (20,21). In addition, the binding of VEGF and its receptor KDR has been reported to initiate the downstream effect of neovascularization (22).

The K237 peptide (HTMYYHYQHHL), which was found to be involved in preventing the binding of VEGF to KDR, was isolated from the screening phage-displayed peptide library (10). K237 not only competed with VEGF in binding to KDR, but also specifically inhibited human endothelial cell proliferation in vitro (10). Therefore, the peptide K237 was shown to be an effective antagonist of VEGF. Furthermore, K237 was reported to inhibit tumor angiogenesis as well as reduce tumor growth and metastasis (23).

In the present study, K237 was radiolabeled ($^{131}$I-K237) and the potential applications of $^{131}$I-K237 SPECT imaging were investigated, which were found to target tumor angiogenesis in PCa xenografts in mice (23).

Radiolabeled molecules have important roles in elucidating tumor status and characteristics, including aggressiveness and angiogenesis, as well as roles in monitoring the effectiveness of cancer treatments, including chemotherapy and radiotherapy (24). In the present study, the radiolabeling yield of $^{131}$I-K237 was found to be 73.7±3.2%, as determined using the Idogen method, which was higher compared with that of a study by Wang et al (25).

The results of the present study showed that $^{131}$I-K237 had a high affinity for LNCaP cells, which had a binding ratio of 95.8±1.5%; this result was comparable with that of a study by Wei et al (26). In the present study, the binding ratio of Na$^{131}$I groups were <10%, which confirmed that the higher affinity of the experimental group did not occur due to the contribution of radiodiode-131. In the blocking groups, the affinity of $^{131}$I-K237 for LNCaP cells declined with the increased dose of unlabeled K237. This, therefore, suggested that unlabeled K237 was able to compete for binding sites with $^{131}$I-K237, and this showed the binding specificity of $^{131}$I-K237.

In the present study, the biodistribution data for nude mice bearing LNCaP xenografts indicated that there was a rapid tumor uptake of $^{131}$I-K237. In addition, rapid blood clearance was demonstrated, with >66% of the tracer cleared by 4 h post injection. By contrast, the concentration of $^{131}$I-K237 in tumors gradually increased in a relatively high level until 24 h following injection. As a result, the ratio of T/M remained at a relatively high level until 24 h following injection. As a result, the ratio of T/M reached 8.01±0.33 by 24 h post injection. Consequently, the high ratio of T/M reached 8.01±0.33 by 24 h post injection. Consequently, the high ratio of T/M was comparable with that in a previous study by Wang et al (25).

In conclusion, the results of the present study demonstrated that $^{131}$I-K237 has the potential for use as a probe for the detection of prostate cancer in vivo.
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References